

1 Genetic coupling of mate recognition systems in the genomic era.

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18 Abstract

19 The concept of Genetic Coupling in mate recognition systems arose in the 1960s as a potential
20 mechanism to maintain coordination between signals and receivers during evolutionary divergence.
21 At its most basic it proposed that the same genes might influence trait and preference and therefore
22 mutations could result in coordinated changes in both traits. Since then, the concept has expanded
23 in scope and is often used to include linkage or genetic correlation between recognition system
24 components. Here we review evidence for genetic coupling, concentrating on proposed examples of
25 a common genetic basis for signals and preferences. Mapping studies have identified several
26 examples of tight genetic linkage between genomic regions influencing signals and preferences, or
27 assortative mating. Whether this extends as far as demonstrating pleiotropy remains a more open
28 question. Some studies, notably of *Drosophila*, have identified genes in the sex determination
29 pathway and in pheromonal communication where single loci can influence both signals and
30 preferences. This may be based on isoform divergence, where sex- and tissue-specific effects are
31 facilitated by alternative splicing, or on regulatory divergence. Hence it is not clear that such
32 examples provide compelling evidence of pleiotropy in the sense that “magic mutations” could
33 maintain trait coordination. Rather, co-evolution may be facilitated by regulatory divergence but
34 require different mutations or coevolution across isoforms. Reconsidering the logic of genetic
35 coupling, it may be that pleiotropy could actually be less effective than linkage if distinct but
36 associated variants allow molecular coevolution to occur more readily than potentially “unbalanced”
37 mutations in single genes. Genetic manipulation or studies of mutation order effects during
38 divergence are challenging but perhaps the only way to disentangle the role of pleiotropy versus
39 close linkage in coordinated trait divergence.

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42 **Introduction**

43 A critical stage in the evolution of sexual isolation is divergence in signals involved in sexual
44 communication and their associated preferences. If a major signal trait and an associated preference
45 both diverge between populations in a coordinated manner, assortative mating is likely to result and
46 to contribute to reproductive isolation. Pre-mating isolation can also be favoured by divergence in
47 seasonality, habitat choice or ecologically adapted traits that indirectly lead to assortative mating
48 (Kopp et al. 2018), but sexual isolation in animals is often thought to emerge most directly from
49 coordinated changes in mating signals (here interpreted broadly as any trait in one sex that can
50 influence mate choice by the other sex) and associated preferences (Lande 1981; Panhuis et al.
51 2001; Ritchie 2007). Jointly, we call these coordinated signals and preferences “Mate Recognition
52 Systems” (after Paterson 1978).

53 Sexual Isolation and Coevolution

54 Sexual isolation is often thought to be amongst the first forms of reproductive isolation to appear in
55 many animal groups (see Shaw et al. in this volume) and therefore potentially plays an important
56 role in animal speciation. Ultimately, mate recognition systems can diverge due to sexual selection
57 (Ritchie 2007), including coevolutionary and antagonistic effects, or by divergent ecological selection
58 acting directly on traits or preferences arising from environmental factors that influence signal or
59 preference efficiency, biotic interactions between species with similar signals (reproductive
60 interference) or reinforcing selection between hybridising species. Sexual isolation may result from
61 mutation order effects in similar environments (Mendelson et al. 2014), and divergence could also
62 be initiated by genetic drift (Uyeda et al. 2009).

63 A key factor influencing how divergence of mate recognition systems can lead to speciation is the
64 extent to which both signal and preference evolve together within diverging populations but also
65 jointly diverge sufficiently to generate assortative mating between populations (Ryan and Rand
66 1993; Rodriguez et al. 2013). Coordination between changes in signals and preferences is critical.
67 Mutual coevolutionary processes such as Fisher’s Runaway or some forms of antagonistic
68 coevolution may maintain correlation between male and female traits in a stepwise manner. If one
69 trait diverges first, for example through a change in environmental selection on a male signal or the
70 spread of a mutation of large effect on the preference, then the other trait will need to “catch up”
71 via trait modification to maintain coordination. This is potentially a slow process, especially if new
72 mutations are required rather than standing genetic variants being available.

73 Genetic architecture and coevolution

74 The genetic architecture of signals and preferences may influence the likelihood of coevolution
75 between them. Felsenstein’s seminal work (Felsenstein 1981) highlighted the issue of recombination
76 in speciation (see also Butlin et al. 2021). If genetically independent but interacting traits each have a
77 simple genetic control, tight physical linkage of the loci can facilitate coevolution by maintaining
78 association (linkage disequilibrium) between complementary allelic variants. Speciation often
79 involves ecological adaptation, in which case signals and preferences may also need to be associated
80 with locally adapted alleles. The necessary linkage disequilibrium (LD) can be generated in multiple
81 ways (by mutation, drift, gene flow or selection, Charlesworth and Charlesworth (2010), chapter 8)
82 but assortative mating is a powerful force generating associations. In whatever way it is generated,
83 LD is broken down by recombination. Therefore, close physical linkage can play a key role in
84 maintaining associations among alleles at different loci (Kirkpatrick 1982, Servedio and Burger 2018).
85 Factors reducing recombination among relevant genes can be favoured in such circumstances, by

86 inversions or other mechanisms, facilitating the appearance of coadapted gene complexes
87 influencing multiple traits (Ravinet et al. 2017; see chapter by Berdan et al. this volume).

88 "Genetic Coupling" potentially greatly facilitates coordinated changes in signals and preferences. In
89 the 1960s Alexander proposed the concept of "genetic coupling" of traits and preferences
90 (Alexander 1962). In this original idea, genetic coupling essentially meant genetic pleiotropy, i.e.
91 substitution of one allele resulting in changes in multiple traits. The way the term was used
92 specifically suggested that a new mutation could simultaneously influence both traits in a
93 complementary manner such that substitutions of large effect would simultaneously alter the
94 expression of both signals and preferences. This was expected to maintain some coordination and
95 reduce the selection that would otherwise oppose new variant signals or preferences (Doherty and
96 Hoy 1985; Ronacher 2019). Theoretically such variants could clearly influence rapid coevolution of
97 these traits: a single mutation could alter both signal and preference, and recombination would not
98 break the association between traits. Initial studies of interspecific hybrids, usually in acoustic
99 communication systems, were thought to support the idea (e.g. Hoy et al. (1977)), but most were of
100 low genetic resolution and did not provide conclusive evidence. In 1989, we (Butlin and Ritchie 1989)
101 reviewed the evidence proposed to support the genetic coupling hypothesis and concluded that, at
102 the time, there was no single convincing example. We also advanced several arguments to suggest
103 that coupling was unlikely. Boake (1991) reached broadly similar conclusions. In nearly 35
104 intervening years, methods of genomic mapping of complex traits have been revolutionised and
105 gene manipulation studies are becoming increasingly adept at examining (and illuminating)
106 pleiotropy. Several studies have hinted at new evidence supporting genetic coupling and at least two
107 have claimed positive support for genetic coupling. Here we provide an update of Butlin & Ritchie
108 (1989). We discuss important changes in the concept of genetic coupling and the improved
109 resolution of more recent studies.

