

1 **Another biomineralising protostome with an *msp130* gene and conservation of *msp130* gene**  
2 **structure across Bilateria.**

3

4 Réka Szabó and David E.K. Ferrier.

5 Affiliation: The Scottish Oceans Institute, Gatty Marine Laboratory, University of St Andrews, East

6 Sands, St Andrews, Fife, KY16 8LB, UK.

7 e-mails: [rs386@st-andrews.ac.uk](mailto:rs386@st-andrews.ac.uk)

8 [dekf@st-andrews.ac.uk](mailto:dekf@st-andrews.ac.uk)

9

10 Corresponding author: David E.K. Ferrier

11 Tel.: +44 (0)1334 463480

12 e-mail: [dekf@st-andrews.ac.uk](mailto:dekf@st-andrews.ac.uk)

13

14

1 MSP130 proteins form a distinctive family of cell surface glycoproteins associated with mineralised  
2 skeletal elements in larval and adult echinoderms (Anstrom et al. 1987; Illies et al. 2002; Mann et al.  
3 2008). In a recent paper in *Evolution and Development*, Etensohn (2014) investigated the  
4 phylogenetic distribution of the MSP130 family, finding that it is only present in prokaryotes and a  
5 handful of disparate eukaryotic clades. Among animals, only deuterostomes and certain molluscs  
6 possessed *msp130* genes. Based on this patchy distribution, Etensohn proposed that animals  
7 acquired *msp130* genes by two events of horizontal gene transfer, once in the deuterostome stem  
8 and once in the molluscan lineage. He also hypothesised that these acquisitions were related to the  
9 evolution of biomineralisation in these clades. However, he recognised the need for more data from  
10 other animals, such as calcifying annelids. Here we report the presence of an *msp130* gene in such  
11 an animal (the keelworm, *Spirobranchus* (formerly *Pomatoceros*) *lamarcki*), thus providing the first  
12 evidence for protostome *msp130* genes outside the phylum Mollusca, enhancing Etensohn's  
13 hypothesised link between *msp130* genes and biomineralising animals. However, we question the  
14 hypothesis of multiple origins, based on comparisons of gene structure between deuterostomes and  
15 molluscs. Etensohn allows for the possibility that bilaterian *msp130* genes originated in a single HGT  
16 event followed by extensive losses. Our findings of conserved introns lend support to a single origin,  
17 either through vertical inheritance or via a single horizontal transfer from a eukaryotic source.

### 18 **Annelids possess MSP130**

19 We have obtained transcriptome data from intact and regenerating specimens of the calcified head  
20 appendage (operculum) of the serpulid annelid *S. lamarcki*. Two contigs from our combined  
21 assembly likely represent two halves of a single *msp130* family transcript. This transcript is  
22 reasonably abundant in both intact and regenerating opercula, but with more reads in the  
23 regenerating (pre-calcifying and strongly calcifying) datasets. The transcript appears to contain a  
24 complete open reading frame (ORF) encoding a 604-amino acid protein with an N-terminal signal  
25 peptide predicted by TargetP v1.1 (Emanuelsson et al. 2000) with high confidence. We successfully

1 amplified and cloned the 3' end of the transcript from cDNA derived from mixed-stage opercular  
2 regenerates. The PCR fragment is predicted to span two conserved intron locations (see next  
3 section). Thus, an *mSP130* gene is present in *S. lamarcki* and expressed in a structure with calcareous  
4 components.

5 Alignment of the *S. lamarcki* protein with selected eukaryotic MSP130 proteins and one of the  
6 partial sequences we cloned is shown in supplementary Figure S1. We used the full list of MSP130  
7 proteins provided by Etensohn (2014) to generate neighbour-joining and maximum likelihood trees  
8 (Fig. 1). In our analysis, the *S. lamarcki* transcript groups among other metazoan sequences, but  
9 support values at nodes separating protostome from deuterostome genes within Metazoa are not  
10 significant, making it difficult to establish whether protostome and deuterostome sequences form  
11 distinct clades. However, the tree features a well-supported branch uniting all metazoan sequences,  
12 consistent with a single origin for metazoan *mSP130* genes.

### 13 **Conserved gene structure across Bilateria**

14 To further address the question of multiple origins, we compared the exon-intron structures of  
15 *mSP130* genes in the chordate *Branchiostoma floridae*, the sea urchin *Strongylocentrotus purpuratus*  
16 and the mollusc *Lottia gigantea* using available whole genome sequences. Where possible, exon  
17 boundaries were also confirmed using publicly available EST data. We discovered several intron  
18 locations that appear conserved across most or all genes in these three bilaterians; alignments of  
19 these regions are shown in Figure 1B. While conservation between bilaterians and algae is more  
20 limited, at least one site is predicted to be in a highly conserved sequence motif in all examined  
21 genes, and is supported by EST evidence in both *Lottia* and the brown alga *Ectocarpus siliculosus*.

