

Research



Cite this article: Hitchcock TJ, Gardner A. 2021 Sex-biased demography modulates male harm across the genome. *Proc. R. Soc. B* **288**: 20212237.
<https://doi.org/10.1098/rspb.2021.2237>

Received: 13 October 2021

Accepted: 26 November 2021

Subject Category:

Evolution

Subject Areas:

evolution, genetics, theoretical biology

Keywords:

cytoplasmic inheritance, intragenomic conflict, overlapping generations, sex chromosome, sexual conflict, soft selection

Author for correspondence:

Thomas J. Hitchcock

e-mail: th76@st-andrews.ac.uk

Electronic supplementary material is available online at <https://doi.org/10.6084/m9.figshare.c.5752431>.

Sex-biased demography modulates male harm across the genome

Thomas J. Hitchcock and Andy Gardner

School of Biology, University of St Andrews, St Andrews KY16 9TH, UK

TJH, 0000-0002-6378-5023; AG, 0000-0002-1304-3734

Recent years have seen an explosion of theoretical and empirical interest in the role that kin selection plays in shaping patterns of sexual conflict, with a particular focus on male harming traits. However, this work has focused solely on autosomal genes, and as such it remains unclear how demography modulates the evolution of male harm loci occurring in other portions of the genome, such as sex chromosomes and cytoplasmic elements. To investigate this, we extend existing models of sexual conflict for application to these different modes of inheritance. We first analyse the general case, revealing how sex-specific relatedness, reproductive value and the intensity of local competition combine to determine the potential for male harm. We then analyse a series of demographically explicit models, to assess how dispersal, overlapping generations, reproductive skew and the mechanism of population regulation affect sexual conflict across the genome, and drive conflict between nuclear and cytoplasmic genes. We then explore the effects of sex biases in these demographic parameters, showing how they may drive further conflicts between autosomes and sex chromosomes. Finally, we outline how different crossing schemes may be used to identify signatures of these intragenomic conflicts.

1. Introduction

Sexual conflict [1–4] and kin selection [5–9] represent central strands of evolutionary biology, and recent years have seen an explosion of interest in the connection and interplay between the two [10–30]. Much of the theoretical attention that has been devoted to this topic has focused on how the incentive for male harm (i.e. traits that increase a male's mating success at the expense of the females with whom he interacts) may be curbed by relatedness between mates and between mate competitors, in a range of ecologically and demographically explicit population settings [24–29]. These theoretical predictions have motivated a growing body of empirical work on a diversity of organisms including flies [12–15,30], chickens [16], mites [17,18] and beetles [19–21,23].

However, this body of theory has focused on autosomal inheritance and has not considered how ecological and demographic factors shape sexual conflict across the rest of the genome. With recent interest in the evolution of sexual conflict on sex chromosomes [31] and an improving understanding of the molecular basis of harming traits [32], extending this theory to consider how kin selection may differentially mould male harm with respect to non-autosomal portions of the genome is crucial for guiding and interpreting future empirical work. In addition to providing an array of further comparative predictions for populations that differ with respect to ecological and demographic factors, this new theory would also yield comparative predictions at an intragenomic level, which is a particularly powerful approach as within-individual comparisons naturally control for variation in a diversity of confounding factors [33].

To investigate this, we adapt previous models of sexual conflict [24,26,28] for application to autosomal, sex chromosomal and cytoplasmic inheritance. Our analysis encompasses both male (XY and XO) and female (ZW and ZO) heterogametic systems, and therefore we investigate the possibility of male harm loci

occurring on X, Y and Z chromosomes. With respect to cytoplasmic factors, we consider the full range from strictly maternal to strictly paternal inheritance. We first provide a general overview, showing how sex-specific relatedness, reproductive value and the intensity of kin competition combine to determine the potential for male harm. We next analyse a series of ecologically, demographically and genetically explicit models [34,35], revealing how dispersal, overlapping generations, reproductive skew and the mechanism of population regulation modulate sexual conflict across different parts of the genome, and ignite intragenomic conflicts between nuclear and cytoplasmic genes. We then explore the effects of sex biases in these demographic factors, showing how they may drive further intragenomic conflicts between autosomes and sex chromosomes. Finally, we discuss how these theoretical predictions can be tested empirically, including how different crossing schemes may be used to identify signatures of intragenomic conflict.

2. Reproductive value, relatedness and intensity of kin competition modulate the potential for harm

Different portions of the genome flow between the sexes in different ways. These different patterns of transmission may consequently generate differences both in the reproductive values of males and females (i.e. the fraction of the ancestry that flows through them [36–38]), and in their relatedness to same-sex and opposite-sex patchmates. Such differences may therefore alter the value that males place upon their different social partners and thus modulate the evolutionary potential for male harm. To investigate the consequences of different modes of genetic transmission, we follow previous models of sexual conflict [24,26,28], considering a population subdivided into patches with the following life cycle: (1) n_f adult female and n_m adult males reside on each patch, (2) males compete to mate with the females on their patch, (3) females produce broods of offspring, (4) adult males and females die, (5) juveniles compete for breeding spots, with a proportion a of the resulting competition occurring against natal patchmates, and (6) successful juveniles then become adults, starting the life cycle anew. A full description of the life cycle is given in the electronic supplementary material, figure S1.