110 What is "Coupling"?

111 Since the 1960s, the concept of "Coupling" in reproductive isolation has developed and now often
112 refers to the interaction between multiple barriers that jointly contribute to reproductive isolation
113 (Smadja and Butlin 2011; Butlin and Smadja 2018). If different barriers become coupled, in the sense
114 that they operate together to reduce gene flow between a pair of populations, then the overall
115 isolation may become more effective. Under this broad view, genetic and demographic factors, as
116 well as natural and sexual selection, can influence the build-up of coupling. However, there is no
117 general agreement on usage of the term: different perspectives and their implications for the role
118 and extent of coupling are discussed in detail in Dopman et al. (this volume), as well as the possible
119 roles of genetic linkage and pleiotropy in promoting coupling between barrier effects. "Genetic
120 coupling", as used in the original literature, refers to the particular case where pleiotropy leads to
121 coordinated effects of a locus on both signals and responses, potentially leading to assortative
122 mating. We refer to this as "Narrow-sense genetic coupling" and Table 1 places it in the context of
123 other relevant terms and concepts (which are not mutually exclusive). "Narrow sense" Genetic
124 Coupling concerns the evolution of a single barrier effect, because a barrier to gene flow exists only
125 when signal and preference both diverge, and so falls outside the scope of the coupling of separate
126 barrier effects, such as assortative mating and reduced hybrid fitness (Butlin and Smadja 2018,
127 Dopman et al. this volume, Perspective 3). However, "Broad-sense genetic coupling", which relies on
128 linkage between signal and preference genes rather than pleiotropy (Table 1), may be considered
129 part of the general coupling process under Perspectives 1 and 2 of Dopman et al. (this volume).
130 Narrow sense genetic coupling is related and relevant to the idea of "magic traits" (Gavrilets 2004;
131 Servedio et al. 2011) where a single gene (pleiotropy) or a single trait ("multiple effect" trait; Smadja

132 and Butlin 2011) influences more than one component of reproductive isolation. These mechanisms
 133 also have similarities to the case of “one allele” models of speciation (Felsenstein 1981), because
 134 they all circumvent the problem of recombination opposing speciation by breaking down allelic
 135 associations required for reproductive isolation. Finally, another relevant concept is the idea of
 136 matching traits (Kopp et al. 2018), where there are no separate signal and response traits but rather
 137 assortative mating depends on similarity between males and females for a single shared trait, for
 138 example size-assortative mating.

139

Concept	Description
Coupling I	The build-up of linkage disequilibrium among loci underlying barriers to gene exchange (Dopman et al., this volume)
Coupling II	The build-up of genome-wide linkage disequilibrium (Dopman et al., this volume)
Coupling III	The process generating a coincidence of distinct barrier effects (Dopman et al., this volume)
Broad-sense genetic coupling	A genetic association between mating signals and preferences due to tight linkage or recombination suppression (this paper).
Narrow-sense genetic coupling	A genetic association between mating signals and preferences due to pleiotropy, i.e. influence of the same allele on both traits (Alexander 1962, Butlin and Ritchie 1989, this paper)
Magic or multiple-effect trait	A single trait that influences more than one component of reproductive isolation, such as effects on both divergent adaptation and assortative mating (for example, Batesian warning colours also contributing to mate choice). The two terms are not directly equivalent (Servedio et al. 2011, Smadja and Butlin 2011, Dopman et al., this volume).
Single trait process	Divergent ecological selection on one trait indirectly leads to assortative mating (for example nest site preferences in some birds or fish) (Rice and Hostert, 1993, developed this, but called it the ‘single variation model’).
Single gene process	Divergence at a single locus results in one or more barriers to gene flow between populations (opposed by gene flow but not by recombination)
One-allele process	Substitution of the same allele, or evolution of a polygenic trait in the same direction, in two populations causes reproductive isolation between them, (for example the evolution of increased female choosiness) (Felsenstein 1981)
Matching trait assortment	Sexual isolation arising due to assortative mating based on phenotypes expressed in both sexes, e.g. assortative mating by body size (Kopp et al. 2018).

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141 Table 1. Coupling and related concepts in speciation. These ideas all have in common the reduction
 142 or removal of the opposing effect of recombination on the build-up of reproductive isolation.

143

144 Arguments have been made in favour of all of these potential mechanisms. One possible case of a
145 one-allele effect, influencing the level of choosiness, was identified in *Drosophila*, though the locus
146 has not been characterised in detail (Ortiz-Barrientos and Noor 2005). Other possible examples have
147 been suggested, especially the evolution of mate or habitat choice based on imprinting (Butlin et al.
148 2021). Servedio et al. (2011) and Smadja and Butlin (2011) argue that magic and multiple-effect
149 traits might be common, frequently suggested potential examples including chirality in snails
150 (though empirical studies suggest this is unlikely to be a really effective single-trait barrier, Richards
151 et al. 2017), host choice in phytophagous insects (e.g. *Rhagoletis* fruit flies; Tait et al. (2016),
152 *Acyrtosiphon* pea aphids; Hawthorne and Via (2001)) and wing patterns in *Heliconius* butterflies.
153 *Heliconius* wing patterns experience divergent selection due to mimicry but are also involved in
154 assortative mating, at least as signals (Kronforst et al. 2006; Merrill et al. 2019). Most examples are
155 of this type, only explaining divergence in one component of the communication system, either
156 signal or preference, being under ecological selection. The other trait also must diverge to generate
157 assortative mating. Reproductive isolation involving matching traits might also be common (Kopp et
158 al. 2018). Assortative mating for body size is a case in point, as exemplified by stickleback fish
159 (Ólafsdóttir et al. 2006).

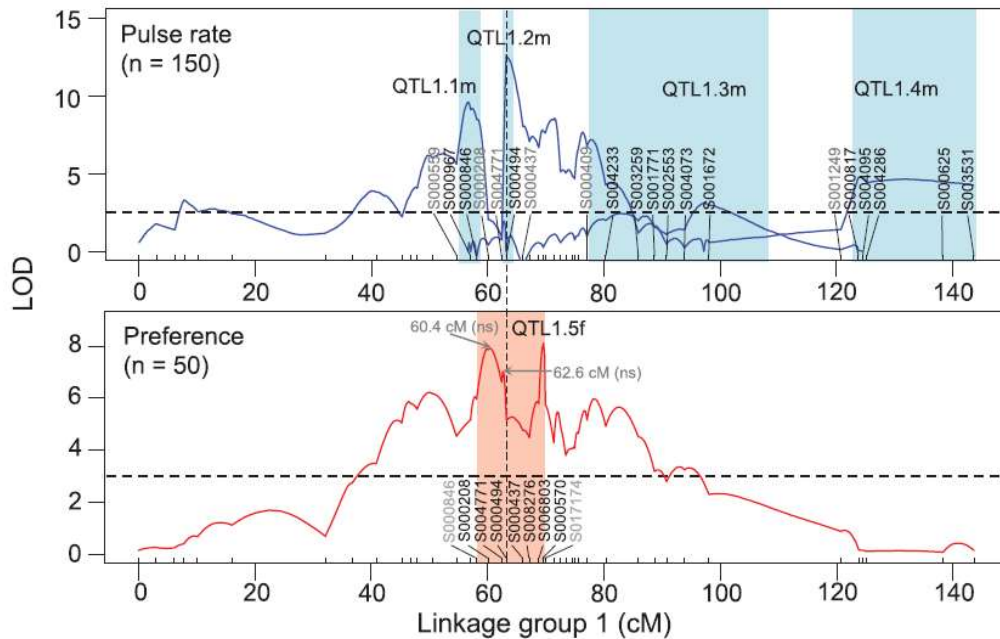
160 A one-allele system relies on the fixation of the same allele in different populations bringing about
161 isolation between them. An alternative simple mechanism is where one gene substitution (of a
162 different allele) would bring isolation due to matching effects on males and females leading to
163 isolation Perhaps the clearest example of a potential single-gene effect in the literature is the *period*
164 gene in *Drosophila*. This is a clock gene with extensive pleiotropic effects on most behaviours that
165 involve intrinsic rhythmicity. (Tauber et al. 2003) completed a genetic transformation experiment,
166 introducing the *period* gene of *D. pseudoobscura* into *D. melanogaster* and showing its effect on
167 altered diurnal activity cycle differences could lead to assortative mating between flies with the
168 same *period* allele and hence rhythmicity. This is a fascinating “proof of concept” study illustrating
169 that single gene effects on pre-mating isolation (in this case allochronic isolation, a type of ‘matching
170 trait assortment’) are possible, though the fact that the two species involved are only distantly
171 related perhaps questions if this is a good example of a direct role in speciation. The *period* gene
172 may also contribute to an oligogenic determination of tidal isolation between closely related marine
173 midges (Briševac et al. 2023). If further studies confirm a key role of this gene in this timing
174 difference, it could be a key demonstration of one gene influencing allochronic speciation in animals.