22 The high level of conservation in bilaterian intron positions makes convergent evolution an unlikely  
23 explanation. In our opinion, these data are most consistent with the presence of *mSP130* genes in  
24 the urbilaterian, with secondary gene loss in some lineages such as the annelids *Capitella teleta* and

1 *Helobdella robusta*, and several arthropods (as surveyed by Etensohn, 2014) for which whole  
2 genome sequences are available. Additionally, we did not find an *msp130* gene in the whole genome  
3 sequence of the biomineralising cnidarian, *Acropora digitifera*, in addition to the other supposedly  
4 non-biomineralising non-bilaterian taxa surveyed by Etensohn (2014). Thus, this cnidarian data does  
5 not resolve whether the *msp130* genes in bilaterians stem from a HGT event early in bilaterian  
6 evolution, before the origin of protostomes and deuterostomes, or instead the gene was present at  
7 the origin of animals and has been secondarily lost, not only from some bilaterian lineages, but also  
8 several non-bilaterian lineages. Given the preponderance of gene loss across animal evolution  
9 (Putnam et al. 2007; Takahashi et al. 2009; Maeso et al. 2012) such extensive secondary loss of  
10 *msp130* genes is perhaps not so surprising. Etensohn (2014) hypothesised that the acquisition of  
11 *msp130* genes was related to the acquisition of mineralised skeletons. While our findings about gene  
12 structure call for a revision of this hypothesis, it would be equally interesting to see whether the  
13 *retention* of this family is associated with biomineralisation (skeletal or non-skeletal), although the  
14 link between *msp130* and animal biomineralisation is clearly not absolute, given the absence of the  
15 gene from a biomineralising cnidarian.

16

17

1   References

- 2   Anstrom, J.A., Chin, J.E., Leaf, D.S., Parks, A.L., and Raff, R.A. 1987. Localization and expression of  
3   msp130, a primary mesenchyme lineage-specific cell surface protein in the sea urchin embryo.  
4   Development *101*, 255–265.
- 5   Emanuelsson, O., Nielsen, H., Brunak, S., and von Heijne, G. 2000. Predicting subcellular localization  
6   of proteins based on their N-terminal amino acid sequence. J Mol Biol *300*, 1005–1016.
- 7   Ettensohn, C.A. 2014. Horizontal transfer of the *msp130* gene supported the evolution of metazoan  
8   biomineralization. Evol Dev *16*, 139-148.
- 9   Guindon, S., Dufayard, J.-F., Lefort, V., Anisimova, M., Hordijk, W., and Gascuel, O. 2010. New  
10   algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of  
11   PhyML 3.0. Syst Biol *59*, 307–321.
- 12   Illies, M.R., Peeler, M.T., Dechtiaruk, A.M., and Ettensohn, C.A. 2002. Identification and  
13   developmental expression of new biomineralization proteins in the sea urchin *Strongylocentrotus*  
14   *purpuratus*. Dev Genes Evol *212*, 419–431.
- 15   Maeso, I., Roy, S.W., and Irimia, M. 2012. Widespread recurrent evolution of genomic features.  
16   Genome Biol Evol *4*, 486–500.
- 17   Mann, K., Poustka, A.J., and Mann, M. 2008. The sea urchin (*Strongylocentrotus purpuratus*) test and  
18   spine proteomes. Proteome Sci *6*, 22.
- 19   Putnam, N.H., Srivastava, M., Hellsten, U., Dirks, B., Chapman, J., Salamov, A., Terry, A., Shapiro, H.,  
20   Lindquist, E., Kapitonov, V.V., et al. 2007. Sea anemone genome reveals ancestral eumetazoan gene  
21   repertoire and genomic organization. Science *317*, 86–94.
- 22   Takahashi, T., McDougall, C., Troscianko, J., Chen, W.-C., Jayaraman-Nagarajan, A., Shimeld, S., and  
23   Ferrier, D.E.K. 2009. An EST screen from the annelid *Pomatoceros lamarckii* reveals patterns of gene  
24   loss and gain in animals. BMC Evol Biol *9*, 240.
- 25   Tamura, K., Stecher, G., Peterson, D., Filipowski, A., and Kumar, S. 2013. MEGA6: Molecular  
26   Evolutionary Genetics Analysis version 6.0. Mol Biol Evol mst197.
- 27   Tu, Q., Cameron, R.A., Worley, K.C., Gibbs, R.A., and Davidson, E.H. 2012. Gene structure in the sea  
28   urchin *Strongylocentrotus purpuratus* based on transcriptome analysis. Genome Res. *22*, 2079–  
29   2087.

