cmdc.201402272

Non-natural acetogenin analogues as potent Trypanosoma brucei inhibitors

Gordon J. Florence,* Andrew L. Fraser, Eoin R. Gould, Elizabeth F. B. King,

Stefanie K. Menzies, Joanne C. Morris, Lindsay B. Tulloch and Terry K. Smith^[a]

[a] Dr G. J. Florence,* Mr A. L. Fraser, Dr E. R. Gould, Miss E. F. B. King, Miss S.

K. Menzies, Dr J. C. Morris, Dr L. B. Tulloch and Professor T. K. Smith

EaStCHEM School of Chemistry,

Biomedical Sciences Research Complex

University of St Andrews

North Haugh, St Andrews, KY16 9ST, U. K.

Fax: (+) 44 1334 463808

E-mail: gjf1@st-andrews.ac.uk, tks1@st-andrews.ac.uk

Abstract

A series of novel bis-tetrahydropyran 1,4-triazole analogues based on the acetogenin

framework display low micromolar trypanocidal activities towards both bloodstream

and insect forms of Trypanosoma brucei, the causative agent of African sleeping

sickness. A divergent synthetic strategy was adopted for the synthesis of the key

tetrahydropyran intermediates to enable rapid access to diastereochemical variation

either side of the 1,4-triazole core. The resulting diastereomeric analogues displayed

varying degrees of trypanocidal activity and selectivity in structure activity

relationship studies.

Introduction

Neglected tropical diseases are a continuing health concern in developing countries

due to the lack of effective prevention methods and therapeutic agents. [1] One of these

prevalent Neglected tropical diseases which has been slowly attracting attention over the past few years is African sleeping sickness or Human African Trypanosomiasis (HAT), caused by the protozoan parasite *Trypanosoma brucei* that is transmitted by the bite of the Tsetse fly. HAT is a major health concern in sub-Saharan Africa threatening more than 60 million people. The annual mortality rate of HAT estimated by the World Health Organization (WHO) stands at approximately 8000 with less than 30,000 new cases per year based on recent reported cases. [2] At present the treatment for HAT is extremely limited and dependent upon the disease stage of HAT infection *i.e.* the lymphatic first stage or the second neurological stage when the T. brucei parasite has crossed the blood brain barrier. Four drugs are used in the treatment of HAT (Figure 1), namely suramin (1) and pentamidine (2) for stage one HAT and melarsoprol (3) and effornithine (4) for stage two. These drugs are difficult to administer to patients often requiring lengthy infusion rates, have varying degrees of human toxicity and resistance is becoming a significant problem. [3] In an effort to reduce the costs associated with these drugs, improve the logistics of their administration and reduce drug resistance (a common problem associated with parasitic diseases), a nifurtimox-effornithine combination therapy (NECT, nifurtimox 5 is used clinically to treat Chagas disease) has been recently introduced for stage 2 HAT by the WHO.^[4] This combination therapy maintains the efficacy of effornithine at lower dosages, while reducing the toxic side effects associated with effornithine monotherapy. Despite the success of NECT, there is a lack of new effective therapeutic agents and the onset drug resistance remains a serious threat to current therapies, [5] highlighting the urgent demand for the development of new drug-like molecules and their clinical implementation as effective HAT therapies.

[INSERT FIGURE 1]

The acetogenins are a class of polyketide natural products isolated from the annonaceae plant species found in the tropical regions of West Africa and South America. [6,7] Chamuvarinin (6) was isolated in 2004 by Laurens et. al. from the roots of the bush banana plant *Uvaria chamae* and showed significant cytotoxicity against the KB 3-1 cervix cancer cell line (IC₅₀ value of 0.8 nM), shown in Figure 2.^[8] In 2011 we reported the total synthesis of chamuvarinin, we showed that 6 and a series of synthetic derivatives exhibited low micromolar activities towards both the bloodstream and procyclic forms of *T. brucei*. [9] These encouraging preliminary results and the limited investigations into trypanocidal activity of the acetogenins^[10] prompted our initial interest in designing simplified analogues of 6 which retain the important structural features of the acetogenin family of natural products. In an effort to reduce structural complexity it was hypothesized that a 1,4-triazole motif could form an effective spatial mimic of the central C20-C23 tetrahydrofuran (THF) motif found in chamuvarinin (6). The synthesis and incorporation of THF motifs are notoriously difficult hence, the C16-19 THF ring system of 6 was substituted with the readily accessible and manageable tetrahydropyran (THP) motif, 7, Figure 2. The heterocyclic spacer was envisioned to arise from the "click" reaction of two THP subunits, whilst the butenolide moiety could be appended to the ether core in an analogous manner to 6 by a suitable length alkyl spacer.