175 Narrow-sense genetic coupling is different from this single-gene, single-trait mechanism in that it
176 proposes a pleiotropic effect of one gene on two traits, a signal and an associated preference. Butlin
177 & Ritchie (1989) were unconvinced by any of the proposed examples of genetic coupling available at
178 the time. What would constitute such an example, and is it feasible? Narrow sense genetic coupling
179 implies that a mutation has a complementary effect on signalling and preference, that essentially the
180 traits are controlled by the “same gene” (Shaw and Lesnick 2009). At first glance, this may seem
181 unlikely because signals and preferences are superficially very different traits. However, in theory
182 this is possible if there is some underlying biochemical, physiological or developmental link. For
183 example, early work invoked the possibility of a single oscillator or common neurons underlying both
184 the frequency of an acoustic signal and the frequency-sensitivity of a receptor (Hoy et al. 1977). An
185 opsin gene in a fish may be under selection due to differences in colour propagation in water that
186 influence both colour production and detection of that colour. This example is only partly
187 hypothetical (Terai et al. 2006): opsins influence colour perception but are not known to influence
188 colour production directly. Some colour mutants in medaka seem to influence assortative mating,
189 although the mechanism is not clear (Fukamachi et al. 2009). The alternative to coupling would be
190 that loci other than opsins produce matching changes in colour production that are more easily

191 perceived due to selected changes in opsins. The changes may still be tightly coordinated by
192 selection but not due to a direct simultaneous effect of a single mutation on both trait and
193 preference. Narrow-sense genetic coupling, i.e. genetic coupling as originally proposed, requires
194 direct pleiotropic effects on signal and preference. Genetic associations due to tight linkage, co-
195 localization in an inversion or very strong epistatic selection are not the same. However, they may be
196 more feasible routes to rapid coevolution and have been considered as genetic coupling in much
197 recent literature, implying only genetic linkage or genomic clustering and not necessarily pleiotropy.
198 (Mead and Arnold 2004) used the term to include a genetic association between signal and
199 preference generated by assortative mating, without reference to its genetic basis, but more often
200 the term is invoked when genetic linkage between traits is tight. We will refer to this mechanism as
201 “broad-sense genetic coupling”.

202 The evidence.

203 Genetic mapping of traits, using either traditional quantitative trait locus (QTL) mapping or genomic
204 association studies, is now much more feasible and cost-effective in a wide range of organisms.
205 Mapping of signal traits and preferences has been completed in a number of systems, and tantalising
206 examples of co-localization have been found. Perhaps one of the most thoroughly studied cases is
207 QTLs for song pulse rate in Hawaiian crickets and, following an ingenious experimental design,
208 female preferences for these songs in crosses between *Laupala kohalensis* and *L. paranigra*. Initial
209 studies identified common QTL peaks affecting both traits (Shaw and Lesnick 2009), though these
210 were broad and contained multiple loci. Xu and Shaw (2021) refined the mapping of the signal and
211 preference loci and interval mapping showed that they clearly overlap, with ~3 cM difference
212 between peaks (Figure 1). Xu and Shaw (2019) also examined different QTL on another linkage
213 group. This chromosome also carried extremely tightly linked loci affecting song and preference,
214 with estimates of recombination distances between them of around only 0.06 cM. These studies
215 are an elegant demonstration of genetic linkage in the sense of a very tight, almost intimate, genetic
216 association. It is particularly striking how tightly these pairs of major loci co-localize on two different
217 chromosomes since both traits are polygenic, and each QTL explains ~10% of the species difference.
218 Whether these different tight peaks include genes with pleiotropic effects (i.e. narrow-sense genetic
219 coupling), and so the potential for a common influence on songs and preferences, remains to be
220 established, though annotations suggest neural functions and potential pattern generator genes lie
221 within the peaks (see also (Xu and Shaw 2020)). Pleiotropy in the case of polygenic traits is most
222 likely where there is some common underlying functional connection, such as an oscillator.



223

Figure 1. QTL mapping of loci influencing song (pulse rate) and preferences for pulse rate in crosses between the Hawaiian cricket species *Laupala kohalensis* and *L. paranigra*. Shaded areas indicate ranges of mapped QTL locations and some QTLs are named (e.g. QTL1.1m; 1st on linkage group 1 in males, etc) From Xu & Shaw (2019). (Inset, *Laupala kohalensis*, photo Kerry Shaw)