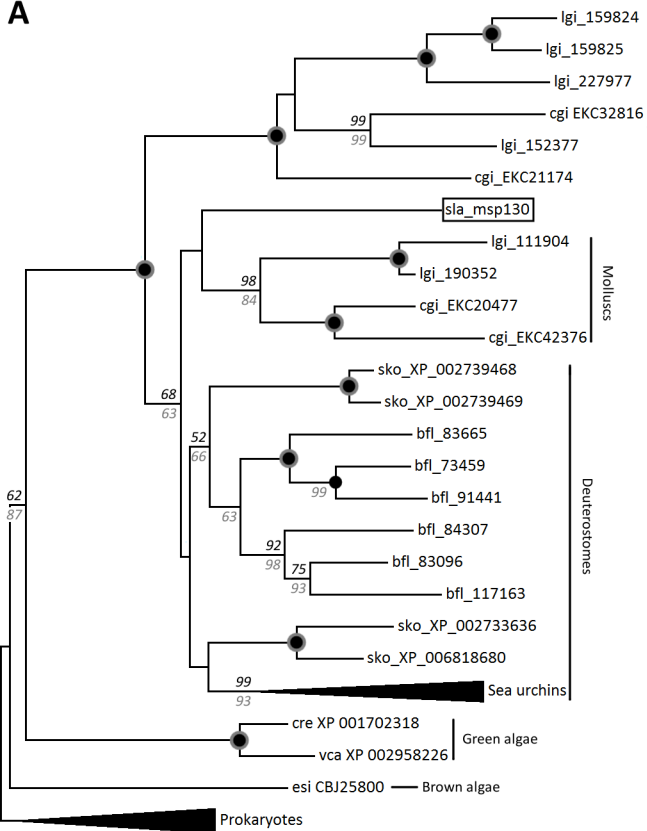
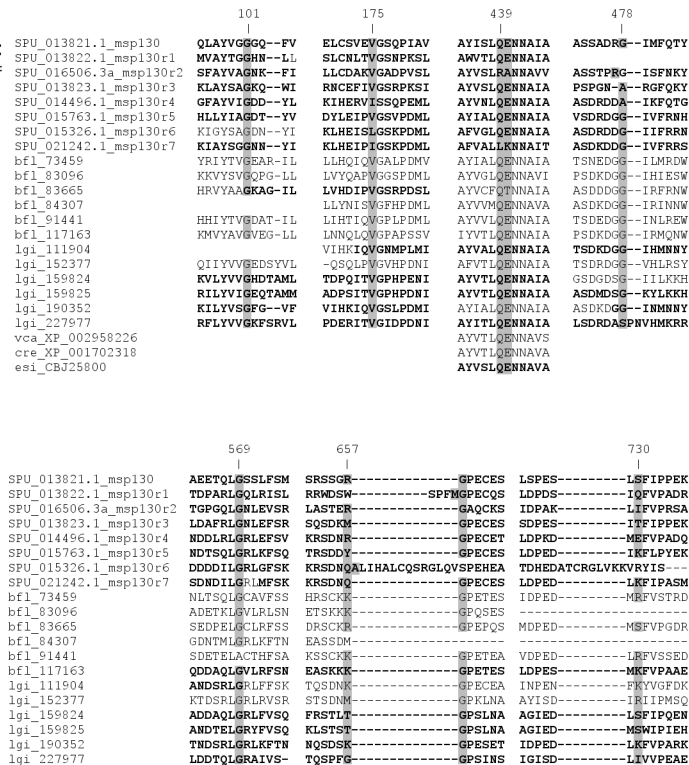
30

1 Figure legends

2 **Figure 1. A.** Maximum likelihood phylogenetic tree of available MSP130 protein sequences. The tree  
3 was generated in PhyML 3.0 (Guindon et al. 2010; <http://atgc.lirmm.fr/phyml/>) using the LG+G  
4 model with 500 bootstrap replicates. Support values over 50% are indicated; dots represent 100%  
5 support. Support values (percentage of 500 replicates) from a neighbour-joining tree of the same  
6 dataset generated in MEGA 6 (Tamura et al. 2013) are indicated in grey. Prokaryotic and echinoid  
7 sequences have been collapsed to reduce tree size. The *Spirobranchus lamarcki* sequence is  
8 highlighted with a box. **B.** Alignment of the residues surrounding conserved intron positions in  
9 bilaterian or eukaryotic MSP130 sequences. Sequences are based on gene models; bolded  
10 sequences have EST support or, in case of *Strongylocentrotus purpuratus*, mapped RNA-seq data (Tu  
11 et al. 2012). Highlighted residues indicate the position of the intron; numbers above the alignment  
12 indicate the position of the intron in *Strongylocentrotus purpuratus* MSP130. Algal sequences are  
13 only included where they show conservation with animals. *Spu-msp130r1* has been omitted from  
14 the alignment at position 478 because of a large insertion in this region. *Bfl\_84307* and *bfl\_83096*  
15 are incomplete. Species abbreviations: *bfl* *Branchiostoma floridae*, *cgi* *Crassostrea gigantea*, *cre*  
16 *Chlamydomonas reinhardtii*, *esi* *Ectocarpus siliculosus*, *lgi* *Lottia gigantea*, *sla* *Spirobranchus*  
17 *lamarcki*, *sko* *Saccoglossus kowalevskii*, *spu* *Strongylocentrotus purpuratus*, *vca* *Volvox carteri*.

