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Title: Intraluminal magnetisation of bowel by ferromagnetic particles for retraction and manipulation by magnetic probes

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Abstract: Feasibility studies are needed to demonstrate that safe and effective manipulation of bowel during Minimal Access Surgery (MAS) can be obtained by use of magnetic force. This paper characterises two classes of magnetic particles: stainless steel microparticles (SS- μ Ps) and iron oxide nanoparticles (IO-nPs) in terms of their magnetisation, chemical composition, crystallinity, morphology and size distribution. Both magnetic particles were dispersed in a high viscosity biological liquid for intraluminal injection of bowel. Ex-vivo porcine bowel segments were then retracted by permanent magnetic probes of 5.0 and 10 mm diameter. Strong retraction forces reaching 6 N maximum were obtained by magnetic fluid based on dispersion of SS- μ Ps. In contrast, the IO-nP-based magnetic liquid generated less attraction force, due to both lower magnetic and solution properties of the IO-nPs. The comparison of the two particles allowed the identification of the rules to engineer the next generation of particles. The results with SS- μ Ps provide proof on concept that intraluminal injection of magnetic fluid can generate sufficient force for efficient bowel retraction. Thereafter we shall carry out in-vivo animal studies for efficacy and safety of both types of ferrofluids.

1 **Intraluminal magnetisation of bowel by ferromagnetic particles for**
2 **retraction and manipulation by magnetic probes**

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11 **Running head:** Intraluminal bowel magnetization by magnetic micro and nanoparticles

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4 characterises two classes of magnetic particles: stainless steel microparticles (SS- μ Ps) and
5 iron oxide nanoparticles (IO-nPs) in terms of their magnetisation, chemical composition,
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16

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19

1 **1. Introduction**

2 Retraction of bowel during minimal access surgery (MAS) remains problematic as the
3 moist low friction serosal surface is difficult to grasp with laparoscopic graspers. One study
4 documented the low percentage (62 %) of successful grasping actions and indicated the need
5 for improvement in laparoscopic grasper design [1].

6 We have been investigating tissue magnetization by magnetic nano- and micro-particles
7 for MAS applications and have previously reported two tissue ferromagnetisation approaches
8 for tissue retraction: *i*) surface magnetization by applying a small volume of glue-based
9 magnetic media to the mucosal/ serosal surface [2], and *ii*) by interstitial injection of
10 phosphate-buffered saline (PBS) ferrofluids [3]. In these experiments injected
11 ferromagnetisation was shown to be superior to surface magnetisation by surface magnetic
12 pellets, as the latter tended to peel off the tissue during retraction by magnetic probes.
13 However, the restricted sub-mucosal space limited our injected media volume thus retraction
14 force in previous interstitial injection method.

15 In the present study, we report a novel method of magnetisation of a bowel segment by
16 intra-luminal injection of magnetic ferrofluids using a custom-designed intra-abdominal
17 suction-injection probe. We compare the behaviour of stainless steel microparticles (SS- μ Pts)
18 and iron oxide nanoparticles (IO-nPs). The latter family of nPs was chosen due to their well-
19 established clinical use for applications including MRI contrast agents [4-8]. Detailed
20 characterisation of magnetic particles used for making the magnetic ferrofluids for
21 intraluminal bowel magnetisation and the retraction forces obtained in ex-vivo experiments
22 using porcine bowel segments are reported and the comparison of four micro- and nano-
23 particles allowed the identification of the rules to engineer the next generation of particles.

24

25 **2. Materials and methods**

1 2.1. Magnetic particles

2 Stainless steel microparticles and iron oxide nanoparticles were used as model magnetic
3 particles. Two types of stainless steel microparticles (SS- μ Ps) were investigated: stainless
4 steel type 410 microparticles (SS410- μ Ps, from Goodfellow) and stainless steel type 430
5 microparticles (SS430- μ Ps, from Alfa). Iron oxide nanoparticles were made in-house by
6 forced hydrolysis (IO_{IH}-nPs) and compared with commercial iron oxide powder (IO_{Alfa}-nPs,
7 from Alfa). These types of materials were previously used in reported studies on
8 medical/surgical applications [9-11].

