Check for updates

OPEN ACCESS

EDITED BY Niccolò Zampieri, Helmholtz Association of German Research Centers (HZ), Germany

REVIEWED BY Francesco Roselli, Ulm University, Germany

CORRESPONDENCE
 Ilary Allodi
 iallodi@sund.ku.dk;
 ia51@st-andrews.ac.uk

RECEIVED 19 April 2023 ACCEPTED 08 June 2023 PUBLISHED 04 July 2023

CITATION

Mora S and Allodi I (2023) Neural circuit and synaptic dysfunctions in ALS-FTD pathology. *Front. Neural Circuits* 17:1208876.

doi: 10.3389/fncir.2023.1208876

COPYRIGHT

© 2023 Mora and Allodi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Neural circuit and synaptic dysfunctions in ALS-FTD pathology

Santiago Mora¹ and Ilary Allodi^{1,2*}

¹Integrative Neuroscience Unit, Department of Neuroscience, Panum Institute, University of Copenhagen, Copenhagen, Denmark, ²Neural Circuits of Disease Laboratory, School of Psychology and Neuroscience, University of St Andrews, St Andrews, United Kingdom

Action selection is a capital feature of cognition that guides behavior in processes that range from motor patterns to executive functions. Here, the ongoing actions need to be monitored and adjusted in response to sensory stimuli to increase the chances of reaching the goal. As higher hierarchical processes, these functions rely on complex neural circuits, and connective loops found within the brain and the spinal cord. Successful execution of motor behaviors depends, first, on proper selection of actions, and second, on implementation of motor commands. Thus, pathological conditions crucially affecting the integrity and preservation of these circuits and their connectivity will heavily impact goal-oriented motor behaviors. Amyotrophic Lateral Sclerosis (ALS) and Frontotemporal Dementia (FTD) are two neurodegenerative disorders known to share disease etiology and pathophysiology. New evidence in the field of ALS-FTD has shown degeneration of specific neural circuits and alterations in synaptic connectivity, contributing to neuronal degeneration, which leads to the impairment of motor commands and executive functions. This evidence is based on studies performed on animal models of disease, post-mortem tissue, and patient derived stem cells. In the present work, we review the existing evidence supporting pathological loss of connectivity and selective impairment of neural circuits in ALS and FTD, two diseases which share strong genetic causes and impairment in motor and executive functions.

KEYWORDS

amyotrophic lateral sclerosis (ALS), frontotemporal dementia (FTD), synapses and neurons, motor control, cognitive functions

Introduction

Neurodegenerative diseases entail a major challenge as life expectancy increases in modern societies (Arthur et al., 2016). As one remarkable example, amyotrophic lateral sclerosis (ALS) is a fatal neurodegenerative disorder affecting motor neurons found in the spinal cord and brainstem, as well as corticospinal neurons. The disease is characterized by muscle wasting and progressive paralysis (Schweingruber and Hedlund, 2022). Frontotemporal dementia (FTD), instead, is a progressive condition characterized by the degeneration of neurons in the frontal and temporal lobes, leading to changes in behavior and personality, frontal executive deficits, and language dysfunctions. Depending on symptoms and etiology, FTD can be distinguished in its behavioral-variant FTD (bvFTD) or in two language variants, e.g., semantic variant primary progressive aphasia (svPPA),

10.3389/fncir.2023.1208876

and non-fluent variant primary progressive aphasia (nfvPPA) (Butler and Chiong, 2019). ALS has an incidence of two per 100,000, with almost 120,000 new cases every year world-wide (Chio et al., 2013), while FTD is the second most common early onset dementia under 65 years of age (Harvey et al., 2003) with a conservative incidence of 1.6-4.1 cases per 100,000 per year (Butler and Chiong, 2019). An epidemiologic projection has shown that the number of ALS-FTD cases across the globe will increase from 222.801 in 2015 to 376.674 in 2040, which represents an increase of 69% (Arthur et al., 2016). It is now clear that ALS and FTD can overlap at genetic and pathological level, since genetic mutations in transactive response DNA-binding protein 43 (TARDBP), Fusedin-Sarcoma (FUS) and C9orf72 locus are associated with both disorders (Lomen-Hoerth et al., 2002; Renton et al., 2011; Al-Chalabi et al., 2012). ALS appear sporadically in 90% of cases (sALS), while in 10% of the cases the mutations are inherited (familial-fALS) (Al-Chalabi et al., 2012). Almost 50% of ALS cases develop FTD-like symptoms, while 20% of FTD cases develop ALS (Arthur et al., 2016). These discoveries have transformed the investigations on disease mechanisms; however, the origin and progression of ALS and FTD remain largely unclear and curative therapies do not exist. At a pathological level, dysfunctional cortical inhibition, leading to excitotoxicity, has been previously reported in both ALS and FTD (Vucic and Kiernan, 2006; Zhang et al., 2016). Our recent findings, obtained in the SOD1^{G93A} ALS mouse model, showed loss of spinal inhibitory inputs on motor neurons early in disease, preceding motor neuron death (Allodi et al., 2021). Imbalance between inhibition and excitation can lead to aberrant excitability, intracellular ion dysregulation and cell death, extensively reported in ALS-FTD (Roselli and Caroni, 2015; Jensen et al., 2020). Thus, maladaptive changes within neural circuits and alterations in connectivity might play a pivotal role in disease. An increasing number of studies performed on human post-mortem patients' material, animal models of disease and patient-derived stem cells has reported alterations in synaptic functions in ALS-FTD (Allodi et al., 2021; Scekic-Zahirovic et al., 2021; Brown et al., 2022; Laszlo et al., 2022; Ma et al., 2022). Here, we summarize some of the latest findings while reviewing the neural circuits contributing to disease.