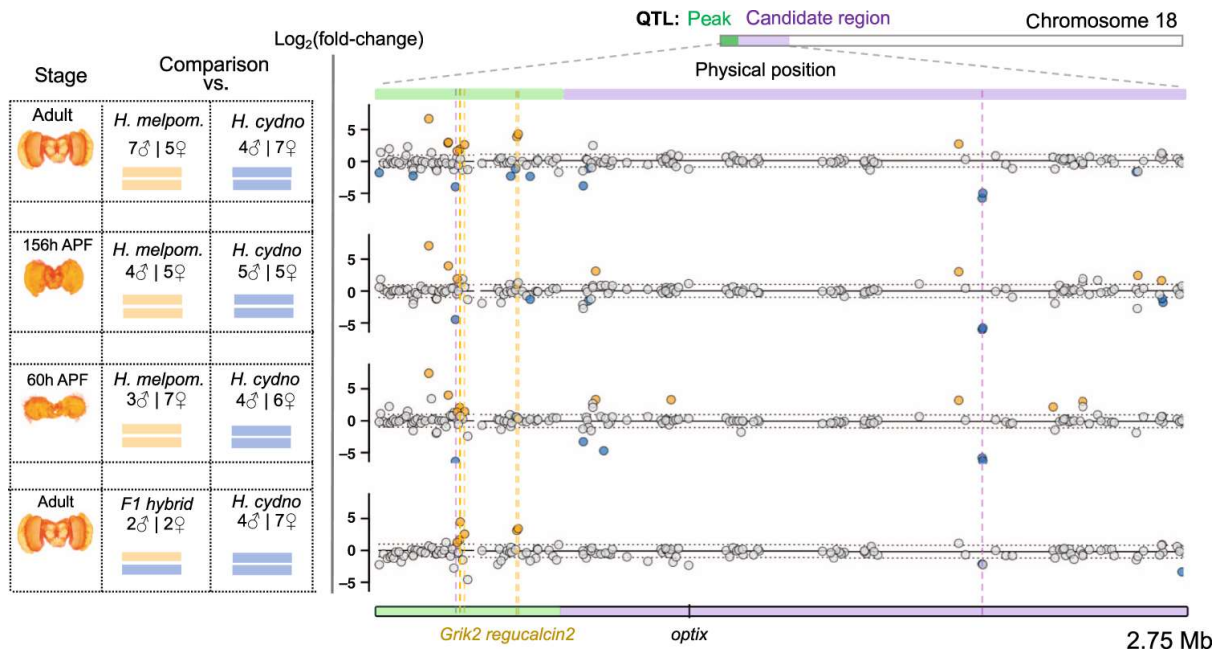


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226 Another system producing apparently compelling examples of co-localisation is provided by the
 227 warning colours of *Heliconius* butterflies, which are also used as mating cues by males (they are
 228 “magic” or “multiple-effect traits”). The question is whether preference genes co-localize with the
 229 pattern genes. Kronforst et al. (2006) identified QTLs for colour differences between *H. cydno* and *H.*
 230 *pachinus* and preferences for these patterns and found a “perfect” association between them,
 231 potentially implicating pleiotropic effects of the *wingless* gene and not consistent with tight linkage,
 232 unless the region also lacked recombination due to an unknown inversion. They suggested that
 233 genetic coupling, while unlikely in most systems, could reflect pleiotropy here if the pigments
 234 influencing wing colour are also expressed in the eyes and influence spectral sensitivity, a fascinating
 235 mechanism if true. Colour pattern differences between *Heliconius cydno* and *H. melpomene* and
 236 their perception have also been studied. Traits and preferences are genetically clustered (Merrill et
 237 al. 2011) but detailed association mapping resolves this to a number of male choice loci, some of
 238 which are tightly linked (~1cM) with the colour pattern gene *optix* (Merrill et al. 2019) (Rossi et al.
 239 2020). Detailed analyses of hybrids, including expression analyses of genes associated with choice,
 240 reveal that the initial larger QTL breaks down to tight linkage between loci, *optix* for colour pattern
 241 and others for male mate choice (Figure 2). QTL for sex pheromones, which influence female mate
 242 choice, are also clustered in the genome and may be loosely linked with some of the colour
 243 patterning loci, which may mean the two signalling traits have the potential for coordinated change

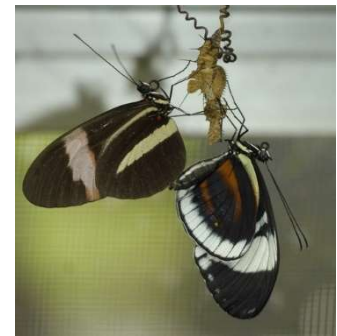
244 (Byers et al. 2021) but do not make as compelling a case for pleiotropy as the (Kronforst et al. 2006)
 245 study.

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Figure 2. Differential expression analyses of brain samples from the butterflies *Heliconius melpomene* and *H. cydno* and F1 hybrids identified candidate genes involved in pattern choice (*Grik2* and *regucalcin2*) are very tightly linked to *optix*, which controls wing pattern differences. From (Rossi, Haussmann et. al. 2020) Vertical lines highlight genes with differential expression between species (see paper for further details). (Inset; *Heliconius melpomene rosina* male x *H. cydno chionus* female, photo Richard Merrill).



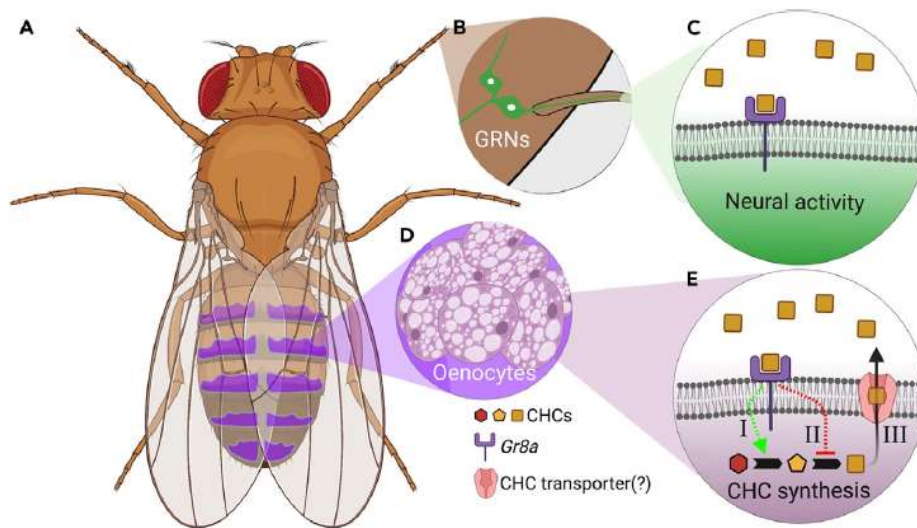
254 In some cases, assortative mating is studied directly, rather than the underlying signals and
 255 preference, which may not be known. Usually studies of assortative mating as a phenotype have led
 256 to the conclusion that this has, perhaps unsurprisingly, a polygenic basis with numerous small effect
 257 loci. This would seem to make coupling unlikely (Ting et al. 2001; Civetta and Cantor 2003).
 258 However, such loci may be more densely distributed on some chromosomes (e.g. sex chromosomes
 259 (Qvarnstrom and Bailey 2009; Abbott et al. 2017)) or co-inherited due to reduced recombination, or
 260 multiple loci might underlie a single trait that drives assortment through phenotype matching. Some
 261 studies have inferred genetic associations between traits involved, or components of assortative
 262 mating itself. An intriguing study investigated size-based assortative mating between limnetic and
 263 benthic stickleback morphs. This assortment is based on differences in morphology between the
 264 morphs, which are clearly multichromosomal and polygenic. However, (Bay et al. 2017) identified
 265 two QTLs for size-based mate choice which co-localised with some QTLs affecting body shape. Model
 266 fitting demonstrated that variation in choice QTLs explained a significant component of shape,
 267 suggesting a strong genetic correlation between the traits. Interestingly, the females' own shape was
 268 correlated with mate choice so phenotypic matching could be involved (Kopp et al. 2018). It would
 269 be interesting to see this examined in the medaka example mentioned above, another case where
 270 phenotype matching could influence assortative mating (Fukamachi, Kinoshita et al. 2009)

271 Using introgression experiments between *D. simulans* and *D. mauritiana*, McNiven and Moehring
272 (2013) identified two small chromosomal regions that contained genes influencing both male and
273 female components of assortative mating (preferences and unidentified factors influencing male
274 species-specific attractiveness), suggesting close linkage or pleiotropy of these unidentified loci. In a
275 study of *D. simulans* and *D. melanogaster*, deficiency mapping had identified a region including the
276 transcription factor *fruitless* and Chowdhury et al. (2020) then used mutants to demonstrate that
277 this locus influences female species-specific preference. This is extremely interesting: *fruitless* is
278 studied extensively because of its dramatic effects on male courtship behaviour, including the
279 production of courtship song (Rideout et al. 2007; Clyne and Miesenböck 2008). By alternative
280 splicing, the *fruitless* gene produces a number of different transcripts that may be male-specific,
281 female-specific or common to both sexes, with the potential to influence courtship song amongst
282 other, usually sex-specific traits. Transcripts vary in exon structure and content and arise from
283 different promoter regions and splicing variants (Parker et al. 2014). Mutations in male-specific
284 transcripts of *fruitless* cause males to sing aberrant song or no song at all (Neville et al. 2014). The
285 fact that *fruitless* can influence preference is tantalising, though the manner in which *fruitless*
286 influences female behaviour is not known: it seems to involve one of the non-sex-specific transcripts
287 produced by a different promoter region from that producing male-specific transcripts that establish
288 male courtship behaviours. Intriguingly, this transcript does not seem to influence female receptivity
289 to male song. This example nicely illustrates how the same gene might underlie signal and
290 preference without the effect of any one allelic substitution acting pleiotropically. It is strictly-
291 speaking an example of tight linkage rather than narrow-sense genetic coupling.