18

19

**A****B**

0.2

## Supplementary information

### Methods summary

#### Identification of *sla\_msp130*

Three total RNA samples (derived from unoperated, 2-day and 6-day regenerating opercular filaments) were sequenced on the Illumina HiSeq2000 platform as 100-bp paired-end reads. The three sets of reads were pooled and assembled de novo using Trinity. Two *msp130* contigs were identified in the assembly using *spu-msp130* as a TBLASTN query; the four-codon overlap between these contigs was confirmed using the raw reads. To confirm the expression of this gene in the regenerating opercular filament, Primer3web was used to design primers to amplify the 3' end of the predicted transcript including two conserved splice junctions (see Figure S1) and part of the 3' UTR. A mixed-stage cDNA sample containing material from 8-hour and 1-4 day regenerates (kindly donated by Mr Tom Barton-Owen) was used as the template. PCR bands of the expected size were extracted and transformed into competent cells using the pGEM-T Easy® vector. Eight clones were sequenced, all of which were close matches (90-98% nucleotide level identity) to the contig sequence. Clone 5, which produced the closest match, was uploaded to Genbank under accession KM588349.

#### Alignment and phylogeny

Sequences used for phylogenetic reconstruction were based on the list provided in Ettensohn's (2014) supplementary information. Species, databases and accessions are given in Table S1, along with notes on corrections to automatic gene models. The *Acropora digitifera* genome (version 1.1, available at <http://marinegenomics.oist.jp/>) was also searched, but no good matches were found. Sequences were aligned with MAFFT and edited manually to remove poorly aligned regions and long repetitive stretches such as the polyG tracts in several sea urchin sequences. A neighbour-joining tree was constructed in MEGA 6 using the default settings, and a maximum likelihood tree was built in PhyML3.0 with the subtree pruning and regrafting search method, 500 bootstrap replicates, and the LG+G model selected by all three information criteria in Modelgenerator v0.85.

#### Gene structure in bilaterians and algae

Gene models were examined in six eukaryotes with sequenced genomes, representing the breadth of the distribution of *msp130* genes in this domain based on Ettensohn's (2014) findings. Species and genome databases were as follows: the deuterostomes *Strongylocentrotus purpuratus* (SpBase, version 3.1 assembly) and *Branchiostoma floridae* (JGI, Brafl1 and 2), the protostome *Lottia gigantea* (JGI, Lotgi1), the green algae *Chlamydomonas reinhardtii* (JGI, Chlre4) and *Volvox carteri* (Phytozome v10), and the brown alga *Ectocarpus siliculosus* (<http://bioinformatics.psb.ugent.be/genomes/view/Ectocarpus-siliculosus>). To confirm predicted splice junctions, the NCBI EST database was searched for matches to each predicted protein except those in the sea urchin, for which the RNA-seq data available through SpBase were used; in addition, ESTs were sought from the amphioxus cDNA resource (<http://amphioxus.icob.sinica.edu.tw/>) for *B. floridae*, and the aforementioned genome resource for *E. siliculosus*.



Figure S1. Alignment of sla\_msp130 with the best matching PCR clone derived from cDNA from a mixed sample of 8-hour and 1- to 4-day opercular regenerates, and selected bilaterian and algal sequences. The animal sequences are indicated by boxes. Columns are shaded according to the level of conservation, and intron positions where known are highlighted in red. Non-conserved N- and C-termini have been trimmed off; no other columns were removed. Species abbreviations: sla *Spirobranchus lamarcki*, spu *Strongylocentrotus purpuratus*, bla *Branchiostoma floridae*, lgi *Lottia gigantea*, vca *Volvox carteri*, esi *Ectocarpus siliculosus*.

10 20 30 40 50 60 70 80  
Animals  
sla\_msp130 YVLEHLNVTVVVFPAY----PDVYRYH-SVGEQIAINKDFPPYVYVIGNN---LI---HVIDISTVSNASLIYHKEVIAEDSGS--  
sla\_msp130\_clone5  
SPU\_016506.3a\_msp130r2 WILKWMATKXIPSDYINNGV-GEHKYD-QGAANSVAPDPASSFAYVAENK---FI---QVVDYSRFARPNIVRRQKTT---AP---  
bfl\_117163 IHLQPLSTVVLWNEF-NNDGTLDYDLD-KGAABCVAYHFQKQWVAVSEVGE--LL---TVIDVSVPTAARVHTQELPNAQMKK--  
lgi\_159824 VVLEBRGLQLQPD-----RNNFLRFDSTAGESAVDVQRKVLVYVGHDTA--ML---HVINNVNVDLSTLILSHNFPATEGK--

Algae  
vca\_XP\_002958226.1 ---MSSL-----KAGNWDGASMEVMDYDFPSKLAIVEA--RRSPIPLALLIVNYAVNSPFIH--PRILVGNSSSGGN  
esi\_CBJ25800.1 IGL-WLSPL-----S-----LTRGISSLPPPPSGQHVFAA--SATGV---SILKLGDDLSELWEL--GFYDSMTTFG--