Neural circuits affected in amyotrophic lateral sclerosis

Amyotrophic lateral sclerosis is characterized by progressive paralysis, hence progressive inability to perform movements. While degeneration of motor neurons is the immediate cause of this incurable disease, several cell non-autonomous mechanisms have been demonstrated to contribute to disease onset and progression in combination with the cell autonomous ones (for review Schweingruber and Hedlund, 2022). Here, we will focus on the contribution of intraspinal neurons as well as afferents and corticospinal descending inputs. The spinal cord is a hub for integration of inputs coming from supraspinal descending pathways, local interneuron circuits and sensory afferents. Spinal motor neurons are the ultimate output of the brain, since they directly connect to the muscles, however, within the spinal cord, they are activated by circuits of premotor neurons (Kiehn, 2006). Thus, execution of movements requires the synchronized or reciprocal activation of complex networks of inhibitory and excitatory spinal neurons (Kiehn, 2006). Spinal interneurons can be divided into ventral and dorsal populations depending on their location, and expression of lineage defined molecular markers and neurotransmitters (Briscoe et al., 2000; Lu et al., 2015). Dorsal populations relay somatosensory information coming from afferent pathways (Harding et al., 2020). Here, dorsal inhibitory and excitatory interneurons, as well as projection neurons send information from the spinal cord to the brain (Harding et al., 2020). Ventral interneurons are divided into V0 (V0_C, V0_G, V0_D, and $V0_V$), V1, V2 (V2a and V2b), and V3 cardinal populations. These interneurons regulate different aspects of locomotion, e.g., flexor-extensor coordination (Zhang et al., 2014), muscle force (Zagoraiou et al., 2009), left-right alternation (Crone et al., 2009; Dougherty et al., 2013; Talpalar et al., 2013; Bellardita and Kiehn, 2015) and speed of gait (Gosgnach et al., 2006). Molecular markerspecific expression allows for targeted silencing or depletion of interneuron populations leading to motor alterations and allowing identification of interneuron specific functions (Gosgnach et al., 2006; Zagoraiou et al., 2009; Talpalar et al., 2013; Zhang et al., 2014; Bellardita and Kiehn, 2015). Silencing of inhibitory V1 and V2b neurons disrupts the flexor-extensor muscle reciprocal recruitment (Zhang et al., 2014), while V1 silencing alone reduces the speed of locomotion and decreases stride length (Gosgnach et al., 2006; Allodi et al., 2021). Depletion of V0 commissural neurons causes the loss of left-right alternation of the limbs (Talpalar et al., 2013; Bellardita and Kiehn, 2015), as well as silencing of V2a interneurons perturbs the left-right alternation and the rhythm of locomotion (Crone et al., 2009; Dougherty et al., 2013).

Spinal interneuron loss in human post-mortem ALS tissue was first described in 1895 by Marie Pierre in its "Lectures on diseases of the spinal cord" (London, The New Sydenham). Quantitative studies reported reduction of interneurons in the ventral and intermediate areas of the spinal cord of ALS subjects (Oyanagi et al., 1989; Terao et al., 1994). Moreover, anatomical analysis in sporadic ALS patients showed similar loss of interneurons (56%) and motor neurons (64.4%) within the ventral horn of the spinal cord (Stephens et al., 2006). Not all motor neurons within the spinal cord are equally vulnerable to ALS degeneration. α motor neurons, innervating muscle fibers, degenerate in disease while γ motor neurons, innervating muscle spindles, are spared (Lalancette-Hebert et al., 2016). Among α motor neurons, the ones innervating slow-twitch (S) muscle fibers are more resistant compared to those innervating the fast twitch fatigable (FF) ones (Pun et al., 2006). Spinal motor neurons are known to become hyperexcitable already at presymptomatic stages (Jensen et al., 2020). Our recent findings demonstrated that, under physiological conditions, FF motor neurons receive stronger inputs from the V1 inhibitory population, found in the ventral and intermediate areas of the spinal cord and positive for the transcription factor Engrailed-1 (En1) (Allodi et al., 2021). We also showed that these interneurons are affected in presymptomatic SOD1^{G93A} mice, leading to preferential loss of inhibitory inputs on FF motor neurons. Loss of V1 synaptic inputs paralleled the onset of locomotor phenotype which is characterized by loss of speed, reduction of stride length and step frequency, as well as hyperflexion of the limbs (Allodi et al., 2021). These symptoms are also reported during early stages of disease in ALS patients (Radovanovic et al., 2014). This loss

of synaptic inputs onto FF motor neurons can contribute their imbalanced excitability (Roselli and Caroni, 2015; Jensen et al., 2020). Reduction of V1 inhibitory inputs was reported also in a SOD1 zebrafish model (McGown et al., 2013). V1 interneurons are 80% glycinergic and among them are Renshaw cells and Ia inhibitory interneurons (Alvarez et al., 2005); a reduction of their synaptic inputs onto motor neurons was previously observed in presymptomatic (Chang and Martin, 2009, 2011) as well as in symptomatic mice (Salamatina et al., 2020). Moreover, inhibitory interneuron loss in the intermediate area and dorsal horn of the spinal cord was also reported in a BAC C9ofr72 mouse model (Liu et al., 2016). Interestingly, a recent electrophysiological study reported that, in the early postnatal SOD1^{G93A} mice, glycinergic spinal interneurons were less excitable, showed smaller somas and larger neurites compared to control mice (Cavarsan et al., 2023). Moreover, the glycinergic interneurons most affected in disease, were the most ventrally located (putative Renshaw cells) and the ones within lamina IX (putative Ia) (Cavarsan et al., 2023). Thus, inhibitory interneuron presymptomatic changes can contribute to ALS pathology.

Intraspinal excitatory interneurons seem to degenerate at later stages during disease progression. Reduced synaptic inputs on lumbar spinal motor neurons was reported excitatory in the symptomatic FUS-R521C mouse model (Qiu et al., 2014). In the SOD1^{G93A} mouse, degeneration of V2a excitatory neurons, positive for the transcription factor Chx10 and providing vGlut2 positive inputs, was observed at later symptomatic stages (Romer et al., 2017; Salamatina et al., 2020). C-boutons are specialized synapses known to modulate motor neuron activity (Zagoraiou et al., 2009) and originate from the V0_c cholinergic interneurons positive for Dbx1 and Ptx2 (Zagoraiou et al., 2009). C-boutons were described to undergo structural changes in ALS patients (Nagao et al., 1998) and mouse models of disease (Herron and Miles, 2012; Landoni et al., 2019; Bak et al., 2023). Also, selective loss of tripartite synapses was observed in ALS postmortem spinal cords as well as the SOD1G93A mice spinal cords (Broadhead et al., 2022). Here, astrocytes regulate synaptic transmission by forming tripartite specialized connections, which were found preferentially affected within excitatory synapses (Broadhead et al., 2022). Thus, intraspinal circuits and their connection to motor neurons are affected in ALS throughout disease progression. Evidence from ALS mice suggests that intraspinal inhibitory interneurons might play a role early in disease, while intraspinal excitatory interneurons seem to be affected at later stages.