[INSERT FIGURE 2]

Results and Discussion

Chemistry

As outlined in Scheme 1, intermediate THP alcohols 8a and 9a could be readily assembled in three steps from (S)-epoxyoctane (10a)[11] via Cu(I)-promoted ring opening of 10a with homoallyl magnesium bromide followed by m-CPBA epoxidation and subsequent in situ acid mediated ring closure to provide a readily separable mixture of diastereomeric alcohols 8a and 9a. This divergent approach provided an excellent opportunity to introduce structural variation within the THP scaffolds. With the alcohols in hand, both diastereomers could be independently converted to the azide by Mitsunobu reaction (DPPA, DIAD, iPr2NEt, PPh3) of alcohols 8a and 9a to give 11a and 12. [12] The enantiomeric syn azide (13) could be accessed in an analogous manner from (R)-epoxyoctane. [11] For the alkyne subunits, syn-8a and anti-9a alcohols were oxidized under Swern conditions to afford aldehydes 14a and 15a in excellent yields and subsequent exposure of 14a to Ohira-Bestmann homologation^[13] gave exclusively syn-16a in 72% yield. Due to epimerization of aldehyde 15a under the mildly basic conditions an alternative twostep Corey-Fuchs homologation^[14] approach was adopted, giving 17a in 70% yield. The incorporation of terminal oxygenation as a functional handle for extended elaboration was achieved by synthesis of the corresponding benzylated and silylated series of azides 11b-c and alkynes 16b and 17b from epoxides 10b and 10c.

[INSERT Scheme 1]

The central 1,4-triazole scaffold was conveniently installed by application of the Cu(I) catalyzed Huisgen 1,3-dipolar cycloaddition (Scheme 2).^[15] Thus, diastereomeric analogues of the alkyl, benzylated and silylated bis-THP triazole motifs were synthesized by treatment of the corresponding azides **11a-c**, **12**, **13** and

alkynes **16a-b**, **17a-b** under "click" reaction conditions (CuSO₄, Na ascorbate) to afford triazole products **18a-d**, **19a-b**, **20a-b**, **21a-b** and **22** in good yields.

[INSERT Scheme 2]

Benzylated **18b-c**, **19b**, **20b**, **21b** and **22** analogues, outlined in Scheme 3, were deprotected under atmospheric hydrogenolysis (20% Pd(OH)₂/C, H₂) to the corresponding alcohols **23-28** (entries 1-6) in excellent yields.

[INSERT Scheme 3]

Table in Scheme to be below Scheme 3 – both original and revised submitted

Entry	Triazole	Product	Yield (%)	
1	18b : $R^1 = Et$, $R^2 = OBn$	23: R ¹ = Et, R ² = OH	92	
2	19b : $R^1 = Et$, $R^2 = OBn$	24 : $R^1 = Et$, $R^2 = OH$	95	
3	21b : $R^1 = Et$, $R^2 = OBn$	25 : R ¹ = Et, R ² = OH	93	
4	22 : $R^1 = Et$, $R^2 = OBn$	26 : R ¹ = Et, R ² = OH	92	
5	22 : $R^1 = Et$, $R^2 = OBn$	27 : $R^1 = Et$, $R^2 = OH$	90	
6	18c : $R^1 = OBn$, $R^2 = Et$	28 : R ¹ = OH, R ² = Et	100	

Having previously established SAR data for the advanced analogues of chamuvarinin, it was observed that incorporation of the butenolide side chain impacted positively on the trypanocidal activity. Thus, installation of the butenolide motif to successfully mimic chamuvarinin was accomplished in four steps from lead alcohol 23, outlined in Scheme 4. Alcohol 23 was manipulated to sulfone 29 *via* a two-step Mitsunobu reaction and catalytic Mo[VI]^[7] oxidation protocol.^[16] Subsequent Julia-Kocienski olefination^[16] with aldehyde 30^[9,18] and diimide reduction^[19] (TsNHNH₂, NaOAc)

provided advanced analogue **31**. To ascertain whether the triazole analogue would bind in a directional manner to the unknown protein target, analogue **32** was prepared in an analogous manner to **31** from alcohol **28** *via* sulfone **33**.

[INSERT Scheme 4]

Having successfully installed the butenolide motif and in an effort to further extend the SAR data, bis-directional analogue 34, outlined in Scheme 5, was synthesized in six steps from advanced triazole 18d. Silyl deprotection of 18d to alcohol 35, followed by a Mitsunobu/oxidation protocol gave sulfone 36. Julia-Kocienski olefination of sulfone 36 with 30 and subsequent diimide reduction gave triazole 37. Benzyl deprotection of 37 with boron trichloride furnished alcohol 34 in 23% yield. Diol analogue 38 was accessed in excellent yield from alcohol 35 under atmospheric hydrogenolysis.