100 110 120 130 140 150 160 170  
sla\_msp130 -LEDVITCG-----DWAVNIDSKD-SPLERVKVFTFR-----YNPSNDP--PMTLVDEIVG-----ALDSSIFTRDCSKLIV  
sla\_msp130\_clone5  
SPU\_016506.3a\_msp130r2 -AADIARCG-----FLVAVITRGV-KITDPGTVHWDM-----YRRVSKK---WLLCDAKVG-----ADVSLKRFHNCETIVI  
bfl\_117163 -YNDLALCG-----DSIAVIQSNPL-DALPSTLYIYGV-----Y-NGFSD--MALNNQLQVG-----PSSSVKFRSDCARLVV  
lgi\_159824 -PLDVEFCSTPTS--SVLAISSFSPLYEQAEHVILLYEH-----VSVGNEVVLVQKNPTDPQITVG-----PHEENIKETSDCALLIV

vca\_XP\_002958226.1 YVGTPTSVAVV-N--GFALLTMDGVP-YTASGILRIYHM-----ASAKVAEAPLTGCSMEDSVKMSKDGHRMVI  
esi\_CBJ25800.1 --EPTSVAYN-PVFDEVAISVRAFD-PLTPGRVYVVKSCEDWI--CDFCDDVQI-----LEAG-----FLEDMITPTPNKRIILT

190 200 210 220 230 240 250 260  
sla\_msp130 AIEGEPN--VGGQVV-----DEEGATIIDFGTNP--S-----SGSQSVT-F  
sla\_msp130\_clone5  
SPU\_016506.3a\_msp130r2 ANKTPAADAATNTFT-----DEEETISIVRIVKAQQQQQGGQPNYPGGAGAGTRFATTQGN---CAAQFGRHMIVT-T  
bfl\_117163 VNEGKAGLD-SFGNFK-----NPPETISVVDPIINSL-G-----GGTLASY-T  
lgi\_159824 SNEGIEGE--YDGKPV-----DEPSVSIISVTVAN--T-----DPT-----IST

vca\_XP\_002958226.1 ACEGEPN--T-----DEVQGSDDLVEPINSALAIAYVS-VSSYT-----PAGASWPFASFSISIK-L  
esi\_CBJ25800.1 ANEGEPN--NYVSAEN-----DEEVSISIFKRITK-----DNT-----YATAC-E

280 290 300 310 320 330 340 350  
sla\_msp130 VDETSF---NN--RKAEVVANGVFKSWNGEGANNIPST---TFSODLEPENIALNDDDES-L--VMTLOENNAVAVLNL--QQTQVW  
sla\_msp130\_clone5  
SPU\_016506.3a\_msp130r2 IDFRFF---NNTNRPFLNG--VMPYDGSMDGSRP-----TLKALRPYTVVGDPTYNDT--AVVSIKRNNAVQLILTPGNEAIT  
bfl\_117163 INELRF---DH--TQDEYTAGVWVYRGGEGTVGPG-----RMSDELEPEDFSFDETEN-K--IIVTLOENNAIAVNL--TTLIVD  
lgi\_159824 VSESEV---DTEPQRFLLNHVWMLRTEP-NTNIIIN---PFSNNIEPEYIITISPNNAY--AVVTLQENNAIAKIDI--ENGVYN

vca\_XP\_002958226.1 LDYCCYIDSLSN--SAYNALLARGFIDPR-----LTKATAAKDIRPEYVALHSDPQ-VNLAVTLOENNAVSAIDLTPGSEKIL  
esi\_CBJ25800.1 VGEKY---NTADRTNPLVDGGMRVGGM---NF---TTLSDVPEEYIAVTEFGT-T--AVVSIKRNNAVAKIDIKGCE--VK

370 380 390 400 410 420 430 440  
sla\_msp130 DIYALGTISFS-SGNEFGSKLDASDRDENIASY-PHGMYPQPSLKYKYSKGS-----SYLFMANEGDSREFTVDDIGMDEW--  
sla\_msp130\_clone5  
SPU\_016506.3a\_msp130r2 GFNPMKMTWF-N---NDLDASTPREGISFKNYN-MYSMRQFNAIESPEI---RNQYNLASYLETDEGAT-----  
bfl\_117163 EIYPLGANMA-N---YMLDPSDKDGGIRMQNW-PIFGLYQDDVVSFTVGSR---KLLATANEGNSRELTV---GGVD-I---  
lgi\_159824 QIYPLGVKWR-Y---STFGSDGDSSEIILKKH-NISSFYQPKIAFFEWKNR---LYLITADEGKEFSVP

vca\_XP\_002958226.1 SIWPLGLFSWK-A---SPVDPSDVG--IRIRNHTGVYSWYQPDIVHATIGTG-----SYLFMANEGDSK-----  
esi\_CBJ25800.1 RMYPLGFKKAGDD---MKFDASDEDGMNLQEWPTVRGMYMEDAITTFKTGGR---RFLITANEGRDREYGE-----EDT