Depression of afferent inputs, known to provide excitation to motor neurons, has been previously reported in ALS patients (Sangari et al., 2016). Alterations in afferent inputs can contribute to changes in motor neuron excitability. Ia proprioceptive afferents provide a large proportion of vGlut1 positive synapses on α motor neurons (Alvarez et al., 2011) and are fund in higher density on FF motor neurons (Basaldella et al., 2015). Significant loss of vGlut1 synapses was found in symptomatic SOD1^{G93A} and TDP43^{A315T} mice (postnatal day 90) but not at presymptomatic stage (Vaughan et al., 2015). However, electrophysiological studies showed presymptomatic impairment of the excitatory connectivity between Ia afferents and motor neurons, as well as disruption of the vGlut1 postsynaptic density in SOD1 mice (Baczyk et al., 2020). Moreover, inhibition of Ia afferents and spared γ motor neurons was found protective in SOD1^{G93A} mice (Lalancette-Hebert et al., 2016). These data suggest dysfunctions of excitatory afferent inputs early in disease.

Cortical hyperexcitability is a known hallmark of the disease and has been reported in ALS patients (Vucic and Kiernan, 2006). Pyramidal neurons within layer V (also referred to as upper motor neurons) which form the corticospinal tract and project to the spinal cord are affected in disease (Vucic and Kiernan, 2006). Studies performed in ALS mouse models, suggest that this event contributes to the early stages of ALS degeneration, which triggers changes within the spinal cord (Marques et al., 2021; Reale et al., 2023). Interestingly, corticospinal neurons and spinal motor neurons seem to degenerate in a somatotopically related manner (Marques et al., 2021). Motor cortex specific mislocalization of TDP-43 led to cortical hyperexcitability and spinal motor neuron loss selectively in the lumbar segments 1–3 (Reale et al., 2023), the spinal cord segments where the central pattern generators supposedly located (Kiehn, 2006).

In ALS, main efforts have been directed to improve motor neuron survival and muscle reinnervation. However, it is now clear that loss of synapses happens not only in the peripheral nervous system, but also at different levels within the central nervous system. Our recent work shows that rescue of synaptic connectivity between V1 interneurons and motor neurons leads to increased number of spared motor neurons and amelioration of motor phenotype (Mora et al., 2022). Moreover, other studies point towards synaptic connectivity being a therapeutic target for treatment (Baczyk et al., 2020; Catanese et al., 2021). Hence, further investigations on changes in synaptic connectivity in intraspinal, afferent and corticospinal circuits could play an important role in deepening disease understanding and finding new targets for treatment.

Neural circuits affected in frontotemporal dementia

Frontotemporal dementia is characterized by alterations in behavioral domains, including executive functions and social interactions. These symptoms generally appear subtle early in disease, leading to underdiagnosis of the condition. As mentioned above, FTD is found in comorbidity with ALS, due to mutations in the C9orf72, FUS, and TDP-43 genes (Lomen-Hoerth et al., 2002; Al-Chalabi et al., 2012). However, FTD can occur independently from ALS and liked to mutations in the progranulin (GRN) and microtubule-associated tau protein (MAPT) genes among others (Couratier et al., 2017; Butler and Chiong, 2019). In FTD, complex behavioral symptoms are paralleled by complex pathophysiological changes in cortical and subcortical circuits of the brain. Positron emission tomography enabling synaptic density quantification in bvFTD studies, showed severe synaptic loss of multiple brain regions including, the medial and dorsolateral frontal regions of the brain, the inferior frontal gyri, the anterior and posterior cingulate gyrus, the insular cortex, and the medial temporal lobe (Malpetti et al., 2023), here, synaptic loss was more severe than overall brain atrophy (Malpetti et al., 2023). Moreover, lower levels of synaptic proteins such as synaptotagmins and synaptophysin were



FIGURE 1

Neural circuits involved in ALS-FTD. Schematic representation of the circuits associated with amyotrophic lateral sclerosis (ALS) and frontotemporal dementia (FTD) as manifestations of the same continuum. (A) In ALS, pathological changes can be first seen in the brainstem somatic motor neurons, whose death results in upper-body phenotype, known as "bulbar onset," or in spinal somatic motor neurons, whose death leads to lower limbs symptomatology in the so-called "lumbar onset," along with death of the corticospinal neurons of the motor cortex. Affected areas are shown in dark pink. FTD affects the frontal and temporal lobes of the brain. Areas contributing to the disease are depicted in light blue. (B) Schematic of intraspinal lumbar circuits known to play a role in motor control. Spatial localization of ventral cardinal populations (VO_C, VO_D, VO_V, V1, V2a, V2b, and V3) and some dorsal populations. Inhibitory (pink) and excitatory (purple) populations and inputs are depicted, dashed lines indicate known structure degeneration. (C) At a systems level, frontotemporal dementia has been linked to altered corticostriatal circuits [adapted from Dalley et al. (2011) and Radulescu et al. (2017)]: cortex sends excitatory glutamatergic projections to the medium spiny neurons of striatum expressing dopamine receptor 1 and 2, which inhibit each other and the internal and external, respectively, regions of globus pallidus via GABAergic activity. Its external part sends inhibitory projections to striatum and cortex.

observed years before disease onset in FTD patients (Goetzl et al., 2016).

The C9orf72 pathology is characterized by hexanucleotide repeat expansion in the first intron of the C9orf72 gene, a common ALS-FTD mutation that results in the formation of nuclear RNA foci, dipeptide repeat protein inclusions and TDP43 aggregates and is the most frequent reported genetic cause of ALS-FTD (Renton et al., 2011). Studies performed on synaptoneurosomes obtained from ALS post-mortem human cortex carrying C9orf72 hexanucleotide repeat expansion revealed extensive synaptic dysfunction for both excitatory and inhibitory synapses (Laszlo et al., 2022). Mice carrying C9orf72 hexanucleotide repeat expansion show abnormal social interaction, hyperactivity, anxious phenotype and, in some cases, motor deficits (Chew et al., 2015;

Jiang et al., 2016; Liu et al., 2016) and memory impairment (Jiang et al., 2016). Anatomical studies performed on these mice have reported lower neuron count within the cortex (especially the motor areas) and the cerebellum, paralleled with reactive gliosis (Chew et al., 2015; Jiang et al., 2016; Liu et al., 2016). Reduction of hippocampal neurons was also shown in two BAC C9orf72 transgenic mouse lines (Jiang et al., 2016; Liu et al., 2016), while only one of them in spinal motor neuron and interneuron loss was observed (Liu et al., 2016). However, while neuronal loss has been more thoroughly investigated, less is known about synaptic connectivity in the C9orf72 mouse models.