[INSERT Scheme 5]

Structure-activity relationship summary

The primary mode of action for acetogenins in mammalian cells is inhibition of Complex I of the mitochondrial electron transport chain (ETC).^[20] In contrast, bloodstream (*BSF*) *T. brucei* lacks functional Complexes I, III and IV and instead relies on alternative oxidases (TAO, AOX2) to enable mitochondrial respiration, while the procyclic (*Pro*) form expresses both a fully functional ETC and the alternative oxidases.^[21] Thus in order to assess whether the triazole based analogues share a common mitochondrial target and identify key structural features required for

parasitic inhibition screening against the bloodstream and procyclic forms of *T. brucei* and the mammalian HeLa cell line, [22] was performed (Table 1). From the data it was evident that the stereochemistry surrounding the THP ring systems and the functionalization of the terminal motifs impacted on the *T. brucei* inhibition profile. In comparison to chamuvarinin (6), the initial set of diastereomeric alkyl triazoles 20a, 21a and 18a (entries 1, 4, 7) were >17 times less potent than 6 and indicated that stereochemistry had little effect on the inhibition profile. Surprisingly analogue 19a (entry 12) displayed good levels of T. brucei inhibition with an EC₅₀ value of 7.6 \pm 0.3 µM and moderate levels of parasite selectivity (SI = 21.9). Introduction of terminal oxygenation as a handle for further manipulation to one side of the alkyl side chain, analogues 21b, 18b, 19b and 22 (entries 5, 8, 13, 15), resulted in good micromolar activities with EC₅₀ values <10 μM. Interestingly, the *anti-anti* analogue **20b** (entry 2) was devoid of HeLa and *T. brucei* activity. By comparison, analogue 21b (entry 5) displayed good levels of parasite inhibition and selectivity with an SI of 23. For analogues 24 and 27 (entries 14 and 16), removal of the benzyl group resulted in ~2-fold loss in parasite activity. Removal of the benzyl group from analogue 21b (entry 5) to give 26 (entry 6) led to a significant loss in parasitic activity and selectivity, EC₅₀ values of 3.1 \pm 0.1 and 72.4 \pm 3.9 μ M and SI's of 23 and 0.8, respectively. The *anti-anti* analogue **25** (entry 3) was still devoid of parasitic activity, despite transformation to the free hydroxyl. Gratifyingly, introducing the free hydroxyl to analogue 23 (entry 9) led to a slight increase in T. brucei inhibition and 23 has shown comparable levels of parasitic activity to that of chamuvarinin (6), EC₅₀ values of 1.80 ± 0.10 and 1.4 ± 0.1 μ M, respectively. On the basis of 23, the bisdirectional analogue 35 (entry 10) was tested in the hope of further improving the activity. Unfortunately, 35 was 9 times less potent than 23 and subsequent removal of the benzyl group to give diol **38** (entry 11) resulted in significantly diminished activities. Comparison of the biological data for the ethyl, benzyl and alcohol triazole series, implies that the benzyl group may be potentially interacting with residues at a protein target site, which an alcohol group cannot accommodate. Alternatively, or as well as the benzyl group may be able to readily insert itself into the lipophilic membrane, whereas the hydrophilic alcohol incurs a greater penalty for membrane insertion.

[Insert Table 1 – double column width – appended at end of manuscript]

Caption for Table 1

Table 1. SAR data for ethyl analogues **20a**, **21a**, **18a**, **19a**, benzyl analogues **20b**, **21b**, **18b**, **35**, **19b**, **22** and alcohol analogues **23-27**, **38**; selectivity index refers to *BSF* parasitic inhibition versus mammalian HeLa inhibition; chamuvarinin (**6**, *T. brucei* (*BSF*) $EC_{50} = 1.4 \pm 0.1 \mu M$; *T. brucei* (*Pro*) $EC_{50} = 1.9 \pm 0.1 \mu M$; HeLa $EC_{50} = 2.9 \pm 0.7 \mu M$ taken from ref. 9).

The synthesis of triazole analogues with terminal oxygenation has provided a functional handle for further elaboration (Table 2). The non-natural chamuvarinin-like analogues previously reported revealed that introduction of the butenolide side chain resulted in a >5-fold increase in *T. brucei* activity. Encouraged by the potent activity of lead alcohol 23 (entry 1), it was decided to incorporate the butenolide moiety based on this structure. Analogues 31 and 32 (entries 2 and 3) clearly highlight the importance of the spatial orientation of the pendent butenolide sidechain. While 31 was essentially inactive, 32 displayed low micromolar selective activities against both forms of *T. brucei* with EC₅₀ values of 3.2 ± 0.1 and 5.7 ± 0.6 µM respectively and a

selectivity index of 15.8. This suggests that although the structures are only subtly different at face value, their binding/interaction is highly specific and indicative of a protein target, rather than the biophysical properties of the compounds.

[Insert Table 2] – Figure containing structures to be inserted above table 2 Caption for Table 2

Table 2. Biological profiles for analogues 23, 31-32, 34 and 37, selectivity index refers to BSF parasitic inhibition versus mammalian HeLa inhibition.