9 Preparation of in-house IO_{IH}-nPs: aqueous FeCl₃ and FeCl₂ solutions were prepared
10 separately in degassed DI-H₂O (Milipore). The reaction environment was conditioned with a
11 continuous and sustained flow of nitrogen and with a mechanical stirrer. Co-precipitation of
12 the iron ions was completed at room temperature and by dropwise injection of a NH₄OH
13 solution at a rate of 60 mL/h. The iron oxide formed a black precipitate, which was collected
14 at the bottom of the container and washed with degassed DI-H₂O several times to remove
15 free amine. The NH₃-free nanoparticles were then dried in vacuum before use.

16 All chemical reagents, unless otherwise stated, were purchased from Sigma, used without
17 further purification and degassed before use only if specified: Iron(III) chloride (FeCl₃,
18 reagent grade >97 %), Iron(II) chloride tetrahydrate (FeCl₂.4H₂O, puriss. p.a. \geq 99.0 %),
19 ammonium hydroxide aqueous solution (NH₄OH, 28-30 %).

20

21 2.2. Characterisation of magnetic particles

22 The properties of the four types of particles were characterised with the following
23 experimental techniques.

24 *SQUID*: A 5.0 Tesla Superconducting Quantum Interference Device (SQUID)
25 magnetometer from Quantum Design (MPMS XLTM) was used to quantify the particles

1 magnetic properties. They were dispersed in a polymeric matrix to prevent interaction
2 between nearby particles. The resulting sample was loaded into a low magnetic background
3 gelatin capsule. Hysteresis measurements were completed at 300 K. The magnetization of the
4 gelatine capsules and the polymeric matrix was subsequently subtracted.

5 *XRD*: Wide-angle powder X-ray diffraction (XRD) was used to assess the particle
6 crystallinity and grain size. The data were collected on a Stoe STADI/P powder
7 diffractometer operating in transmission mode and with a small angle position sensitive
8 detector. Incident radiation was generated using a $\text{Fe}_{K\alpha 1}$ source ($\lambda=1.936 \text{ \AA}$). The strongest
9 peak was fitted with Lorentzian-shaped peaks using STOEwinXpow and KaleidaGraph
10 software packages to determine the diffraction peak positions and widths. The crystalline
11 grain size, D_{XRD} , of the particles was calculated according to Scherrer's formula[12]:

$$12 \quad D_{XRD} = \frac{0.9\lambda}{B \cos\theta} \quad (1)$$

13 where D_{XRD} is the "average" dimension of the crystallites, λ is the wavelength of the X-ray
14 source (for Fe source is equal to 0.193604 nm), B is the full width at half maximum of the
15 peak intensity, θ is the glancing angle.

16 *Electron microscopy*: Their sizes and morphology determined by electron microscopy
17 (Transmission (TEM) and Scanning (SEM) electron microscope). TEM images were
18 recorded using a Gatan CCD camera on a JEOL JEM-2011 microscope operating at 200 kV.
19 SEM images were recorded using a Hitachi S-4800 microscope.

20 *ICP-OES*: The chemical composition of all particles was analysed by Inductively
21 Coupled Plasma Optical Emission Spectrometry (ICP-OES). A Perkin Elmer Optima 5300
22 DV was used as Inductively Coupled Plasma Optical Emission Spectrometer. The particles
23 were mixed with hydrochloric acid (Trace SELECT®, $\geq 37 \%$) and nitric acid (Trace
24 SELECT®, $\geq 69.0 \%$) in a 3:1 acid volume ratio and left overnight at room temperature. After

1 full dissolution of the particles, the solution was diluted with deionised water to prepare
2 inductively coupled plasma-atomic emission spectrometry measurements.