TDP-43 and FUS play an important role in regulating strength of synaptic transmission (for review Ling, 2018). Gene expression analysis in FTD post-mortem human tissue with TDP-43 pathology

TABLE 1	Altered	biomarkers	underlyi	ing clinica	l symptoma	itology.
---------	---------	------------	----------	-------------	------------	----------

	Structure	Clinical output	Animal model				
			C9ORF72	TDP-43	FUS	Tau	GRN
	Frontal cortex	Apathy	-	↓ Synaptic activity and formation; gene expression	 ↓ Synaptic connectivity ↓ Dendritic spine number 		↓ Late dendritic arborization
		Dietary shifts			 ↓ EPSCs amplitude ↑ neuronal activity 		
	Striatum	Compulsive behavior				●↓ NMDAR synaptic density ●↓ firing	↑ Excitability of spiny neurons
	Motor cortex	Stereotypies	Reactive gliosis	•Hyperactive Sst neurons •↓ GABA transmission •↑ Excitability, excitotoxicity	 ↑ Excitatory synapses markers ↓ Inhibitory synapses markers ↓ Dendritic length 		
	Hippocampus	Memory deficits	↓ Neuron count	 ↑ Microglial activity ↓ Ventral mature and precursor granular cells 	 ↓ Synaptic connectivity ↓ Dendritic spine number 	 ↓ Ventral mature and precursor granular cells ↑ GABAergic innervation 	 ↑ Microglial activity ↓ Synaptic connectivity ↓ LTP ↓ Dendritic length and spine density and number
	IN type	Function	Animal model				
			C9ORF72	OD1 TDP-43 FUS			
	V0c	Muscle tone		Early enlargement, then loss of synapses			
SR	V2a	Rhythm of locomotion		•↓ Chx10+ neuron count •↓ Synapses			
	V1	Speed of locomotionStride length		•↓ En1+ neuron count •↓ Synapses			
	*Not specified		↓ Interneuron count	↓ Excitatory synapses (vGlut1)	 ↑ excitatory synapses (vGlut2) ↑ inhibitory synapses (GAD65/67) 	↓ excitatory synapses	

It summarizes the link between the aforementioned pathophysiological alterations in ALS-FTD by brain area and relevant symptomatology associated with alterations of such regions. Although more or less defined links between brain pathology and clinical outcomes are stated for the sake of simplicity, future studies should aim to investigate the phenomenon from a circuit neuroscience perspective given the nosological complexity of it and the roles of these structures in more than one behavior under normal conditions, especially when taking into account fronto-striatal circuits (Dalley et al., 2011).

reported downregulation of genes involved in synaptic activity and synapse formation (Mishra et al., 2007). Similar results were obtained by cross-linking and immunoprecipitation coupled with high-throughput sequencing performed in the brain of a mouse model upon reduction in TDP-43 (Polymenidou et al., 2011). Several of the generated TDP-43 transgenic mice recapitulate ALS-FTD pathology and symptoms (among others Wegorzewska et al., 2009; Wils et al., 2010; Swarup et al., 2011). Here, layer V pyramidal neurons in the frontal, motor and somatosensory corteces, spinal motor neurons (Wegorzewska et al., 2009; Wils et al., 2010; Igaz et al., 2011) as well as hippocampal neurons (Igaz et al., 2011) were found preferentially affected. In a TDP-43 transgenic mouse, hyperactive somatostatin positive interneurons were shown to indirectly disinhibit layer V pyramidal neurons leading to excitotoxicity (Zhang et al., 2016). Severe cognitive impairment within the learning and memory domains was reported in TPD-43 transgenic mice (Swarup et al., 2011), also when TDP-43 overexpression was restricted to the forebrain (Tsai et al., 2010). Here, progressive motor dysfunctions were also observed (Tsai et al., 2010). FUS is an important protein involved in the processing of coding and non-coding RNA, and in the maintenance of genomic integrity. Mutations in the FUS gene induce pathological cytoplasmic mislocalization of the protein, especially within the stress granules, which can be observed in human post-mortem tissue and FUS mouse models, in absence of TDP-43 pathology (Nolan et al., 2016). FUS pathology has been reported to affect neurons within frontal and temporal neocortex, hippocampus, and striatum (Mackenzie et al., 2011). Several FUS transgenic mice were generated, however, not all of them showed neuronal degeneration even in presence of cognitive and motor dysfunctions (for review Nolan et al., 2016). Changes in synaptic connectivity and dendritic spine morphology were described in the sensorimotor cortex (Qiu et al., 2014). In the Fus^{Δ NLS/+} mouse, increase of FUS at synaptic level was observed early in disease and led to changes in density and size of GABAergic synapses (Shiihashi et al., 2017; Sahadevan et al., 2021). The Fus^{Δ NLS/+} transgenic mice showed increased activity in the layers II/III of the frontal cortex and overall reduced brain volume, which were paralleled by increased social interaction and hyperactivity, along with worse learning and memory (Shiihashi et al., 2017; Scekic-Zahirovic et al., 2021).

Other known FTD-linked mutations are found in the progranulin (GRN) and microtubule-associated tau protein (MAPT) genes. GRN deficiency leads to progranulin haploinsufficiency, and has been linked to several behavioral impairments and central nervous system alterations (Roberson, 2012). Here, top-down cognitive control is impaired (Dalley et al., 2011), and cortico-striatal-thalamic alterations are commonly reported. GRN transgenic mouse models show decreased synaptic connectivity and impaired plasticity (Petkau et al., 2012; Lui et al., 2016). Changes in dendritic arborization and electrophysiological alterations in prefrontal regions, amygdala and thalamus were also reported (Arrant et al., 2016; Lui et al., 2016; Nagy et al., 2019). Phenotypically, dysfunctional social interaction and motor coordination (Petkau et al., 2012; Arrant et al., 2016) were shown, as well as impairment of executive functions leading to excessive grooming, deficient exploration of novel objects, and increased reward seeking (Petkau et al., 2012; Lui et al., 2016; Nagy et al., 2019). Tau pathology also leads to protein accumulation and is linked to behavioral and neurophysiological changes. Transgenic mouse models show degeneration in the hippocampus and increased GABAergic inhibition (Llorens-Martin et al., 2011), as well as NMDA synaptic deficits in the ventral striatum and insula (Warmus et al., 2014). Alterations in executive function and emotional processing were also reported, such as compulsive grooming and increased immobility (Llorens-Martin et al., 2011; Warmus et al., 2014). Altogether, these findings support the idea that alterations in synaptic connectivity within specific neural circuits play a pivotal role in FTD pathophysiology.