460 470 480 490 500 510 520 530  
sla\_msp130 ---EEFSRGDDL---KDDKLSSS-VDPS--VVDMLND--DNCLGPIRFSLV-DG-----LDS--GKIETLHTFGGSIIVFR  
sla\_msp130\_clone5  
SPU\_016506.3a\_msp130r2 ---TSYRSKSKSF---TDAVPSR-LQMPNYISRNHTG--PGCLQMLEVSRV-DG-----LRNPQPNNSGYDYVSFFGGRGISMFR  
bfl\_117163 ---TDEWKGKDFI---TUGAVAPT-VPQT--LVAALQD--DAQLGVLRFSNY-DG-----KSASNP-GQYEQYAFGGGFSLWK  
lgi\_159824 IYFYTDFDRAKMLH---TUGEF---LVDP--LDAELAD--DAQLGPIRFSVQFHDG-----RATGS--SKINRVFTFGGGSFVFD

vca\_XP\_002958226.1 ---GESRRVKEL--AS---LDRHT-----TQDSELGRINVDPL-FGLKKGYNTRAWNAQGPYKLIAYGGRSWSILD  
esi\_CBJ25800.1 BEFFTDEIRVEDLATELG---LDAITMTP-----YTPBALGPIRVTATA-GP-----IDGSDIYELAFGGGSISIFD

550 560 570 580 590 600 610 620  
sla\_msp130 VSEDSVW-TAVHDTSELADBIAR---STPRVFNADSRKGD---ETPEETKESRSNDRGCEPIIEMAAI-GDTQLLFIAGIERPGLIAVY  
sla\_msp130\_clone5  
SPU\_016506.3a\_msp130r2 SNTSSNLDLVWDSGDQVARGVAE---TYPCVFNKSTTADAQSHGPKSQDLASTERACCKSLAVAKVDRDTLLEVVGADGFSVIGIF  
bfl\_117163 ASD---L-TQWDSGADVELQHTR---HLPLIFNCFPDANPN-KSPMDEKDEASKKGPETESLVVAV-YGKTVIIIGNERPSLLMIV  
lgi\_159824 ANN---M-IRQYESGDEIEKYSRL---PHNVFNQDC---SDANLAPFOEMDFRSTLTPGSLNIAVDGDF-LGRKLLIFSGTNGILYVY

vca\_XP\_002958226.1 AKNG---RLVYSESSMEAFIAAHFVAS-QCFNCDRDR-----ND-PDSRSDNAGPEPEALEVFCI-GSRTYAAIQLERMGGFLLY  
esi\_CBJ25800.1 GKTG---ELVWDSGDFIEQFTADPANGVSDIFNSNGEV---DTFDGRSDDKGPPEPEGLAMARI-DGSIYVVELERIGGWMCW

640 650 660 670 680 690 700 710  
sla\_msp130 TV-SMGDQDVTMEFCSIYCYGQYKTKWQCLYDERR-----THALDVEDLRFISAADSP--NKKDMIAAGSVSGTISFF  
sla\_msp130\_clone5  
SPU\_016506.3a\_msp130r2 SV---SQNGSYPAYESVYRKAPMDGHSILMDNKN-----MGDIDPAKLTIVPPSATP--DNYNYLVMGKKSGLSMY  
bfl\_117163 SV-DTVT--GIPSEESIFRAGDISKNYDAAVNDRN-----IGDLDEPESMGVPAESEP-DGTPLLLVVGNISGTVAVY  
lgi\_159824 ELVNPVSSRAEMEFQSVHRRGSLTDWGLYSSGI-----IGDAGIDLSRIPQEMSVFTGSPVLVSVSSKGSVSAY

vca\_XP\_002958226.1 DV---SVPSAPVFGAIYIN-----RNFSAPRTAPSTA-LGDIAPEGLIREDVAKDSS--SGTALILMSNE-----  
esi\_CBJ25800.1 DV---TDATAPVFCYVNS-----YEDTAPESATVISADMSR--SGNTLLVAAVEEINLAVF

Inferred nucleotide and protein sequence of sla\_msp130 based on two manually merged Trinity contigs