3

4 2.3. Preparation of magnetic ferrofluids for intra-luminal bowel injection

5 Magnetic particles were dispersed in phosphate buffered saline (PBS) together with high
6 viscosity fluids such as glycerol (80 % v/v) in order to improve their suspension and
7 dispersion [3]. Concentrations of magnetic particles ranging from 0.1 to 1 g/mL were used for
8 the magnetic bowel retraction studies. In a typical preparation protocol, for one ferrofluid
9 formulation containing 0.25 g/mL fluid, 1.0 g of magnetic particles were added into a
10 Sure/Seal™ bottle which already contained 4 mL glycerol/PBS (80 % v/v) fluid. The
11 magnetic particles were suspended in the liquid by vigorous hand and mechanical shaking of
12 the sealed bottle prior to using the ferrofluid for intraluminal bowel injection.

13 A 10 mm diameter intra-abdominal suction-injection probe was designed and made-in-
14 house. The probe applies initial suction through small suction holes at its distal end before its
15 long needle (19-gauge) is inserted, via a central channel of the probe, into the lumen of the
16 bowel for injection of ferrofluid in large volume. More detailed illustration of our proposed
17 transperitoneal injection can be found from our recently published glue-based magnetic fluid
18 for bowel magnetisation and retraction [13]. This design facilitates intraluminal injection
19 without the need for use of additional graspers.

20 Harvested porcine colons were injected with the ferrofluids, after which the injection
21 device was removed and the magnet probe was brought into contact with the bowel at the
22 injection site. In these experiments, the magnetic attraction force was measured using a
23 tensiometer [2] (Model 5564, Instron Ltd, Buckinghamshire, UK). The magnetic probe was
24 made from neodymium iron boron (NdFeB) disc magnets with a remanence of 1.20 T (grade
25 N35, Eclipse Magnetics Ltd, Sheffield, UK), and diameters of both 5 mm and 10 mm (30 mm

1 in length). The magnetic field at the surface of the magnets was measured with a
 2 magnetometer (Model #DCM 2320, AlphaLab, Inc., Salt Lake City, UT) and found to be
 3 equal to 0.46 T and 0.53 T for the 5mm and 10mm probes, respectively.

4

5 3. Results

6 3.1. Characterisation of magnetic particles

7 Four types of magnetic particles were investigated, Table 1 lists the physical properties as
 8 obtained by SEM and XRD, SQUID at 300 K, and ICP-OES.

9

10 **Table 1**

11 Characterization of the magnetic particles: size as obtained by SEM and XRD characterizations,
 12 magnetization at saturation (M_s) and coercivity (H_c) measured with a SQUID magnetometer at 300 K,
 13 Iron and chromium percentage as deduced from ICP-OES measurements, average maximum of the
 14 magnetic retraction forces F_{max} of a magnetised bowel segments in 20 ex-vivo experiments using the 10
 15 mm magnet probe (injected magnetic media at concentration 0.25 g/mL: intraluminally injected 4 mL).
 16 Normalised retractions forces with the surface of the magnet and the magnetization at saturation.