Entry	Analogue	T. brucei (BSF) EC ₅₀ (μM)	T. brucei (Pro) EC ₅₀ (μM)	HeLa EC ₅₀ (μM)	Selectivity Index	
1	23	1.8 ± 0.1	0.8 ± 0.11	7.0 ± 1.0	3.9	
2	31	>500	>1000	>1000	-	
3	32	3.2 ± 0.1	5.7 ± 0.6	50.8 ± 3.7	15.8	
4	37	5.2 ± 0.3	19.2 ± 0.8	2.1 ± 0.4	0.4	
5	34	28.5 ± 4.6	.6 5.6 ± 0.6 8.3 ± 1.1		0.3	

On the basis of the SAR data, incorporation of both the butenolide moiety and a benzyl-protected alcohol in place of the alkyl side chain indicated that the benzyl analogue 37 (entry 4) had similar activity to 32. Removal of the benzyl group to reveal the free hydroxyl analogue 34 (entry 5), was found to diminish activity and was 9 times less active than 32, with EC₅₀ values of 28.5 ± 4.6 and 3.2 ± 0.1 µM, respectively. This may indicate that a non-hydrophilic arm on the opposite side of the butenolide moiety is required for maximum protein/analogue/lipid interaction.

Lipophilic efficiency

A theoretical comparison of the triazole analogues molecular properties was conducted in an effort to highlight the lipophilicity of these compounds and provide an indication as to whether they would be suitable drug candidates. As outlined in Table 3, the data indicated that our lead compound alcohol 23 (entry 1) displayed a suitable log P of 5.163, had a molecular weight less than 500 and had a suitable number of hydrogen bond donors and acceptors, adhering to Lipinski's rule of 5. Introduction of the butenolide moiety on one end of the THP ring system and an alkyl side chain on the opposite side in analogue 32 (entry 2), led to an increased log P value of 9. Exchange of the hydrophobic alkyl side chain with either a benzyl group (37, entry 4) or a free hydroxyl group (34, entry 3) decreased log P values while increasing the potential for hydrogen bond donation. Chamuvarinin itself (6, entry 5) displayed a high log P value of 9 in line with the observed ability of acetogenin family members to cross the blood-brain barrier^[23] The lipophilic efficiency (LipE) is a combination of the calculated lipophilicity (clogP) and the potency (pEC₅₀) of analogues to estimate the drug-likeness (LipE >5) of compounds. As highlighted in Table 3, triazoles 23, 32, 34, 37 and chamuvarinin 6 exhibited values ranging from +0.6 to -3.7, which are lower than the proposed optimal LipE of typical drug candidates. The calculated LipE values (Table 3) for triazoles 23, 32, 34, 37 and chamuvarinin 6 fall in the range +0.6 to -3.7, which although below the optimal value are in line with values often displayed by natural products and their derivatives.^[24] Furthermore the LipE values of our designed triazole analogues are comparable with bioactive compounds that cross the blood brain barrier^[23,24] which when coupled with their toxicity towards T. brucei provides the exciting prospect of developing an

effective dual pronged treatment for Stage 1 and 2 HAT if selectivity profiles can be further improved.

[INSERT TABLE 3]

Table 3. Theoretical data of molecular properties for triazole analogues 6, 23, 32, 34 and 37.

Entry	Compound	T. brucei (BSF) EC ₅₀ (μM)	pEC ₅₀	clog P	LipE	MW	H bond donors	H bond acceptors
1	23	1.8 ± 0.1	5.75	5.163	0.59	407.6	1	6
2	32	3.2 ± 0.1	5.49	9.056	-3.57	571.8	0	7
3	34	28.5 ± 4.6	4.55	7.336	-2.79	559.8	1	8
4	37	5.2 ± 0.3	5.29	8.935	-3.65	649.9	1	8
5	6	1.4 ± 0.1	5.86	9.208	-3.35	604.9	1	6

Molecular modelling

Our initial rationale for the design and synthesis of simplified triazole analogues was based on the molecular modelling of the central tricyclic core of chamuvarinin (6), as outlined in Figure 3a, indicating that 6 adopts a "U-shape" conformation. We hypothesized that a 5-membered heterocyclic spacer (Figures 3b and 3c) would act as an effective spatial mimic for the central THF core of 6 and thus retain potent biological activity. Modelling highlights that the lowest energy conformation of the *syn-syn* bis-THP rings acts a better mimic of this 'U-shaped' conformation than the corresponding *anti-anti* and this would appear to be broadly born out by the biological results. The encouraging trypanocidal activity of the triazole compounds discussed in this paper has now formed the basis of further design iterations and lead structure optimization. The evolving structure-activity-relationship

will be enabled by further cycles of iterative design towards the development of more potent and selective trypanocidal compounds.