Discussion

This minireview focuses on the existing evidence that several neural circuits are affected in ALS-FTD pathology. Here, results obtained from ALS-FTD human post-mortem tissue and mouse models of disease suggest that changes in synaptic connectivity and subsequent imbalance between inhibitory and excitatory networks might play a role, together with other pathological events, at different stages of disease. Imaging studies suggest that FTD affects the salience network, which includes multimodal networks identifying salient information (Roberson, 2012). The salience network is found in the anterior cingulate and ventral anterior insular cortices, and it is phylogenically conserved between mice and men (Roberson, 2012). Thus, while in humans, ALS and FTD are characterized by the degeneration of Betz cells and von Economo neurons within the prefrontal and frontal cortices, which are not present in mice, network activity and degeneration can still be mimicked in mouse models. Cell types are not the only non-conserved feature among species, the corticospinal tract is also known to differ anatomically and functionally between humans and mice. Moreover, intraspinal neural networks controlling different aspects of motor control and forming the central pattern generators are under investigated in the human spinal cord, although new comparative studies points toward maintenance of spinal neuron molecular markers (Yadav et al., 2023) and functions (Kathe et al., 2022). Overall, further investigations are needed to deepen our understanding of neural networks alteration in ALS-FTD and their contribution to neuronal vulnerability in disease.

Author contributions

IA: conceptualization and funding acquisition. Both authors contributed to writing - original draft, figure, table and the article and approved the submitted version.

Funding

This work was supported by the Lundbeck Foundation (IA) (Grant No. R346-2020-202) and the Department of Neuroscience, University of Copenhagen and School of Psychology and Neuroscience, University of St Andrews.

Acknowledgments

We thank Peter Simonsson for the help in the drawings in Figure 1. Illustrations in Table 1 were created with BioRender.com.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships

References

Al-Chalabi, A., Jones, A., Troakes, C., King, A., Al-Sarraj, S., and Van Den Berg, L. H. (2012). The genetics and neuropathology of amyotrophic lateral sclerosis. *Acta Neuropathol.* 124, 339–352.

Allodi, I., Montanana-Rosell, R., Selvan, R., Low, P., and Kiehn, O. (2021). Locomotor deficits in a mouse model of ALS are paralleled by loss of V1-interneuron connections onto fast motor neurons. *Nat. Commun.* 12:3251. doi: 10.1038/s41467-021-23224-7

Alvarez, F. J., Jonas, P. C., Sapir, T., Hartley, R., Berrocal, M. C., Geiman, E. J., et al. (2005). Postnatal phenotype and localization of spinal cord V1 derived interneurons. *J. Comp. Neurol.* 493, 177–192. doi: 10.1002/cne.20711

Alvarez, F. J., Titus-Mitchell, H. E., Bullinger, K. L., Kraszpulski, M., Nardelli, P., and Cope, T. C. (2011). Permanent central synaptic disconnection of proprioceptors after nerve injury and regeneration. I. Loss of VGLUT1/IA synapses on motoneurons. J. Neurophysiol 106, 2450–2470. doi: 10.1152/jn.01095.2010

Arrant, A. E., Filiano, A. J., Warmus, B. A., Hall, A. M., and Roberson, E. D. (2016). Progranulin haploinsufficiency causes biphasic social dominance abnormalities in the tube test. *Genes Brain Behav.* 15, 588–603. doi: 10.1111/gbb.12300

Arthur, K. C., Calvo, A., Price, T. R., Geiger, J. T., Chio, A., and Traynor, B. J. (2016). Projected increase in amyotrophic lateral sclerosis from 2015 to 2040. *Nat. Commun.* 7:12408.

Baczyk, M., Alami, N. O., Delestree, N., Martinot, C., Tang, L., Commisso, B., et al. (2020). Synaptic restoration by cAMP/PKA drives activity-dependent neuroprotection to motoneurons in ALS. J. Exp. Med. 217:e20191734. doi: 10.1084/jem.20191734

Bak, A. N., Djukic, S., Kadlecova, M., Braunstein, T. H., Jensen, D. B., and Meehan, C. F. (2023). Cytoplasmic TDP-43 accumulation drives changes in C-bouton number and size in a mouse model of sporadic Amyotrophic Lateral Sclerosis. *Mol. Cell Neurosci.* 125:103840. doi: 10.1016/j.mcn.2023.103840

Basaldella, E., Takeoka, A., Sigrist, M., and Arber, S. (2015). Multisensory signaling shapes vestibulo-motor circuit specificity. *Cell* 163, 301–312. doi: 10.1016/j.cell.2015. 09.023

Bellardita, C., and Kiehn, O. (2015). Phenotypic characterization of speed-associated gait changes in mice reveals modular organization of locomotor networks. *Curr. Biol.* 25, 1426–1436. doi: 10.1016/j.cub.2015.04.005

Briscoe, J., Pierani, A., Jessell, T. M., and Ericson, J. (2000). A homeodomain protein code specifies progenitor cell identity and neuronal fate in the ventral neural tube. *Cell* 101, 435–445. doi: 10.1016/s0092-8674(00)80853-3

Broadhead, M. J., Bonthron, C., Waddington, J., Smith, W. V., Lopez, M. F., Burley, S., et al. (2022). Selective vulnerability of tripartite synapses in amyotrophic lateral sclerosis. *Acta Neuropathol.* 143, 471–486. doi: 10.1007/s00401-022-02412-9

Brown, A. L., Wilkins, O. G., Keuss, M. J., Hill, S. E., Zanovello, M., Lee, W. C., et al. (2022). TDP-43 loss and ALS-risk SNPs drive mis-splicing and depletion of UNC13A. *Nature* 603, 131–137. doi: 10.1038/s41586-022-04436-3

Butler, P. M., and Chiong, W. (2019). Neurodegenerative disorders of the human frontal lobes. *Handb. Clin. Neurol.* 163, 391-410.

Catanese, A., Rajkumar, S., Sommer, D., Freisem, D., Wirth, A., Aly, A., et al. (2021). Synaptic disruption and CREB-regulated transcription are restored by K(+) channel blockers in ALS. *EMBO Mol. Med.* 13:e13131. doi: 10.15252/emmm.2020 13131

Cavarsan, C. F., Steele, P. R., Genry, L. T., Reedich, E. J., Mccane, L. M., Lapre, K. J., et al. (2023). Inhibitory interneurons show early dysfunction in a SOD1 mouse model of amyotrophic lateral sclerosis. *J. Physiol.* 601, 647–667. doi: 10.1113/JP28 4192

that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Chang, Q., and Martin, L. J. (2009). Glycinergic innervation of motoneurons is deficient in amyotrophic lateral sclerosis mice: a quantitative confocal analysis. *Am. J. Pathol.* 174, 574–585. doi: 10.2353/ajpath.2009.080557

Chang, Q., and Martin, L. J. (2011). Glycine receptor channels in spinal motoneurons are abnormal in a transgenic mouse model of amyotrophic lateral sclerosis. *J. Neurosci.* 31, 2815–2827. doi: 10.1523/JNEUROSCI.2475-10.2011

Chew, J., Gendron, T. F., Prudencio, M., Sasaguri, H., Zhang, Y. J., Castanedes-Casey, M., et al. (2015). Neurodegeneration. C9ORF72 repeat expansions in mice cause TDP-43 pathology, neuronal loss, and behavioral deficits. *Science* 348, 1151–1154.