292 Another example in *Drosophila* that has been genetically dissected with great precision provides one
293 of the most compelling cases yet for pleiotropy. Major contributors to assortative mating in the
294 melanogaster group of *Drosophila* are cuticular hydrocarbons, and loci involved in their production
295 and perception are well characterised. The desaturase gene family has been shown to control
296 important changes in species-specific pheromonal components of CHCs (Jallon and Wicker-Thomas
297 2003; Wicker-Thomas 2007). Very elegant work has further shown how they can alter sex-specific
298 CHC expression and change the ratio of key compounds, often produced by females and detected by
299 males (Shirangi et al. 2009; Fang et al. 2009). Marcillac et al. (2005) took a candidate gene approach
300 and mutated a key enzyme, *desat1*, using transposon insertion. Pheromone production was altered
301 in both sexes, especially key sex pheromones. Males carrying the mutation were unable to
302 distinguish the sexes, at least in the dark when pheromones are essential for sex recognition.
303 Transposon excision affected these traits differently, perhaps suggesting the precise molecular
304 mechanism for production and detection differed between the sexes. This was explored further by
305 Bousquet et al. (2012) in a very elegant study. They demonstrated that *desat1* has 5 transcripts, each
306 with the same protein but different regulatory regions. Its potential pleiotropic role was examined
307 by generating mutants and reporters for each transcript. One promoter was strongly associated with
308 changes in the production of saturated/desaturated CHCs and this was expressed in female
309 oenocytes. Another was associated with male sex-discrimination, and was expressed in the antennal
310 lobes and a sexually dimorphic glomerulus. Clearly the development of sex- and tissue-specific
311 transcripts may well be a key to diversifying gene function in sexual communication systems and,
312 unlike *fruitless*, this effect is primarily regulatory rather than involving both coding and regulatory
313 variation. However, like *fruitless*, it may also be the case here that no single mutation influences
314 signals and preferences in a coordinated fashion.

315 The desaturase gene family is known for its role in fatty acid production, including the cuticular
316 hydrocarbons, and these studies suggest it also influences perception. Remarkably, a recent study
317 provides a very similar interpretation of potential genetic coupling in *Drosophila* pheromones, but in

318 this case a gustatory receptor (Gr) locus known to influence chemical perception has been
 319 demonstrated to influence pheromone production as well. Vernier et al. (2023) measured tissue-
 320 specific expression of members of the Gr family across several tissues in *Drosophila* and identified
 321 that *Gr8a* is expressed in both sensory tissues and oenocytes. In males it is also expressed in the
 322 ejaculatory bulb and is suggested to be involved in the production of an inhibitory compound
 323 (“antiaphrodisiacs” are passed in the ejaculate to alter female attractiveness to rivals; Billeter and
 324 Levine (2015)). Knockdown males produce altered CHCs, though not known antiaphrodisiacs such as
 325 CVA. Knockdown females have altered mating behaviour, mating more quickly, and knockdown
 326 males are both more sexually attractive and less likely to discriminate against mated females. The
 327 authors’ interpretation is that *Gr8a* is involved in regulating the behavioural responses to an
 328 inhibitory mating pheromone in females and males, and its production in males. Knockdown is a
 329 valuable method to demonstrate the role of a gene product in multiple functions but it cannot
 330 distinguish strict pleiotropy from the effects of different substitutions in the same gene.



331
 332 **Figure 3.** Model for the pleiotropic effect of *Gr8a* in both production and perception of pheromones.
 333 (A). Male fly, with oenocytes in magenta (B) *Gr8a*-expressing GRNs and in the sensory terminal tarsi
 334 (C) *Gr8a* functions as an inhibitory pheromone receptor in the tarsi (D) Oenocytes produce the
 335 pheromones in the abdomen (E) *Gr8a* functions as an autoreceptor in oenocytes, which regulates
 336 synthesis (I-II) and secretion (III) via feedback loops. From Vernier et al. (2023)

337 Other studies of pheromonal communication systems have disentangled the genetics of signal-
 338 receiver systems very clearly. Moths are ideal systems for such work and an important study system
 339 is the European corn borer, *Ostrinia nubilalis*. This system has two strains that show assortative
 340 mating due to alternate (E/Z) volatile long-range pheromone blends and has been examined for
 341 many years and in many localities. The pheromone polymorphism is mainly influenced by alleles at
 342 the *pgFAR* locus, coding for a fatty acyl reductase enzyme (Lassance et al. 2013). Pheromone
 343 discrimination was assumed to be due to candidate receptor loci expressed on the antenna, but
 344 recently very neat work (Unbehend et al. 2021) demonstrated the key locus responsible was *bric-a-*
 345 *brac* (*bab*), a gene including a BTB domain known to be involved in morphological pattern generation
 346 in *Drosophila*. Association analyses and CRISPR knockout demonstrated that the key difference
 347 influencing pheromone discrimination in *Ostrinia* lies in a regulatory intron rather than any exon.
 348 *pgFAR* and *bab* are on different chromosomes but a key finding from this work is that, in natural

349 populations, *pgFAR* and intron1 of *bab* are in strong disequilibrium and show heterozygote deficit
350 due to assortative mating. Hence, this is a very nice example of the ability of strong assortative
351 mating to generate linkage disequilibrium between distinct, unlinked loci affecting signal and
352 preference. If assortative mating, perhaps combined with epistatic effects on fitness, can maintain
353 associations even among unlinked loci, then perhaps genetic coupling is not important for the
354 coevolution of signals and preferences. It is fascinating that *bab* has also been implicated in male UV
355 signalling in *Colias* butterflies (Ficarrotta et al. 2022). Here the Z chromosome, in addition to carrying
356 *bab*, also influences female preference, which is genetically associated with signalling in a hybrid
357 zone between *C. eurytheme* and *C. philodice*, but it is not yet known whether this could be due to
358 pleiotropy.

359 Whither “genetic coupling”?

360 This is a short and probably not exhaustive review of potential cases of genetic coupling in sexual
361 signalling systems. It is striking that in the last few decades there have still only been two or three
362 potential examples of genetic coupling at the level of key loci that underlie both signal and
363 preference. So far, there is no system where the same allelic substitution has been shown to
364 influence both traits, although that also cannot be ruled out in some cases. Therefore, narrow-sense
365 genetic coupling, i.e. coupling due to pleiotropy rather than linkage, has not been demonstrated.
366 Close linkage has been found, either within the same gene or in nearby genes, and this extends to
367 cases where multiple loci influence both traits, suggesting some underlying common mechanistic
368 connection: evidence for broad-sense genetic coupling is slowly accumulating.