[INSERT FIGURE 3]

Conclusion

A focused library of 1,4-triazole-based acetogenin analogues have been synthesized and have demonstrated low micromolar trypanocidal activities against both bloodstream and procyclic forms of *T. brucei*. Analogue 23 has displayed comparable levels of *T. brucei* inhibition to that of natural product chamuvarinin (6), EC₅₀ values of 1.8 ± 0.1 and 1.4 ± 0.1 µM, respectively demonstrating that this series of triazole analogues are potential lead compounds for the development of an effective therapeutic agent for HAT. The acetogenins have long been established as mitochondrial Complex I inhibitors within mammalian cells, therefore, it is postulated that the analogues are targeting a protein within the mitochondrion of the parasite. The specific protein target and mode of action for parasite inhibition is currently undetermined for these analogues. However, analogues 31 and 32 clearly highlight that spatial orientation of the butenolide side chain impacts heavily on T. brucei inhibition. Current focus is to establish whether the analogues are disrupting mitochondrial functions through the incorporation of fluorescent and affinity tags to lead triazole compounds in order to isolate specific proteins of interest, allowing them to be genetically and chemically validated.

Experimental Section

See supporting information for full experimental details

Acknowledgements

This work was funded by the Royal Society, the Leverhulme Trust, the EPSRC (GJF) and the Wellcome Trust (TKS, WT 093228). We thank the EPSRC National Mass Spectrometry Service Centre, Swansea, UK for mass spectrometry services.

Keywords: acetogenin, Neglected diseases, HAT, *T. brucei*, stereochemistry

- [1] L. Zhou, G. Stewart, E. Rideau, N. J. Westwood, T. K. Smith, *J. Med. Chem.* **2013**, *56*, 796-806.
- [2] *Trypanosomiasis, African*; World Health Organization: Geneva, **2012**; http://www.who.int/topics/trypanosomiasis_african/en/ (Accessed 19 June, **2014**)
- [3] a) D. Steverding, *Parasites & Vectors* **2010**, *3*: 15; b) V. Delespaux, H. P. de Koning, *Drug Resistance Updates*, **2007**, *10*, 30-50.
- [4] G. Priotto, S. Kasparian, W. Mutombo, D. Ngouama, S. Ghorashian, U. Arnold, S. Ghabri, E. Baudin, V. Buard, S. Kazadi-Kyanza, M Ilunga, W. Mutangala, G. Pohlig, C. Schmid, U. Karunakara, E. Torreele, V. Kande, *Lancet*, 2009, 374, 56-64.
- [5] A. Y. Sokolova, S. Wyllie, S. Patterson, S. L. Ozam, K. D. Read, A. H. Fairlamb, Antimicrob. Agents Chemother. 2010, 54, 2893-2900.
- [6] a) F. Q. Alali, X. -X. Liu, J. L. McLaughlin, *J. Nat. Prod.* 1999, 62, 504-540; b)
 M. -C. Zafra-Polo, B. Figadère, T. Gallardo, J. R. Tormo, D. Cortes, *Phytochemistry*, 1998, 48, 1087-1117; c) M. C. Zafra-Polo, M. C. González, E. Estornell, S. Sahpaz, D. Cortes, *Phytochemistry*, 1996, 42, 253-271; d) L. Zeng, Q. Ye, N. H. Oberlies, G. Shi, Z. -M. Gu, K. He, J. L. McLaughlin, *Nat. Prod. Rep.* 1996, 13, 275-306; e) J. K. Rupprecht, Y. H. Hui, J. L. McLaughlin, *J. Nat. Prod.* 1990, 53, 237-278.