Chio, A., Logroscino, G., Traynor, B. J., Collins, J., Simeone, J. C., Goldstein, L. A., et al. (2013). Global epidemiology of amyotrophic lateral sclerosis: a systematic review of the published literature. *Neuroepidemiology* 41, 118–130.

Couratier, P., Corcia, P., Lautrette, G., Nicol, M., and Marin, B. (2017). ALS and frontotemporal dementia belong to a common disease spectrum. *Rev. Neurol.* 173, 273–279.

Crone, S. A., Zhong, G., Harris-Warrick, R., and Sharma, K. (2009). In mice lacking V2a interneurons, gait depends on speed of locomotion. *J. Neurosci.* 29, 7098–7109.

Dalley, J. W., Everitt, B. J., and Robbins, T. W. (2011). Impulsivity, compulsivity, and top-down cognitive control. *Neuron* 69, 680–694.

Dougherty, K. J., Zagoraiou, L., Satoh, D., Rozani, I., Doobar, S., Arber, S., et al. (2013). Locomotor rhythm generation linked to the output of spinal shox2 excitatory interneurons. *Neuron* 80, 920–933. doi: 10.1016/j.neuron.2013.08.015

Goetzl, E. J., Kapogiannis, D., Schwartz, J. B., Lobach, I. V., Goetzl, L., Abner, E. L., et al. (2016). Decreased synaptic proteins in neuronal exosomes of frontotemporal dementia and Alzheimer's disease. *FASEB J.* 30, 4141–4148. doi: 10. 1096/fj.201600816R

Gosgnach, S., Lanuza, G. M., Butt, S. J., Saueressig, H., Zhang, Y., Velasquez, T., et al. (2006). V1 spinal neurons regulate the speed of vertebrate locomotor outputs. *Nature* 440, 215–219. doi: 10.1038/nature04545

Harding, E. K., Fung, S. W., and Bonin, R. P. (2020). Insights into spinal dorsal horn circuit function and dysfunction using optical approaches. *Front. Neural Circuits* 14:31. doi: 10.3389/fncir.2020.00031

Harvey, R. J., Skelton-Robinson, M., and Rossor, M. N. (2003). The prevalence and causes of dementia in people under the age of 65 years. *J. Neurol. Neurosurg. Psychiatry* 74, 1206–1209.

Herron, L. R., and Miles, G. B. (2012). Gender-specific perturbations in modulatory inputs to motoneurons in a mouse model of amyotrophic lateral sclerosis. *Neuroscience* 226, 313–323. doi: 10.1016/j.neuroscience.2012.09.031

Igaz, L. M., Kwong, L. K., Lee, E. B., Chen-Plotkin, A., Swanson, E., Unger, T., et al. (2011). Dysregulation of the ALS-associated gene TDP-43 leads to neuronal death and degeneration in mice. *J. Clin. Invest.* 121, 726–738.

Jensen, D. B., Kadlecova, M., Allodi, I., and Meehan, C. F. (2020). Spinal motoneurones are intrinsically more responsive in the adult G93A SOD1 mouse model of amyotrophic lateral sclerosis. *J. Physiol.* 598, 4385–4403. doi: 10.1113/JP280097

Jiang, J., Zhu, Q., Gendron, T. F., Saberi, S., Mcalonis-Downes, M., Seelman, A., et al. (2016). Gain of toxicity from ALS/FTD-linked repeat expansions in C9ORF72 is alleviated by antisense oligonucleotides targeting GGGGCC-Containing RNAs. *Neuron* 90, 535–550. doi: 10.1016/j.neuron.2016.04.006

Kathe, C., Skinnider, M. A., Hutson, T. H., Regazzi, N., Gautier, M., Demesmaeker, R., et al. (2022). The neurons that restore walking after paralysis. *Nature* 611, 540–547.

Kiehn, O. (2006). Locomotor circuits in the mammalian spinal cord. Annu. Rev. Neurosci. 29, 279–306.

Lalancette-Hebert, M., Sharma, A., Lyashchenko, A. K., and Shneider, N. A. (2016). Gamma motor neurons survive and exacerbate alpha motor neuron degeneration in ALS. Proc. Natl. Acad. Sci. U. S. A. 113, E8316–E8325. doi: 10.1073/pnas.1605210113

Landoni, L. M., Myles, J. R., Wells, T. L., Mayer, W. P., and Akay, T. (2019). Cholinergic modulation of motor neurons through the C-boutons are necessary for the locomotor compensation for severe motor neuron loss during amyotrophic lateral sclerosis disease progression. *Behav. Brain Res.* 369:111914. doi: 10.1016/j.bbr.2019. 111914

Laszlo, Z. I., Hindley, N., Sanchez Avila, A., Kline, R. A., Eaton, S. L., Lamont, D. J., et al. (2022). Synaptic proteomics reveal distinct molecular signatures of cognitive change and C9ORF72 repeat expansion in the human ALS cortex. *Acta Neuropathol. Commun.* 10:156. doi: 10.1186/s40478-022-01455-z

Ling, S. C. (2018). Synaptic paths to neurodegeneration: The emerging role of TDP-43 and FUS in synaptic functions. *Neural Plast.* 2018:8413496. doi: 10.1155/2018/ 8413496

Liu, Y., Pattamatta, A., Zu, T., Reid, T., Bardhi, O., Borchelt, D. R., et al. (2016). C9orf72 BAC Mouse Model with Motor Deficits and Neurodegenerative Features of ALS/FTD. *Neuron* 90, 521–534. doi: 10.1016/j.neuron.2016.04.005

Llorens-Martin, M., Hernandez, F., and Avila, J. (2011). Expression of frontotemporal dementia with parkinsonism associated to chromosome 17 tau induces specific degeneration of the ventral dentate gyrus and depressive-like behavior in mice. *Neuroscience* 196, 215–227. doi: 10.1016/j.neuroscience.2011.08.057

Lomen-Hoerth, C., Anderson, T., and Miller, B. (2002). The overlap of amyotrophic lateral sclerosis and frontotemporal dementia. *Neurology* 59, 1077–1079.