369 Perhaps further consideration suggests that the original idea of a single mutation with coordinated
370 effects was naïve and always unlikely. The idea of such a “magic mutation” with pleiotropic effects
371 inducing changes in signal and preference is tantalising, but what is the likelihood that a mutation in
372 such a gene would have a coordinated effect on both traits? Would the shift in signal necessarily
373 match the shift in preference when the effects on some common stage in a biochemical or
374 developmental pathway have been filtered through later steps to reach the different phenotypes?
375 The *period* gene of *D melanogaster*, part of the clock mechanism, influences both song and female
376 detection of song patterns, but not in a matching manner. Mutations in *per* do both influence song
377 rhythm and disrupt preference but there is no matching of the effects of mutations such that *perL*
378 lengthens rhythm in males and makes females prefer long rhythms (though the mutations studied to
379 date are induced mutations rather than natural alleles) (Greenacre et al. 1993). The gene does
380 influence a common pattern generator and so connects changes in song and recognition, but
381 coordinated mutational effects seem unlikely. The advantage of requiring a single mutation may
382 then be lost, because additional modifier mutations are still needed to maintain signal-response
383 coevolution.

384 *Fruitless* and *desat1* probably indicate more convincing ways that one complex gene can influence
385 sex- and tissue-specific functions, with multiple transcripts being able to diverge in function in
386 different tissues involved in signal production or recognition such as oenocytes and receptors (Figure
387 3). Hence coordination is probably not due to a single mutation, though the alternative transcripts
388 will be intimately linked. However, this would probably still require step-for-step changes in gene
389 function: a mutation influencing one function has to be able to increase in frequency first and then
390 generate selection for a corresponding change in the other function. This might limit evolution to
391 small steps and increase the waiting time for suitable complementary mutations. In *desat1* (and *bab*)
392 it is primarily promoter sequence divergence controlling where and how the gene is expressed
393 (though details of how this influences coordination are not yet known). In *fruitless* there are
394 important changes in sex-specific promoter regions, but also coding sequence divergence. (Parker et

395 al. 2014) showed how most exons of *fruitless* exhibit strong evidence for purifying selection, as
396 might be expected for a key gene in the sex determination pathway with conserved gene function
397 across Diptera, but that divergent selection was concentrated in one sex-specific exon across
398 multiple species. Clearly identifying the mutational steps involved in adaptation and the order of
399 their appearance during evolutionary change is a major challenge but is necessary to disentangle
400 these questions and explore the importance of intralocus epistasis, i.e. interactions between
401 different exons or transcripts from a single gene, or coding and regulatory divergence. Some
402 progress is being made in identifying key mutational steps in other systems (Chan et al. 2010).
403 CRISPR provides a fantastic opportunity to examine the role of specific mutations on multiple traits
404 and recapitulate such steps (Karageorgi et al. 2019).

405 So what?

406 As discussed earlier and elsewhere (Dopman et al., this volume), the concept of genetic coupling has
407 evolved and loosened. Here we have used narrow-sense genetic coupling to suggest strict
408 pleiotropy. Broad-sense genetic coupling, we suggest, includes tight linkage, either within complex
409 genes or between tightly-linked loci. There are increasing numbers of examples of such broad-sense
410 genetic coupling in differences between recognition systems of sibling species, most notably *Laupala*
411 and *Heliconius*, as well as the within-locus cases in *Drosophila*. Similarly, genomic analysis is
412 highlighting the importance of reduced recombination, inversions and “supergenes” in ecological
413 adaptation and speciation (Ravinet et al. 2017; Faria et al. 2019; Berdan et al. 2022). Could such
414 linkage facilitate coordinated change just as effectively as pleiotropy? The corn borer example shows
415 that strong assortative mating can generate linkage disequilibrium even among unlinked loci. Theory
416 suggests that, at least in some circumstances, tight linkage is not just unnecessary for the
417 maintenance of linkage disequilibrium but might actually impede the evolution of reproductive
418 isolation. An intriguing recent study (Servedio and Burger 2018) modelled ecological speciation
419 based on three loci, for an adaptation, a trait and choosiness, in the context of assessing the
420 likelihood of “magic” versus “pseudomagic” traits (i.e. pleiotropic versus linked traits, in this context
421 between an ecological adaptation and a male signalling trait). At least in a secondary contact
422 scenario, recombination could facilitate the evolution of greater choosiness, though the role of the
423 ecological adaptation is important. Whether a similar effect might occur in a non-ecological context
424 is unclear. Similarly counterintuitive effects of recombination can occur when multiple signal traits or
425 mutual mate choice are involved (Aubier et al. 2019; Aubier Chapter, this volume). Indeed, it is
426 possible that pleiotropy could actually be less effective than linkage if distinct but associated loci
427 allowed molecular coevolution to occur more quickly than an ‘unbalanced’ mutation in a single gene.
428 Strict pleiotropy may prevent more favourable combinations of alleles or mutations from becoming
429 associated by recombination, so the search for pleiotropy in mate recognition systems may
430 ultimately be a red herring. We conclude that there is still much to be learned about the genetic
431 bases of signals and preferences, and the co-evolutionary relationships between them, but that
432 there is little reason to propose that strict pleiotropy is likely, or would necessarily facilitate rapid co-
433 evolution.