>comp389772\_merge

ATGTAACTATATGTACCGAATTCTGTCTAATTTTCAGTGGTTGAGAAGAGAGTGATAAAAAATAAGTGACACGC  
CATTCTCGAAAAAGTCATAAAATGTGCTTACGTATAAGTGACAGGAGACGCCGAAGTCATTCCACTTATTACA  
AATGGAAAAAGGATAATTAAGAAGAAACGTCAATGTAATTCAAACTGGAAAAAATCAGATTTAGAATTA  
TTAAATTGTTGAATAATATTCTAATTAATAAATCACACATAATCAACAGCAAATGGCTTTTAAGTGTATATTT  
CTATCGTAACCCTATTGATTTGTGTTGTTTCGTGGAAAGTATGTATTGGAGCATCTCAACACGGTCTATGTGCCT  
TTCGCATATCCGGATGTGTATAGATACCATAGCGGTGTTGGAGAACAAATTGCTATCAATAAGGATTTCCATA  
TGTGTATGTTATAGGAAATAATCTGATACACGTTATCGATATTTCAACTGTGTCAAACGCCTCTCTCATCTATCA  
TAAGGAAGTCATAGCTGAAGATAGTGGTAGCCTTGAGGATGTCATAACATGCGGGGACTGGGTAGCTGTTAA  
TATTGACAGCAAAGATCCCACTTGAAGGGAGGGTAAAGTATTCACGAGATACAACCCAAGCAATGACCCT  
CCCATGACAGAACTAGTGGATGAAATAGTTGGAGCCCTACCCGATAGTTTCGATATTCACACGGGACTGCAGCA  
AGCTGATCGTGGCCATAGAGGGAGAGCCGAGGAATGTGGGCGGCCAGGTCGTGGACCCGGAAGGGGGCGC  
CACAATCATCGACTTTGGTACAAATCCCTCCAGTGGTAGCCAATCAGTTACCTTTGTTGACTTCACCAGCTTAA  
TAACAGGAAGGCGGAATATGTGGCCAATGGTGTTCGCAAGAGTTGGAATGGTGAAGGAGCTAACAATATCC  
ATCAACAACATTTTCTCAAGATCTGGAGCCTGAAAATATTGCGCTGAATGATGATGAATCTTTGGTGTATATGA  
CATTGCAGGAGAATAATGCAGTTGCCGTTCTCAACCTGCAAACACAAACCTGGGTTGATATATACGCGCTGGG  
GACCAAGAGCTTCAGCAGTGGTAACGAATTCGGCAGCAAACCTAGACGCAAGCGACAGAGATGATGAAATCA  
ATATAGCAAGTTATCCTATCCATGGTATGTATCAACCGGATAGTCTGAAGTACAAGTCTACAAAGGGAGCAG  
TTATTTGTTTCATGGCCAACGAGGGAGACTCGCGAGAGTTCACCGTAGATGACATTGGCATGGACGAGGACTG  
GGAGGAGTTCAGCAGAGGAGATGACCTTGTGAAAGATGATAAGCTCAGTTCAAGTGTGGACCCCTCGGTGGT  
AGATATGCTAAACGATGATAACCAGCTAGGTGCGTTACGCTTCAGTCTAGTGGACGGATTGGACAGTGATGG  
CAAATAGAACTCTGCATACATTTGGTGGTTCGAGTATCGCTGTATTTAGAGTATCAGAGGACAGCGTGTGG  
ACGGCGGTACATGATACGGGCAGTGAGCTGGCGGACGAGATAGCTAGGTCCACACCCAGGGTCTTCAACGC  
AGACTCCAGGAAAGGAGATGAAACCCAGAGGAAACCAAAGAAAGTAGATCAGATAACCGGGGATGCGAAC  
CGGAAATAATAGAAATGGCTGCTATCGGTGACACACAACCTACTATTTGCGGGTATAGAGCGCCAGGCCACA  
TCGCCGTCTACACGGTTAGCATGGGCGACCAAGATGTAACCATGGAGTTTCAGAGTATATACTACTGTGGCCA  
GTACGGAAAAACATGGCAACAGCTTTATGACGAGCGCAGGACGCATGCTCTTGATGTGGAGGATTTAAGGTT  
TATTAGCGCAGCGGACAGCCCTAATAATAAGGACATGTTAATTGCCGCGGGCTCGGTTAGTGGCACAATCTCC  
TTCTCCAAGTAACCGATGATGGGAAATGCCCGTGGAAAATAATTCAGGTGAAGGAAGGCATCTGTCAGCG  
GTGCTGGGAGCTCTAATTGGAGCAATAACTTTATTGCTGGTGTCTGCATGAGGACAACCTCTCAATGGACCCA  
AAGAAATGAAGGCACCGCTCGAACTGCCCGATGAAACATCTGTTTCGATGGC

>comp389772\_merge\_ORF

MAFKCIFSIVTLICVVRGKYVLEHLNNTVYVPFAYPDVYRYHSGVGEQIAINKDFPYVYVIGNNLIHVIDISTVSNASLIY  
HKEVIAEDSGSLEDVITCGDWVAVNIDSKDSPLEGRVKVFRYFNPSNDPPMTELVDIVGALPDSSIFTRDCSKLIVAI  
EGEPRNVGGQVVDPEGGATIIDFGTNPSSGSQSVTFVDFTSFNNRKAIEYVANGVRKSWNGEGANNIPSTTFSQDL  
EPENIALNDDESLVYMTLQENNAVAVLNLTQQTWVDIYALGTKSFSSGNEFGSKLDASDRDDEINIASYPIHGMYYQ  
PDSLKYKSYKSSYLFMANEGDSREFTVDDIGMDEDWEEFSRGGDLVKDDKLSSSVDPVVDMLNDDNQLGRLR

FSLVDGLSDGKIETLHTFGGRSIAVFRVSEDSVWTAVHDTGSELADEIARSTPRVFNADSRKGDETPEETKESRSD  
 NRGCEPEIIEMAAIGDTQLLFAGIERPGHIAVYTVSMGDQDVTMEFQSIYYCGQYGKTWQQLYDERRTHALDVED  
 LRFISAADSPNNKDMLIAAGSVSGTISFFQVTDDGEMPVENNSGEGRHLSAVLALGALGAILLLVSA

Table S1. Sources of sequences used for alignment and phylogenetic reconstruction. Where the final sequence used differs from the one under the given accession, a brief note on modifications is given.