Lu, D. C., Niu, T., and Alaynick, W. A. (2015). Molecular and cellular development of spinal cord locomotor circuitry. *Front. Mol. Neurosci.* 8:25. doi: 10.3389/fnmol.2015. 00025

Lui, H., Zhang, J., Makinson, S. R., Cahill, M. K., Kelley, K. W., Huang, H. Y., et al. (2016). Progranulin deficiency promotes circuit-specific synaptic pruning by microglia via complement activation. *Cell* 165, 921–935. doi: 10.1016/j.cell.2016.04.001

Ma, X. R., Prudencio, M., Koike, Y., Vatsavayai, S. C., Kim, G., Harbinski, F., et al. (2022). TDP-43 represses cryptic exon inclusion in the FTD-ALS gene UNC13A. *Nature* 603, 124–130. doi: 10.1038/s41586-022-04424-7

Mackenzie, I. R., Munoz, D. G., Kusaka, H., Yokota, O., Ishihara, K., Roeber, S., et al. (2011). Distinct pathological subtypes of FTLD-FUS. *Acta Neuropathol.* 121, 207–218.

Malpetti, M., Jones, P. S., Cope, T. E., Holland, N., Naessens, M., Rouse, M. A., et al. (2023). Synaptic loss in frontotemporal dementia revealed by [(11) C]UCB-J positron emission tomography. *Ann. Neurol.* 93, 142–154. doi: 10.1002/ana.26543

Marques, C., Burg, T., Scekic-Zahirovic, J., Fischer, M., and Rouaux, C. (2021). Upper and lower motor neuron degenerations are somatotopically related and temporally ordered in the sod1 mouse model of amyotrophic lateral sclerosis. *Brain Sci.* 11, 369. doi: 10.3390/brainsci11030369

McGown, A., Mcdearmid, J. R., Panagiotaki, N., Tong, H., Al Mashhadi, S., Redhead, N., et al. (2013). Early interneuron dysfunction in ALS: insights from a mutant sod1 zebrafish model. *Ann. Neurol.* 73, 246–258. doi: 10.1002/ana.23780

Mishra, M., Paunesku, T., Woloschak, G. E., Siddique, T., Zhu, L. J., Lin, S., et al. (2007). Gene expression analysis of frontotemporal lobar degeneration of the motor neuron disease type with ubiquitinated inclusions. *Acta Neuropathol.* 114, 81–94.

Mora, S., Von Huth Friis, R., Stuckert, A., Noes-Holt, G., Montañana-Rosell, R., Toft Sørensen, A., et al. (2022). Stabilization of V1 interneuron-motor neuron connectivity ameliorates motor phenotype in a mouse model of ALS. *bioRxiv* [Preprint]. doi: 10.1101/2022.12.15.520568

Nagao, M., Misawa, H., Kato, S., and Hirai, S. (1998). Loss of cholinergic synapses on the spinal motor neurons of amyotrophic lateral sclerosis. *J. Neuropathol. Exp. Neurol.* 57, 329–333.

Nagy, D., Martens, L. H., Leventhal, L., Chen, A., Kelley, C., Stoiljkovic, M., et al. (2019). Age-dependent emergence of neurophysiological and behavioral abnormalities in progranulin-deficient mice. *Alzheimers Res. Ther.* 11, 88. doi: 10.1186/s13195-019-0540-x

Nolan, M., Talbot, K., and Ansorge, O. (2016). Pathogenesis of FUS-associated ALS and FTD: insights from rodent models. *Acta Neuropathol. Commun.* 4, 99. doi: 10.1186/s40478-016-0358-8

Oyanagi, K., Ikuta, F., and Horikawa, Y. (1989). Evidence for sequential degeneration of the neurons in the intermediate zone of the spinal cord in amyotrophic lateral sclerosis: a topographic and quantitative investigation. *Acta Neuropathol.* 77, 343–349. doi: 10.1007/BF00687368

Petkau, T. L., Neal, S. J., Milnerwood, A., Mew, A., Hill, A. M., Orban, P., et al. (2012). Synaptic dysfunction in progranulin-deficient mice. *Neurobiol. Dis.* 45, 711–722.

Polymenidou, M., Lagier-Tourenne, C., Hutt, K. R., Huelga, S. C., Moran, J., Liang, T. Y., et al. (2011). Long pre-mRNA depletion and RNA missplicing contribute to neuronal vulnerability from loss of TDP-43. *Nat. Neurosci.* 14, 459–468. doi: 10.1038/nn.2779

Pun, S., Santos, A. F., Saxena, S., Xu, L., and Caroni, P. (2006). Selective vulnerability and pruning of phasic motoneuron axons in motoneuron disease alleviated by CNTF. *Nat. Neurosci.* 9, 408–419. doi: 10.1038/nn1653

Qiu, H., Lee, S., Shang, Y., Wang, W. Y., Au, K. F., Kamiya, S., et al. (2014). ALS-associated mutation FUS-R521C causes DNA damage and RNA splicing defects. *J. Clin. Invest.* 124, 981–999.

Radovanovic, S., Milicev, M., Peric, S., Basta, I., Kostic, V., and Stevic, Z. (2014). Gait in amyotrophic lateral sclerosis: Is gait pattern differently affected in spinal and bulbar onset of the disease during dual task walking? *Amyotroph. Lateral Scler. Frontotemporal Degener.* 15, 488–493. doi: 10.3109/21678421.2014.918148

Radulescu, A., Herron, J., Kennedy, C., and Scimemi, A. (2017). Global and local excitation and inhibition shape the dynamics of the cortico-striatal-thalamo-cortical pathway. *Sci. Rep.* 7, 7608. doi: 10.1038/s41598-017-07527-8

Reale, L. A., Dyer, M. S., Perry, S. E., Young, K. M., Dickson, T. C., Woodhouse, A., et al. (2023). Pathologically mislocalised TDP-43 in upper motor neurons causes a die-forward spread of ALS-like pathogenic changes throughout the mouse corticomotor system. *Prog. Neurobiol.* 226, 102449. doi: 10.1016/j.pneurobio.2023.102449

Renton, A. E., Majounie, E., Waite, A., Simon-Sanchez, J., Rollinson, S., Gibbs, J. R., et al. (2011). A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD. *Neuron* 72, 257–268.

Roberson, E. D. (2012). Mouse models of frontotemporal dementia. Ann. Neurol. 72, 837-849.