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439 References

- 440 Abbott JK, Norden AK, Hansson B. 2017. Sex chromosome evolution: historical insights and future
 441 perspectives. *Proceedings of the Royal Society B-Biological Sciences* **284**. 20162806.
 442 Alexander RD. 1962. Evolutionary change in cricket acoustical communication. *Evolution* **16**: 433-
 443 467.
 444 Aubier TG, Kokko H, Joron M. 2019. Coevolution of male and female mate choice can destabilize
 445 reproductive isolation. *Nature Communications* **10**. 5122.
 446 Bay RA, Arnegard ME, Conte GL, Best J, Bedford NL, McCann SR, Dubin ME, Chan YF, Jones FC,
 447 Kingsley DM et al. 2017. Genetic Coupling of Female Mate Choice with Polygenic Ecological
 448 Divergence Facilitates Stickleback Speciation. *Current Biology* **27**: 3344-3349.
 449 Berdan EL, Flatt T, Kozak GM, Lotterhos KE, Wielstra B. 2022. Genomic architecture of supergenes:
 450 connecting form and function. *Philosophical Transactions of the Royal Society B-Biological*
 451 *Sciences* **377**. 20210192.
 452 Billeter JC, Levine JD. 2015. The role of cVA and the Odorant binding protein Lush in social and sexual
 453 behavior in *Drosophila melanogaster*. *Front Ecol Evol* **3**. 75.
 454 Boake CRB. 1991. Coevolution of senders and receivers of sexual signals: genetic coupling and
 455 genetic coevolution. *Trends Ecol Evol* **6**: 225-227.
 456 Bousquet F, Nojima T, Houot B, Chauvel I, Chaudy S, Dupas S, Yamamoto D, Ferveur JF. 2012.
 457 Expression of a desaturase gene, *desat1*, in neural and nonneural tissues separately affects
 458 perception and emission of sex pheromones in *Drosophila*. *Proceedings of the National*
 459 *Academy of Sciences of the United States of America* **109**: 249-254.
 460 Briševac D, Peralta CM, Kaiser TS. 2023. An oligogenic architecture underlying ecological and
 461 reproductive divergence in sympatric populations. *eLife* **12**: e82825.
 462 Butlin RK, Ritchie MG. 1989. Genetic coupling in mate recognition systems: What is the evidence?
 463 *Biol J Linn Soc* **37**: 237-246.
 464 Butlin RK, Servedio MR, Smadja CM, Bank C, Barton NH, Flaxman SM, Giraud T, Hopkins R, Larson EL,
 465 Maan ME et al. 2021. Homage to Felsenstein 1981, or why are there so few/many species?
 466 *Evolution* **75**: 978-988.
 467 Butlin RK, Smadja CM. 2018. Coupling, Reinforcement, and Speciation. *American Naturalist* **191**: 155-
 468 172.
 469 Byers K, Darragh K, Garza SF, Almeida DA, Warren IA, Rastas PMA, Merrill RM, Schulz S, McMillan
 470 WO, Jiggins CD. 2021. Clustering of loci controlling species differences in male chemical
 471 bouquets of sympatric *Heliconius* butterflies. *Ecology and Evolution* **11**: 89-107.
 472 Chan YF, Marks ME, Jones FC, Villarreal G, Shapiro MD, Brady SD, Southwick AM, Absher DM,
 473 Grimwood J, Schmutz J et al. 2010. Adaptive Evolution of Pelvic Reduction in Sticklebacks by
 474 Recurrent Deletion of a *Pitx1* Enhancer. *Science* **327**: 302-305.
 475 Charlesworth BC, Charlesworth D. 2010. *Elements of Evolutionary Genetics*. Roberts, Greenwood
 476 Village.
 477 Chowdhury T, Calhoun RM, Bruch K, Moehring AJ. 2020. The *fruitless* gene affects female receptivity
 478 and species isolation. *Proceedings of the Royal Society B: Biological Sciences* **287**: 20192765.
 479 Civetta A, Cantor EJF. 2003. The genetics of mating recognition between *Drosophila simulans* and *D*-
 480 *sechellia*. *Genetical Research* **82**: 117-126.
 481 Clyne JD, Miesenböck G. 2008. Sex-Specific Control and Tuning of the Pattern Generator for
 482 Courtship Song in *Drosophila*. *Cell* **133**: 354-363.
 483 Doherty J, Hoy R. 1985. The auditory behaviour of crickets: some views of genetic coupling, song
 484 recognition and predator detection. *Quart Rev Biol* **60**: 457-472.
 485 Fang S, Ting CT, Lee CR, Chu KH, Wang CC, Tsaur SC. 2009. Molecular evolution and functional
 486 diversification of fatty acid desaturases after recurrent gene duplication in *Drosophila*.
 487 *Molecular Biology and Evolution* **26**: 1447-1456.
 488 Faria R, Johannesson K, Butlin RK, Westram AM. 2019. Evolving Inversions. *Trends in Ecology &*
 489 *Evolution* **34**: 239-248.

490 Felsenstein J. 1981. Skepticism towards Santa Rosalia, or why are there so few kinds of animals?
491 *Evolution* **35**: 124-138.

492 Ficarrotta V, Hanly JJ, Loh LS, Francescutti CM, Ren AN, Tunstrom K, Wheat CW, Porter AH,
493 Counterman BA, Martin A. 2022. A genetic switch for male UV iridescence in an incipient
494 species pair of sulphur butterflies. *Proceedings of the National Academy of Sciences of the*
495 *United States of America* **119**. e2109255118.

496 Fukamachi S, Kinoshita M, Aizawa K, Oda S, Meyer A, Mitani H. 2009. Dual control by a single gene of
497 secondary sexual characters and mating preferences in medaka. *Bmc Biology* **7**. 64.

498 Gavrillets S. 2004. *Fitness landscapes and the origin of species*. Princeton University Press.

499 Greenacre M, Ritchie MG, Byrne BC, Kyriacou CP. 1993. Female song preference and the *period* gene
500 of *Drosophila melanogaster*. *Behav Genet* **23**: 85-90.

501 Hawthorne DJ, Via S. 2001. Genetic linkage of ecological specialization and reproductive isolation in
502 pea aphids. *Nature* **412**: 904-907.

503 Hoy RR, Hahn J, Paul RC. 1977. Hybrid cricket auditory behavior – evidence for genetic coupling in
504 animal communication. *Science* **195**: 82-84.

505 Jallon J-M, Wicker-Thomas C. 2003. Genetic studies on pheromone production in *Drosophila*. in
506 *Insect Pheromone Biochemistry and Molecular Biology* (eds. GJ Blomqvist, RG Vogt), pp.
507 253-281. Elsevier.

508 Karageorgi M, Groen SC, Sumbul F, Pelaez JN, Verster KI, Aguilar JM, Hastings AP, Bernstein SL,
509 Matsunaga T, Astourian M et al. 2019. Genome editing retraces the evolution of toxin
510 resistance in the monarch butterfly. *Nature* **574**: 409-412.

511 Kirkpatrick M. 1982. Sexual selection and the evolution of female choice. *Evolution* **36**: 1-12.

512 Kopp M, Servedio MR, Mendelson TC, Safran RJ, Rodriguez RL, Hauber ME, Scordato EC, Symes LB,
513 Balakrishnan CN, Zonana DM et al. 2018. Mechanisms of Assortative Mating in Speciation
514 with Gene Flow: Connecting Theory and Empirical Research. *American Naturalist* **191**: 1-20.

515 Kronforst MR, Young LG, Kapan DD, McNeely C, O'Neill RJ, Gilbert LE. 2006. Linkage of butterfly mate
516 preference and wing color preference cue at the genomic location of wingless. *Proceedings*
517 *of the National Academy of Sciences of the United States of America* **103**: 6575-6580.

518 Lande R. 1981. Models of speciation by sexual selection on polygenic traits. *Proc Natl Acad Sci USA*
519 **78**: 3721-3725.

520 Lassance J-M, Liénard MA, Antony B, Qian S, Fujii T, Tabata J, Ishikawa Y, Löfstedt C. 2013. Functional
521 consequences of sequence variation in the pheromone biosynthetic gene pgFAR for *Ostrinia*
522 moths. *Proceedings of the National Academy of Sciences* **110**: 3967-3972.

523 Marcillac F, Grosjean Y, Ferveur JF. 2005. A single mutation alters production and discrimination of
524 *Drosophila* sex pheromones. *Proceedings of the Royal Society B-Biological Sciences* **272**: 303-
525 309.

526 McNiven VTK, Moehring AJ. 2013. Identification of genetically linked female preference and male
527 trait. *Evolution* **67**: 2155-2165.

528 Mead LS, Arnold SJ. 2004. Quantitative genetic models of sexual selection. *Trends in Ecology &*
529 *Evolution* **19**: 264-271.

530 Mendelson TC, Martin MD, Flaxman SM. 2014. Mutation-order divergence by sexual selection:
531 diversification of sexual signals in similar environments as a first step in speciation. *Ecology*
532 *Letters* **17**: 1053-1066.

533 Merrill RM, Rastas P, Martin SH, Melo MC, Barker S, Davey J, McMillan WO, Jiggins CD. 2019. Genetic
534 dissection of assortative mating behavior. *Plos Biology* **17**.