- [7] A. Bermejo, B. Figadère, M. C. Zafra-Polo, I. Barrachina, E. Estornell, D. Cortes, *Nat. Prod. Rep.* **2005**, *22*, 269-303.
- [8] a) D. Fall, R. A. Duval, C. Gleye, A. Laurens, R. Hocquemiller, J. Nat. Prod. 2004, 67, 1041-1043; b) S. Derbré, E. Poupon, C. Gleye, R. Hocquemiller, J. Nat. Prod. 2007, 70, 300-303.
- [9] a) G. J. Florence, J. C. Morris, R. G. Murray, R. R. Vanga, J. D. Osler, T. K. Smith, *Org. Lett.* 2011, *13*, 514-517; b) G. J. Florence, J. C. Morris, R. G. Murray, R. R. Vanga, J. D. Osler, T. K. Smith, *Chem. Eur. J.* 2013, *19*, 8309-8320.
- [10]a) S. Sahpaz, C. Bories, P. M. Loiseau, D. Cortès, R. Hocquemiller, A. Laurens, A. Cavé, *Planta. Med.* 1994, 60, 538-540; b) A. -I. Waechter, G. Yaluff, A. Inchausti, A. Rojas de Arias, R. Hocquemiller, A. Cavé, A. Fournet, *Phytother. Res.* 1998, 12, 541-544; c) S. Hoet, F. Opperdoes, R. Brun, J. Quetin-Leclercq, *Nat. Prod. Rep.* 2004, 21, 353-364.
- [11]a) G. C. Paddon-Jones, C. S. P. McErlean, P. Hayes, C. J. Moore, W. A. Konig, W. Kitching, *J. Org. Chem.* 2001, 66, 7487-7495; b) S. E. Schaus, A. D. Brandes, J. F. Larrow, M. Tokunga, K. B. Hansen, A. E. Gould, M. E. Furrow, E. N. Jacobsen, *J. Am. Chem. Soc.* 2002, 124, 1307-1315.
- [12] J. P. Scott, M. Alam, N. Bremeyer, A. Goodyear, T. Lam, R. D. Wilson, G. Zhou, Org. Process Res. Dev. 2011, 15, 1116-1123.
- [13] a) S. Müller, B. Liepold, G. J. Roth, H. J. Bestmann, *Synlett* 1996, 521-522; b) G.
 J. Roth, B. Liepold, S. G. Müller, H. J. Bestmann, *Synthesis* 2004, 59-62.
- [14] E. J. Corey, P. L. Fuchs, *Tetrahedron Lett.* **1972**, *13*, 3769-3772.
- [15]a) V. V. Rostovtsev, L. G. Green, V. V. Fokin, K. B. Sharpless, *Angew. Chem. Int. Ed.* 2002, 41, 2596-2599; b) C. W. Tornøe, C. Christensen, M. Meldal, *J. Org. Chem.* 2002, 67, 3057-3064.

- [16] H. S. Schultz, H. B. Freyermuth, S. R. Buc, J. Org. Chem. 1963, 28, 1140-1142.
- [17]a) P. R. Blakemore, W. J. Cole, P. J. Kocieński, A. Morley, *Synlett* 1998, 26-28;
 b) P. R. Blakemore, *J. Chem. Soc. Perkin Trans.* 1, 2002, 2563-2585;
 c) R. I. Dumeunier, I. E. Markó, *Modern Carbonyl Olefination: Methods and Applications* (Ed.: T. Takeda), Wiley-VCH, Weinheim, 2004, 104-150.
- [18] The butenolide aldehyde **30** was synthesized in five steps from enantioenriched (S)-propylene oxide, see Ref 8a.
- [19]a) J. A, Marshall, M. Chen, J. Org. Chem. 1997, 62, 5996-6000; b) M. T. Crimmins, Y. Zhang, F. A. Diaz, Org. Lett. 2006, 8, 2369-2372.
- [20] a) M. Degli, Esposti, A. Ghelli, M. Ratta, D. Cortes, E. Estronell, *Biochem. J.*1994, 301, 161-167; b) N. De Pedro, B. Cautain, A. Melguizo, F. Vicente, O. Genilloud, F. Pelaez, J. R. Tormo, *J. Bioenerg. Biomembr.* 2013, 45, 153-164.
- [21] For a review, see: A. G. Tielens, J. J. van Hellemond, *Trends Parasitol.* **2009**, *25*, 482–490
- [22] Biological testing was performed using the Alamar Blue viability test as described in: J. Mikus, D. Steverding, *Parasitol. Int.* **2000**, *48*, 265-269.
- [23] P. Champy, G. U. Höglinger, J. Féger, C. Gleye, R. Hocquemiller, A. Laurens, V. Guérineau, O. Laprévote, F. Medja, A. Lombès, P. P. Michel, A. Lannuzel, E. C. Hirsch, M. Ruberg, *J. Neurochem.* 2004, 88, 63–69.
- [24] I. Jabeen, K. Pleban, U. Rinner, P. Chiba, G. F. Ecker, *J. Med. Chem.* **2012**, *55*, 3261-3273.
- [25] Schrodinger Maestro (v9.7.014) and Macromodel (v10.4) were used for 10000 step Monte-Carlo searches using the MMFF force field.

Figure Captions

Caption for Figure 1

Figure 1. Current drugs for treatment of Human African trypanosomiasis.

Caption for Figure 2

Figure 2. Rationale for simplified analogue design.

Caption for Scheme 1

Scheme 1. Synthesis of azides **11a-c**, **12** and **13** and alkynes **16a-b** and **17a-b**: a) $CH_2CHCH_2CH_2MgBr$, CuI, THF, -40 °C \rightarrow RT; b) mCPBA, CH_2Cl_2 , 0 °C \rightarrow RT; then (±)-CSA (20 mol%), RT; c) PPh₃, iPr₂NEt, DIAD, DPPA, 0 °C \rightarrow RT; d) (COCl)₂, DMSO, CH_2Cl_2 , Et_3N , -78 °C \rightarrow RT; e) dimethyl diazo-2-oxopropylphosphonate, K_2CO_3 , MeOH, RT; f) CBr_4 , PPh₃, CH_2Cl_2 , 0 °C then -78 °C; g) n-BuLi, THF, -78 °C. mCPBA = 3-chloroperbenzoic acid, (±)-CSA = (±)-camphorsulfonic acid, DIAD = diisopropyl azodicarboxylate, DPPA = diphenyl phosphoryl azide, DMSO = dimethyl sulfoxide

Caption for Scheme 2

Scheme 2. Synthesis of triazole analogues **18a-d**, **19a-b**, **20a-b**, **21a-b** and **22**: a) CuSO₄•5H₂O, Na ascorbate, H₂O, *t*-BuOH, RT.