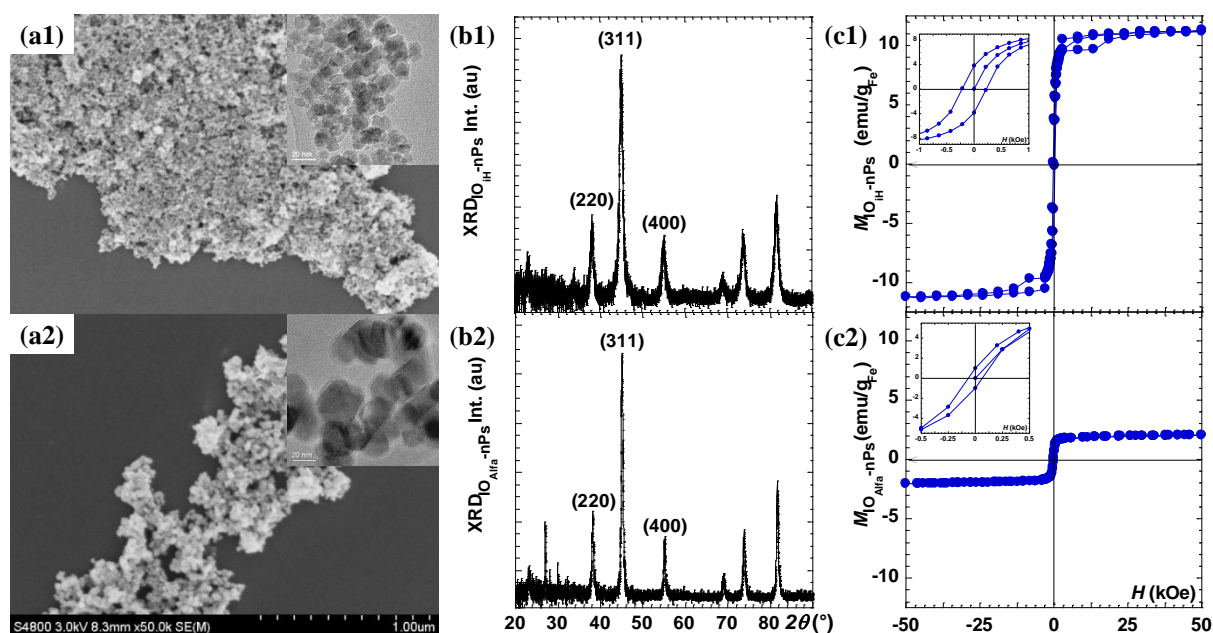
	SS410- μ Ps	SS430- μ Ps	IO _{iH} -nPs	IO _{Alfa} -nPs
Size (diameter)	up to 50 μm^{SEM}	up to 40 μm^{SEM}	$\sim 13 \text{ nm}^{\text{XRD}}$	$\sim 25 \text{ nm}^{\text{XRD}}$
M_s (emu/g)	168.0	110.0	11.0	2.0
H_c (kOe)	0.01	0.02	0.25	0.06
Fe (w %)	86.6 ± 6.8	82.2 ± 6.7	na	na
Cr (w %)	12.6 ± 1.0	16.9 ± 1.4		
F_{max} (N)	5.9 ± 1.0	5.8 ± 0.6	1.3 ± 0.4	2.0 ± 0.1
σ (kPa)	75.67 ± 12.5	74.0 ± 7.0	16.69 ± 4.3	24.97 ± 1.4
F_{max}/M_s (Ng/emu)	0.035 ± 0.006	0.053 ± 0.005	0.12 ± 0.03	0.98 ± 0.10

17

18 Fig. 1a presents the morphology as observed by electron microscopy for iron oxide
 19 nanoparticles. XRD patterns, Fig. 1b, are characteristic of Fe_3O_4 and were used to extract the

1 crystalline grain size. Magnetic properties were quantified at room temperature in terms of
 2 magnetization as function of applied magnetic field and the resulting curves are presented in
 3 Fig. 1c. The morphologies and magnetisation curves for the commercial stainless steel
 4 microparticles were reported earlier elsewhere [13]. As expected, the curves are symmetric
 5 and the magnetization at saturation, M_s , can be deduced from the plateau, when the
 6 magnetization does not increase any more with the magnetic field.

7



8

9 **Fig. 1:** For IOiH-nPs (1) and IOAlfa-nPs (2): Scanning Electron Microscopy images (a), XRD spectra of the
 10 Fe_3O_4 nPs (b) and hysteresis curves completed at room temperature (c). Insets present TEM images of IO-nPS
 11 with a 20 nm scale bar (a) and zoomed magnetisation response for low magnetic fields (c).