535 Merrill RM, Van Schooten B, Scott JA, Jiggins CD. 2011. Pervasive genetic associations between traits
536 causing reproductive isolation in *Heliconius* butterflies. *Proceedings of the Royal Society B-*
537 *Biological Sciences* **278**: 511-518.

538 Neville MC, Nojima T, Ashley E, Parker DJ, Walker J, Southall T, Van de Sande B, Marques AC, Fischer
539 B, Brand AH et al. 2014. Male-Specific Fruitless Isoforms Target Neurodevelopmental Genes
540 to Specify a Sexually Dimorphic Nervous System. *Current Biology* **24**: 229-241.

541 Ólafsdóttir GÁ, Ritchie MG, Snorrason SS. 2006. Positive assortative mating between recently
542 described sympatric morphs of Icelandic sticklebacks. *Biology Letters* **2**: 250-252.

543 Ortiz-Barrientos D, Noor MAF. 2005. Evidence for a one-allele assortative mating locus. *Science* **310**:
544 1467-1467.

545 Panhuis TM, Butlin R, Zuk M, Tregenza T. 2001. Sexual selection and speciation. *Trends Ecol Evol* **16**:
546 364-371.

547 Parker DJ, Gardiner A, Neville MC, Ritchie MG, Goodwin SF. 2014. The evolution of novelty in
548 conserved genes; evidence of positive selection in the *Drosophila fruitless* gene is localised
549 to alternatively spliced exons. *Heredity* **112**: 300-306.

550 Paterson HEH. 1978. More evidence against speciation by reinforcement. *S Afr J Sci* **74**: 369-371.

551 Qvarnstrom A, Bailey RI. 2009. Speciation through evolution of sex-linked genes. *Heredity* **102**: 4-15.

552 Ravinet M, Faria R, Butlin RK, Galindo J, Bierne N, Rafajlovic M, Noor MAF, Mehlig B, Westram AM.
553 2017. Interpreting the genomic landscape of speciation: a road map for finding barriers to
554 gene flow. *Journal of Evolutionary Biology* **30**: 1450-1477.

555 Rice WR, Hostert EE. 1993. Laboratory experiments on speciation: what have we learned in 40
556 years? *Evolution* **47**: 1637-1653.

557 Rideout EJ, Billeter JC, Goodwin SF. 2007. The sex-determination genes fruitless and doublesex
558 specify a neural substrate required for courtship song. *Current Biology* **17**: 1473-1478.

559 Richards PM, Morii Y, Kimura K, Hirano T, Chiba S, Davison A. 2017. Single-gene speciation: Mating
560 and gene flow between mirror-image snails. *Evolution Letters* **1**: 282-291.

561 Ritchie MG. 2007. Sexual selection and speciation. *Annual Review of Ecology Evolution and*
562 *Systematics* **38**: 79-102.

563 Rodriguez RL, Boughman JW, Gray DA, Hebets EA, Hoebel G, Symes LB. 2013. Diversification under
564 sexual selection: the relative roles of mate preference strength and the degree of divergence
565 in mate preferences. *Ecology Letters* **16**: 964-974.

566 Ronacher B. 2019. Innate releasing mechanisms and fixed action patterns: basic ethological concepts
567 as drivers for neuroethological studies on acoustic communication in Orthoptera. *Journal of*
568 *Comparative Physiology a-Neuroethology Sensory Neural and Behavioral Physiology* **205**: 33-
569 50.

570 Rossi M, Hausmann AE, Thurman TJ, Montgomery SH, Papa R, Jiggins CD, McMillan WO, Merrill RM.
571 2020. Visual mate preference evolution during butterfly speciation is linked to neural
572 processing genes. *Nature Communications* **11**. 4763.

573 Ryan MJ, Rand AS. 1993. Species recognition and sexual selection as a unitary problem in animal
574 communication. *Evolution* **47**: 647-657.

575 Servedio MR, Van Doorn GS, Kopp M, Frame AM, Nosil P. 2011. Magic traits in speciation: 'magic' but
576 not rare? *Trends in ecology & evolution* **26**: 389-397.

577 Shaw KL, Lesnick SC. 2009. Genomic linkage of male song and female acoustic preference QTL
578 underlying a rapid species radiation. *Proceedings of the National Academy of Sciences of the*
579 *United States of America* **106**: 9737-9742.

580 Shirangi TR, Dufour HsD, Williams TM, Carroll SB. 2009. Rapid evolution of sex pheromone-
581 producing enzyme expression in *Drosophila*. *PLoS Biol* **7**: e1000168.

582 Smadja CM, Butlin RK. 2011. A framework for comparing processes of speciation in the presence of
583 gene flow. *Molecular Ecology* **20**: 5123-5140.

584 Tait C, Batra S, Ramaswamy S, Feder JL, Olsson SB. 2016. Sensory specificity and speciation: a
585 potential neuronal pathway for host fruit odour discrimination in *Rhagoletis pomonella*.
586 *Proceedings of the Royal Society B-Biological Sciences* **283**. 20162101.

587 Tauber E, Roe H, Costa R, Hennessy JM, Kyriacou CP. 2003. Temporal mating isolation driven a
588 behavioral gene in *Drosophila*. *Current Biology* **13**: 140-145.

589 Terai Y, Seehausen O, Sasaki T, Takahashi K, Mizoiri S, Sugawara T, Sato T, Watanabe M, Konijnendijk
590 N, Mrosso HDJ et al. 2006. Divergent Selection on Opsins Drives Incipient Speciation in Lake
591 Victoria Cichlids. *PLoS Biology* **4**: e433.

592 Ting C-T, Takahashi A, C.-I. W. 2001. Incipient speciation by sexual isolation in *Drosophila*:
593 Concurrent evolution at multiple loci. *Proc Natl Acad Sci USA* **98**: 6709-6713.

594 Unbehend M, Kozak GM, Koutroumpa F, Coates BS, Dekker T, Groot AT, Heckel DG, Dopman EB.
595 2021. bric a brac controls sex pheromone choice by male European corn borer moths.
596 *Nature Communications* **12**: 11.

597 Uyeda JC, Arnold SJ, Hohenlohe PA, Mead LS. 2009. Drift promotes speciation by sexual selection.
598 *Evolution* **63**: 583-594.

599 Vernier CL, Leitner N, Zelle KM, Foltz M, Dutton S, Liang X, Halloran S, Millar JG, Ben-Shahar Y. 2023.
600 A pleiotropic chemoreceptor facilitates the production and perception of mating
601 pheromones. *iScience* **26**: 105882.

602 Wicker-Thomas C. 2007. Pheromonal communication involved in courtship behavior in Diptera.
603 *Journal of Insect Physiology* **53**: 1089-1100.

604 Xu MZ, Shaw KL. 2019. Genetic coupling of signal and preference facilitates sexual isolation during
605 rapid speciation. *Proceedings of the Royal Society B-Biological Sciences* **286**. 20191607.

606 Xu M, Shaw KL. 2020. The genetics of mating song evolution underlying rapid speciation: Linking
607 quantitative variation to candidate genes for behavioral vsolation. *Genetics* **215**: 285-286.

608 Xu MZ, Shaw KL. 2021. Extensive linkage and genetic coupling of song and preference loci underlying
609 rapid speciation in *Laupala* crickets. *Journal of Heredity* **112**: 204-213.

610

611