Caption for Scheme 3

Scheme 3. Synthesis of alcohols 23-28: a) 20% Pd(OH)₂/C, H₂ (1 atm), EtOH, RT.

Caption for Scheme 4

Scheme 4. Synthesis of triazoles **31** and **32**: a) 1*H*-mercaptophenyltetrazole, PPh₃, DIAD, 0 °C; b) (NH₄)₆Mo₇O₂₄•4H₂O, H₂O₂, EtOH, 0 °C \rightarrow RT; c) NaHMDS, THF, -78 °C; then **30**, THF -78 \rightarrow -20 °C; d) TsNHNH₂, NaOAc, DME, H₂O, 100 °C. PT = phenyl tetrazole, NaHMDS = sodium hexamethyldisilylazide, DME = dimethoxyethane.

Caption for Scheme 5

Scheme 5. Synthesis of advanced alcohol **34**: a) TBAF, THF, 0 °C \rightarrow RT; b) 1*H*-mercaptophenyltetrazole, PPh₃, DIAD, 0 °C; c) (NH₄)₆Mo₇O₂₄•4H₂O, H₂O₂, EtOH, 0 °C \rightarrow RT; d) NaHMDS, THF, -78 °C; then **30**, THF -78 \rightarrow -20 °C; e) TsNHNH₂, NaOAc, DME, H₂O, 100 °C; f) BCl₃•SMe₂, CH₂Cl₂, -78 °C \rightarrow RT; g) 20% Pd(OH)₂/C, H₂(1 atm), EtOH, RT. TBAF = tetrabutylammonium fluoride.

Caption for Table 1

Table 1. SAR data for ethyl analogues **20a**, **21a**, **18a**, **19a**, benzyl analogues **20b**, **21b**, **18b**, **35**, **19b**, **22** and alcohol analogues **23-27**, **38**; selectivity index refers to *BSF* parasitic inhibition versus mammalian HeLa inhibition; chamuvarinin (**6**, *T. brucei* (*BSF*) $EC_{50} = 1.4 \pm 0.1 \mu M$; *T. brucei* (*Pro*) $EC_{50} = 1.9 \pm 0.1 \mu M$; HeLa $EC_{50} = 2.9 \pm 0.7 \mu M$ taken from ref. 9).

Caption for Table 2

Table 2. Biological profiles for analogues 23, 31-32, 34 and 37, selectivity index refers to BSF parasitic inhibition versus mammalian HeLa inhibition.

Caption for Table 3

Table 3. Theoretical data of molecular properties for triazole analogues 6, 23, 32, 34 and 37.

Caption for Figure 3

Figure 3. Molecular modelling of the lowest energy conformations for a) chamuvarinin framework; b) *syn-syn* triazole core; and c) *anti-anti* triazole core.

Text for the Table of Contents

A series of novel bis-tetrahydropyran 1,4-triazole analogues based on the acetogenin framework display low micromolar trypanocidal activities towards both bloodstream and insect forms of *Trypanosoma brucei*, the causative agent of African sleeping sickness.

Keywords

acetogenin, Neglected diseases, HAT, T. brucei, stereochemistry

cmdc.201402272 : Figure 1

Na
$$O_3$$
S HN HN S O_3 Na Suramin S O_3 Na So O

cmdc.201402272 : Figure 2

T. brucei (*Pro*) EC_{50} = 1.90 ± 0.11 μM

cmdc.201402272 : Scheme 1

cmdc.201402272 : Scheme 2

cmdc.201402272: Scheme 3 - original

 R^1 = Et or OBn, R^2 = Et or OBn

$$R^1$$
 = Et or OH, R^2 = Et or OH

Entry	Triazole	Product	Yield
1	18b : $R^1 = Et$, $R^2 = OBn$	23 : $R^1 = Et$, $R^2 = OH$	92%
2	19b : $R^1 = Et$, $R^2 = OBn$	24 : $R^1 = Et$, $R^2 = OH$	95%
3	20b : $R^1 = Et$, $R^2 = OBn$	25 : $R^1 = Et$, $R^2 = OH$	93%
4	21b : R ¹ = Et, R ² = OBn	26 : $R^1 = Et$, $R^2 = OH$	92%
5	22 : R ¹ = Et, R ² = OBn	27 : R ¹ = Et, R ² =OH	90%
6	18c : R ¹ = OBn, R ² = Et	28 : R ¹ = OH, R ² = Et	100%

cmdc.201402272 : Scheme 4

23: R¹ = Et, R² = OH

28: $R^1 = OH$, $R^2 = Et$

29:
$$R^1$$
 = Et, R^2 = SO_2PT , 61% over 2 steps **33**: R^1 = SO_2PT , R^2 = Et, 71% over 2 steps