Higher taxon	Species	Database	Accession	Notes
Deuterostomes	Strongylocentrotus purpuratus	SpBase	SPU_013821.1	
		SpBase	SPU_013822.1	
		SpBase	SPU_016506.3a	
		SpBase	SPU_013823.1	
		SpBase	SPU_014496.1	
		SpBase	SPU_015763.1	
		SpBase	SPU_015326.1	Chosen as the more complete of Ettensohn's two accessions for msp130rel6.
		SpBase	SPU_021242.1	
	Helicidaris erythrogramma	Genbank	her_CAC20358.1	
	Helicidaris tuberculata	Genbank	htu_CAC20589.2	
	Eucidaris tribuloides	EchinoBase transcripts	etr_6853_2	
		EchinoBase transcripts	etr_38352	
		EchinoBase transcripts	etr_1749_2	
		EchinoBase transcripts	etr_21290	
		EchinoBase transcripts	etr_10262	
		EchinoBase transcripts	etr_22657_1	
	Saccoglossus kowalevskii	RefSeq	sko_XP_002739468.1	

		RefSeq, Metazome	sko_XP_002739469.1	A probably spurious region in the RefSeq sequence was replaced by the conserved exon beginning ENNAI based on the genomic sequence
		RefSeq	sko_XP_002733636.1	
		RefSeq, Metazome	sko_XP_006818680.1	This sequence was not listed by Ettensohn but appears distinct from the others. A missing conserved exon was reconstructed from genomic sequence.
	Branchiostoma floridae	JGI	73459	A conserved exon was added to the gene model from genomic sequence
		JGI	83096	
		JGI, NCBI EST	83665, FE585043.1	N-terminus of the gene model was replaced by sequence from the EST – JGI pipeline probably missed first exon
		JGI	84307	C-terminus probably spurious (poorly aligned and on far side of 17 kb sequencing gap) and thus removed from alignment
		JGI	91441	
		JGI	117163	
Protostomes	Lottia gigantea	JGI, NCBI EST	111904, FC586646 .1	The gene prediction seems to have missed the first exon and mispredicted portions of the protein due to a frame shift. Corrected based on alignment of the EST to the genome.
		JGI	152377	
		JGI	159824	
		JGI	159825	

		JGI, NCBI EST	190352, FC596551.1, FC761686.1	ESTs were used to extend the N-terminus
	<i>Crassostrea gigas</i>	Genbank	EKC20477.1	
			EKC42376.1	
			EKC21174.1	
			EKC32816.1	
	<i>Spirobranchus lamarcki</i>	own	n/a	Merged from two contigs with a slight overlap confirmed by raw read BLAST.
Green algae	<i>Chlamydomonas reinhardtii</i>	RefSeq	XP_001702318.1	
	<i>Volvox carteri</i>	RefSeq	XP_002958226.1	
Brown algae	<i>Ectocarpus siliculosus</i>	Genbank	CBJ25800.1	
Bacteria and archaea	<i>Comamonas testosterone</i>	RefSeq	WP_003075952.1	
	<i>Corynebacterium lipophiloflavum</i>	RefSeq	WP_006839975.1	
	<i>Cyanothece</i> sp	RefSeq	YP_001804430.1	
	<i>Cyanothece</i> sp	RefSeq	ZP_01729342.1	
	<i>Cyanothece</i> sp	RefSeq	YP_003887791.1	
	<i>Cyanothece</i> sp	RefSeq	YP_002377271.1	
	<i>Cyanothece</i> sp	RefSeq	YP_003138182.1	
	<i>Desulfuromonas acetoxidans</i>	RefSeq	WP_006002570.1	
	<i>Gallaecimonas xiamenensis</i>	RefSeq	WP_008482472.1	
	<i>Haloferax gibbonsi</i>	RefSeq	WP_004975775.1	
	<i>Halorubrum teberiquichense</i>	RefSeq	WP_006630545.1	
	<i>Halosimplex carlsbadense</i>	RefSeq	WP_006883787.1	
	<i>Janibacter hoylei</i>	RefSeq	WP_007928713.1	
	<i>Ketogulonicigenium vulgare</i>	RefSeq	YP_003964898.1	

	<i>Leptothrix cholodnii</i>	RefSeq	YP_001792238.1	
	<i>Planktomyces maris</i>	RefSeq	WP_002648981.1	
	<i>Planktomyces limnophilus</i>	RefSeq	YP_003631837.1	
	<i>Plesiocystis pacifica</i>	RefSeq	WP_006972868.1	
	<i>Pseudoalteromonas</i> sp	RefSeq	WP_008130733.1	