12

13 The magnetization curves of both μPs fell to zero magnetisation when the magnetic field
 14 equalled zero, indicating that there was no significant coercivity (H_c) and no observable
 15 remanence (Table 1). The coercivity refers to the magnetic field which needs to be applied to
 16 reduce the magnetization of a material down to zero after the magnetization of the sample has
 17 been driven to saturation. This implies that SS- μPs even though too large to form stable
 18 solution would not face further difficulty in dispersion in the fluid due to magnetic

1 interactions since the particles experience no mutual magnetic attraction until placed in a
2 magnetic field. In contrast, the smaller size nPs would face magnetic interactions at room
3 temperature so that a degree aggregation/ clumping occurs, as observed on the SEM and
4 TEM images in Fig. 1ab.

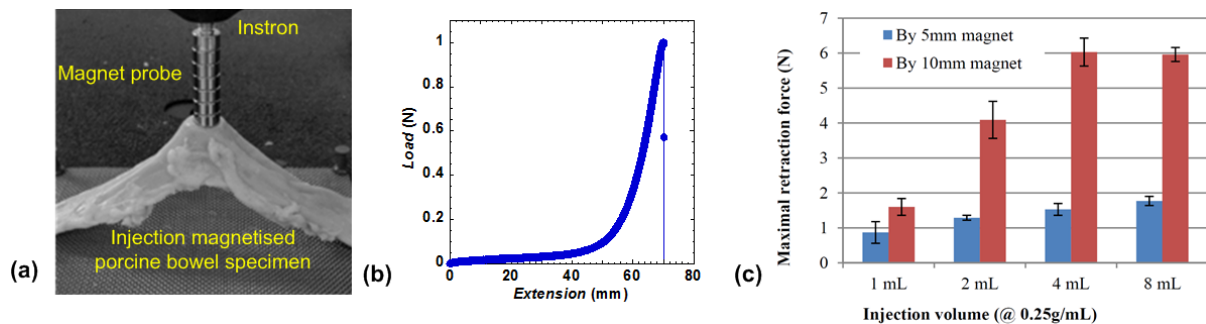
5

6 3.2 Ferrofluid injection and bowel retraction by magnet probe

7 Table 1 also summarises the magnetic retraction forces of a magnetised bowel segments in
8 20 ex-vivo experiments using the 10 mm magnet probes with a 4 mL ferrofluid injection (Fig.
9 2a-b), that is, the number of measurements per type of ferrofluid and probe size was 20 with
10 standard deviations shown in Table 1 and Fig. 2c. Ferrofluids based on both SS-MPs could
11 generate maximal force near 6 N while IO-nPs-based media could only provide about 1 to 2
12 N retraction force.

13 In our previous study [3], we observed that magnetic attraction force increased
14 proportionally with the injected volume of a solution of constant particle concentration.
15 However, the maximal possible injected volume for interstitial (sub-mucosal) injection is
16 limited to 0.2 mL and this limits the retraction force possible with this form of tissue
17 magnetisation. This problem is avoided with intraluminal injection into the bowel lumen.
18 Although one can inject as many fluids as possible into the bowel lumen, a practical threshold
19 could be estimated. For example, as shown in Fig. 2c, there was no or just minor increase in
20 its magnetic retraction force beyond 4 mL injection. Specifically, magnet probes with 10 mm
21 and 5 mm diameters were used. With 10 mm probe, magnetic retraction force increased with
22 increase of injected magnetic volume of ferrofluid and particle content/mass from 0.25 g (1
23 mL) up to 1 g (4 mL). There was almost no retraction force increase beyond 1 g of injected
24 particles, indicative of saturation at this mass/content. The smaller magnet (5 mm) required
25 even less particles to reach its saturation force. As magnetic attraction force between the
26 magnet probe and injected particle decreases exponentially with their distance, any further

1 injection of magnetic ferrofluid simply pushes the magnetic particles further away from the
 2 magnet and thus contributes significantly less to the attraction force.



3
 4 **Fig. 2:** Ex-vivo porcine bowel retraction and force measurement: (a) Photograph of a magnetised bowel
 5 segment retracted during Instron force measurement. (b) Extension-load curve demonstrating a maximum force
 6 before slipping occurs (normalised to unity for illustration purpose). (c) Injection volume versus magnetic
 7 retraction force using two-sized magnet probes.