Romer, S. H., Seedle, K., Turner, S. M., Li, J., Baccei, M. L., and Crone, S. A. (2017). Accessory respiratory muscles enhance ventilation in ALS model mice and are activated by excitatory V2a neurons. *Exp. Neurol.* 287, 192–204. doi: 10.1016/j. expneurol.2016.05.033

Roselli, F., and Caroni, P. (2015). From intrinsic firing properties to selective neuronal vulnerability in neurodegenerative diseases. *Neuron* 85, 901–910. doi: 10. 1016/j.neuron.2014.12.063

Sahadevan, S., Hembach, K. M., Tantardini, E., Perez-Berlanga, M., Hruska-Plochan, M., Megat, S., et al. (2021). Synaptic FUS accumulation triggers early misregulation of synaptic RNAs in a mouse model of ALS. *Nat. Commun.* 12, 3027. doi: 10.1038/s41467-021-23188-8

Salamatina, A., Yang, J. H., Brenner-Morton, S., Bikoff, J. B., Fang, L., Kintner, C. R., et al. (2020). Differential loss of spinal interneurons in a mouse model of ALS. *Neuroscience* 450, 81–95. doi: 10.1016/j.neuroscience.2020.0 8.011

Sangari, S., Iglesias, C., El Mendili, M. M., Benali, H., Pradat, P. F., and Marchand-Pauvert, V. (2016). Impairment of sensory-motor integration at spinal level in amyotrophic lateral sclerosis. *Clin. Neurophysiol.* 127, 1968–1977. doi: 10.1016/j. clinph.2016.01.014

Scekic-Zahirovic, J., Sanjuan-Ruiz, I., Kan, V., Megat, S., De Rossi, P., Dieterle, S., et al. (2021). Cytoplasmic FUS triggers early behavioral alterations linked to cortical neuronal hyperactivity and inhibitory synaptic defects. *Nat. Commun.* 12:3028. doi: 10.1038/s41467-021-23187-9

Schweingruber, C., and Hedlund, E. (2022). The cell autonomous and non-cell autonomous aspects of neuronal vulnerability and resilience in amyotrophic lateral sclerosis. *Biology* 11, 1191. doi: 10.3390/biology1108 1191

Shiihashi, G., Ito, D., Arai, I., Kobayashi, Y., Hayashi, K., Otsuka, S., et al. (2017). Dendritic homeostasis disruption in a novel frontotemporal dementia mouse model expressing cytoplasmic fused in sarcoma. *EBioMedicine* 24, 102–115. doi: 10.1016/j. ebiom.2017.09.005

Stephens, B., Guiloff, R. J., Navarrete, R., Newman, P., Nikhar, N., and Lewis, P. (2006). Widespread loss of neuronal populations in the spinal ventral horn in sporadic motor neuron disease. A morphometric study. J. Neurol. Sci. 244, 41–58. doi: 10.1016/j.jns.2005.12.003

Swarup, V., Phaneuf, D., Bareil, C., Robertson, J., Rouleau, G. A., Kriz, J., et al. (2011). Pathological hallmarks of amyotrophic lateral sclerosis/frontotemporal lobar degeneration in transgenic mice produced with TDP-43 genomic fragments. *Brain* 134, 2610–2626. doi: 10.1093/brain/awr159

Talpalar, A. E., Bouvier, J., Borgius, L., Fortin, G., Pierani, A., and Kiehn, O. (2013). Dual-mode operation of neuronal networks involved in left-right alternation. *Nature* 500, 85–88. doi: 10.1038/nature12286

Terao, S., Sobue, G., Hashizume, Y., Mitsuma, T., and Takahashi, A. (1994). Diseasespecific patterns of neuronal loss in the spinal ventral horn in amyotrophic lateral sclerosis, multiple system atrophy and X-linked recessive bulbospinal neuronopathy, with special reference to the loss of small neurons in the intermediate zone. *J. Neurol.* 241, 196–203. doi: 10.1007/BF00863768

Tsai, K. J., Yang, C. H., Fang, Y. H., Cho, K. H., Chien, W. L., Wang, W. T., et al. (2010). Elevated expression of TDP-43 in the forebrain of mice is sufficient to cause neurological and pathological phenotypes mimicking FTLD-U. *J. Exp. Med.* 207, 1661–1673. doi: 10.1084/jem.20092164

Vaughan, S. K., Kemp, Z., Hatzipetros, T., Vieira, F., and Valdez, G. (2015). Degeneration of proprioceptive sensory nerve endings in mice harboring amyotrophic lateral sclerosis-causing mutations. *J. Comp. Neurol.* 523, 2477–2494. doi: 10.1002/cne. 23848

Vucic, S., and Kiernan, M. C. (2006). Novel threshold tracking techniques suggest that cortical hyperexcitability is an early feature of motor neuron disease. *Brain* 129, 2436–2446.

Warmus, B. A., Sekar, D. R., Mccutchen, E., Schellenberg, G. D., Roberts, R. C., Mcmahon, L. L., et al. (2014). Tau-mediated NMDA receptor impairment underlies dysfunction of a selectively vulnerable network in a mouse model of frontotemporal dementia. *J. Neurosci.* 34, 16482–16495. doi: 10.1523/JNEUROSCI.3418-14. 2014

Wegorzewska, I., Bell, S., Cairns, N. J., Miller, T. M., and Baloh, R. H. (2009). TDP-43 mutant transgenic mice develop features of ALS and frontotemporal lobar degeneration. *Proc. Natl. Acad. Sci. U. S. A.* 106, 18809–18814.

Wils, H., Kleinberger, G., Janssens, J., Pereson, S., Joris, G., Cuijt, I., et al. (2010). TDP-43 transgenic mice develop spastic paralysis and neuronal inclusions characteristic of ALS and frontotemporal lobar degeneration. *Proc. Natl. Acad. Sci. U. S. A.* 107, 3858–3863. doi: 10.1073/pnas.09124 17107

Yadav, A., Matson, K. J. E., Li, L., Hua, I., Petrescu, J., Kang, K., et al. (2023). A cellular taxonomy of the adult human spinal cord. *Neuron* 111, 328–344.e327.

Zagoraiou, L., Akay, T., Martin, J. F., Brownstone, R. M., Jessell, T. M., and Miles, G. B. (2009). A cluster of cholinergic premotor interneurons modulates mouse locomotor activity. *Neuron* 64, 645–662.

Zhang, J., Lanuza, G. M., Britz, O., Wang, Z., Siembab, V. C., Zhang, Y., et al. (2014). V1 and v2b interneurons secure the alternating flexor-extensor motor activity mice require for limbed locomotion. *Neuron* 82, 138–150. doi: 10.1016/j.neuron.2014.02. 013

Zhang, W., Zhang, L., Liang, B., Schroeder, D., Zhang, Z. W., Cox, G. A., et al. (2016). Hyperactive somatostatin interneurons contribute to excitotoxicity in neurodegenerative disorders. *Nat. Neurosci.* 19, 557–559. doi: 10.1038/nn.4257