30

31: R^1 = Et, R^2 = **A**, 53% over 2 steps

32: $R^1 = A$, $R^2 = Et$, 49% over 2 steps

cmdc.201402272: Scheme 5

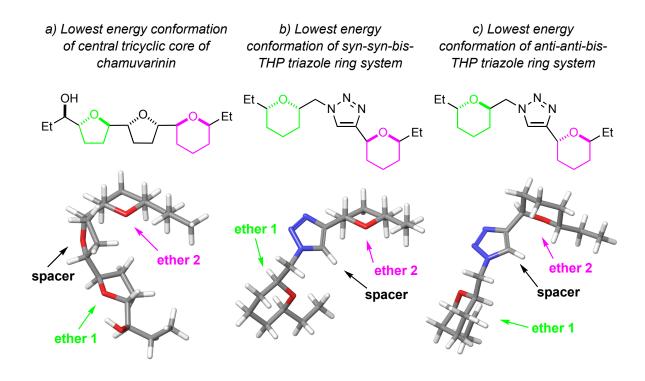
Table 1:

Entry	Analogue	R group		EC ₅₀ (μM)		Selectivity Index	Entry	Analogue	R group		EC ₅₀ (µM)		Selectivity Index
			T. brucei (BSF)	T. brucei (Pro)	HeLa	muox				T. brucei (BSF)	T. brucei (Pro)	HeLa	moox
1		20a	24.6 ±	65.0 ±	33.0 ±	1.3	9		23	1.8 ±	0.8 ±	7.0 ±	3.9
		R = Et	1.4	3.1	2.4				$R^1 = Et$ $R^2 = OH$	0.1	0.11	1.0	
2	$N_{N_{N_{N_{N_{N_{N_{N_{N_{N_{N_{N_{N_{N$	20b	467.2 ±	236.6 ±	>1000	>2	10	,'0',' N'N'N	35	16.2 ±	19.4 ±	>100	>6
		R = OBn	40.3	23.8				R^1 R^2	$R^1 = OH$ $R^2 = OBn$	0.9	0.5		
3	R	25	223.2 ±	87.9 ±	415.8 ±	1.9	11	R*	38	157.8 ±	99.0 ±	>250	>1.5
		R = OH	21.8	4.5	35.3				$R^1 = R^2 = OH$	13.2	2.5		
4		21a	37.2 ±	81.5 ±	>500	>6	12		19a	7.6 ±	1.1 ±	166.6	21.9
	~ .O. ~ N.N.	R = Et	6.6	4.5				~ ,0, ~ N,N	R = Et	0.3	0.1	± 18.0	
5		21b	3.10 ±	48.7 ±	71.4 ±	23	13		19b	10.1 ±	72.3 ±	76.0 ±	7.5
		R = OBn	0.1	3.6	4.1				R = OBn	0.4	5.3	3.0	
6	R	26	72.4 ±	68.3 ±	59.7 ±	0.8	14	Ŕ	24	17.5 ±	25.4 ±	26.9 ±	1.5
		R = OH	3.9	7.2	2.5				R = OH	1.0	1.3	1.1	
7	N	18a	48.7 ±	103.3 ±	>100	>2	15		22	8.2 ±	15.5 ±	37.5 ±	4.6
	, O, , N, N	$R^1 = R^2 = Et$	3.5	3.5				O N.N.N	R = OBn	0.4	1.0	1.5	
8	R^1	18b	8.70 ±	15.6 ±	45.5 ±	5.3	16		27	19.2 ±	42.9 ±	101.0	5.3
	R ²	$R^1 = Et$ $R^2 = OBn$	0.5	0.3	2.5			R	R = OH	1.1	2.4	± 10.4	

cmdc.201402272: Table 2 Graphic - original

Entry	Analogue	T. brucei (BSF) EC ₅₀ (μM)	T. brucei (Pro) EC ₅₀ (μM)	HeLa EC ₅₀ (μΜ)	Selectivity Index	
1	23	1.8 ± 0.1	0.8 ± 0.1	7.0 ± 1.0	3.9	
2	31	>500	>1000	>1000	-	
3	32	3.2 ± 0.1	5.7 ± 0.6	50.8 ± 3.7	15.8	
4	37	5.2 ± 0.3	19.2 ± 0.8	2.1 ± 0.4	0.4	
5	34	28.5 ± 4.6	5.6 ± 0.6	8.3 ± 1.1	0.3	

cmdc.201402272 : Figure 3



cmdc.201402272: TOC Graphic