8

9 4. Discussion

10 The suction-injection probe first tented the target bowel segment and generated large
 11 lumen space in order to facilitate intraluminal injection of ferrofluid into lumen. This avoids
 12 fluid to be injected into sub-mucosa or through the bowel into abdominal space. The injected
 13 fluid was temporarily maintained in the intended lumen section of the intestine due to its high
 14 viscosity (i.e., glycerol suspension), and the magnet probe was then located at the injection
 15 section immediately after withdrawal of the injection probe. Although a very small volume of
 16 fluid, e.g. 4 mL, is needed to interact with the magnet probe for generating sufficient force, in
 17 practical a larger volume (e.g. >10 mL) could be readily injected into the intended lumen
 18 section. Although the fluid would disperse throughout the lumen, in our ex-vivo experimental
 19 setup, we found there was always sufficient ferrofluid remaining to interact with the magnet
 20 probe.

21

1 The average pull force that surgeons use to provide enough tension to the bowel is known
2 to be 2.5 N while the maximal force is just below 5 N.[14] The test results presented in Table
3 1 and Fig. 2 indicate that SS- μ Ps based bowel magnetization would be capable of providing
4 sufficient retraction force for bowel manipulation. The pressure generated between the
5 magnetic fluid and the probe was up to 76 kPa in the present study which would cause less
6 trauma to the target tissue. For example, the pressure generated from this device is
7 significantly lower than that generated by forceps or graspers (210 to 650 kPa [15]), also used
8 for laparoscopic bowel retraction. Much higher trans-mural pressures were used in order to
9 cause tissue ischemia in compressive magnetic anastomosis, for example in gastrointestinal
10 (600 kPa) and bilioenteric (1 MPa) anastomosis [16]. In our application, the probe is removed
11 once the operation is completed, thus the fluid is eliminated in the stools after surgery. A
12 retractable permanent magnet housed inside the probe distal segment could provide an
13 adjustable magnetic attraction force for safe tissue manipulations. For example, in one
14 custom-design, an 8 mm-diameter and 20 mm-long magnet housed within a 10 mm tubular
15 probe produced a maximal retraction force of 3.5 N when the probe was in contact with the
16 magnetised bowel. The force could be reduced to 0.3 N by controlled movement of the
17 magnet, thereby enabling controlled release of the magnetised target bowel. Details about this
18 custom-design magnet probe will be published elsewhere.

19

20 **5. Conclusion**

21 The physical properties of magnetic particles were investigated and the relative
22 performances were compared. Injectable high viscosity magnetic ferrofluids were developed
23 for magnetic bowel retraction and were characterised in an ex-vivo porcine bowel model. The
24 present approach utilizes the magnetic interactions to retract internally magnetised bowels
25 towards an external (intra-abdominal) magnetic probe for safe and effective bowel

1 manipulations. The results of the current experiments indicate that with the technology
2 described, stainless steel micro-particles provide sufficient magnetic attraction force for
3 bowel manipulation. With at least one order of magnitude smaller magnetisation at saturation,
4 IO-nPs fell short by only 20 % to provide of sufficient magnetic attraction force. On-going
5 research is in progress to address the identified challenges and ideal nPs would be engineered
6 to be either super-paramagnetic or coated with a layer providing both solubility and repulsive
7 interactions between the nanoparticles to improve the dispersion in the selected solvent,
8 collection from the jar, and injection into the bowel. To engineer the next generation of
9 particles, these are more important parameters than the magnetisation at saturation of a
10 specific magnetic material. With this in mind, the library of magnetic particles suitable to
11 retract internally magnetised bowels towards an external (intra-abdominal) magnetic probe
12 for safe and effective bowel manipulations should drastically expand. Finally, in-vivo animal
13 studies should next be completed to insure efficacy and safety of ferrofluids.

14

15 **Conflict of interest**

16 The authors report no conflict of interest.

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20

21 **Ethical approval**

22 Ethical approval has not been required for this work.

23

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