Within this life cycle, we focus on a harming trait, expressed exclusively by males. This trait increases the relative competitiveness of its bearer (step 2), but decreases the fecundity of the females in his patch (step 3). Possible examples of such behaviour include male harassment, toxic ejaculates and mating plugs [4]. We determine the conditions under which natural selection favours an increase in the level of this harming trait, using the kin-selection approach of Taylor & Frank [39]. This approach analyses how the relative fitness of a focal individual is altered by both small changes in their own trait value and by correlated changes in the trait values of their social partners, with changes in relative fitness weighted by the reproductive value appropriate to their class [38] and the mode of inheritance exhibited by the focal locus [7,40]. See the electronic supplementary material, S1 for full details. This approach assumes weak selection and additivity, and as such it may be less informative for those alleles whose selective effects are particularly strong or highly non-additive. Applying

this methodology, we find that the condition for increase is given by

$$B[(c_{m \rightarrow f} + c_{m \rightarrow m})(1 - r_{mm})] - C[(1 - a_f)(c_{f \rightarrow f} r_{mf} + c_{m \rightarrow f} r_{mm}) + (1 - a_m)(c_{f \rightarrow m} r_{mf} + c_{m \rightarrow m} r_{mm})] > 0, \quad (2.1)$$

where B is the scaled marginal benefit of increased competitiveness enjoyed by the focal individual male; C is the scaled marginal cost of this harm upon the fecundity of the individual females with whom he interacts; r_{ij} is the coefficient of genetic relatedness [5,7] between a sex- i individual and a sex- j individual drawn at random (with replacement) from the same patch (i.e. whole-group relatedness [41,42]); $c_{i \rightarrow j}$ is the class reproductive value [36–38] associated with gene-flow from sex- i parents to sex- j offspring; a_i is the intensity of kin competition, i.e. the probability that sex- i juvenile natal patchmates compete with one another for breeding spots (equivalent to the ‘spatial scale of density-dependent competition’ from [7]); and f and m indicate female and male, respectively.

Inspecting the left-hand side of condition (2.1), we can isolate the distinct selective effects of male harm, and the weightings placed upon them. The first portion captures the inclusive-fitness effect of increased mating success. This includes the direct benefit enjoyed by the focal male from increased mating success B , weighted by the reproductive value he accrues through his daughters $c_{m \rightarrow f}$ and sons $c_{m \rightarrow m}$, minus the concomitant loss of siring success by the average male on his patch (including himself) $-B$, weighted by his relatedness to them r_{mm} and the reproductive value they would have accrued through their daughters $c_{m \rightarrow f}$ and sons $c_{m \rightarrow m}$. The second portion captures the inclusive-fitness effect of increased male harm upon female fecundity. This includes the loss of fecundity of female patchmates $-C$, weighted by the focal male’s relatedness to these females r_{mf} and the reproductive value they would have accrued through their daughters $c_{f \rightarrow f}$ and sons $c_{f \rightarrow m}$, and also the concomitant loss of fecundity of male patchmates who would have sired these lost offspring $-C$, weighted by the focal male’s relatedness to these males r_{mm} and the reproductive value they would have accrued through their daughters $c_{m \rightarrow f}$ and sons $c_{m \rightarrow m}$, with these losses of fecundity being defrayed to the extent a_f and a_m that competition for resources occurs among female and male natal patchmates, respectively.

We may rewrite condition (2.1) in the form $C/B < H$, where the dimensionless quantity H defines the ‘potential for male harm’ [28,43] and is given by:

$$H = \frac{(c_{m \rightarrow f} + c_{m \rightarrow m})(1 - r_{mm})}{(1 - a_f)(c_{f \rightarrow f} r_{mf} + c_{m \rightarrow f} r_{mm}) + (1 - a_m)(c_{f \rightarrow m} r_{mf} + c_{m \rightarrow m} r_{mm})}. \quad (2.2)$$

The potential for harm summarizes the role of ecology, demography and transmission genetics in modulating the evolution of male harm, separate from the role of the more-contingent fecundity cost and benefit, providing a generalization of equation A6 in [28]. A larger potential for male harm means that harm is more likely to be favoured and, if favoured, is expected to be elaborated to a greater degree. Inspecting equation (2.2), we can see that: increasing relatedness (i.e. higher r_{mf} and/or r_{mm}) will typically decrease the potential for male harm; increasing the intensity of kin competition (i.e. higher a_f and/or a_m) will typically increase the potential for male harm, and increasing male reproductive value (i.e.

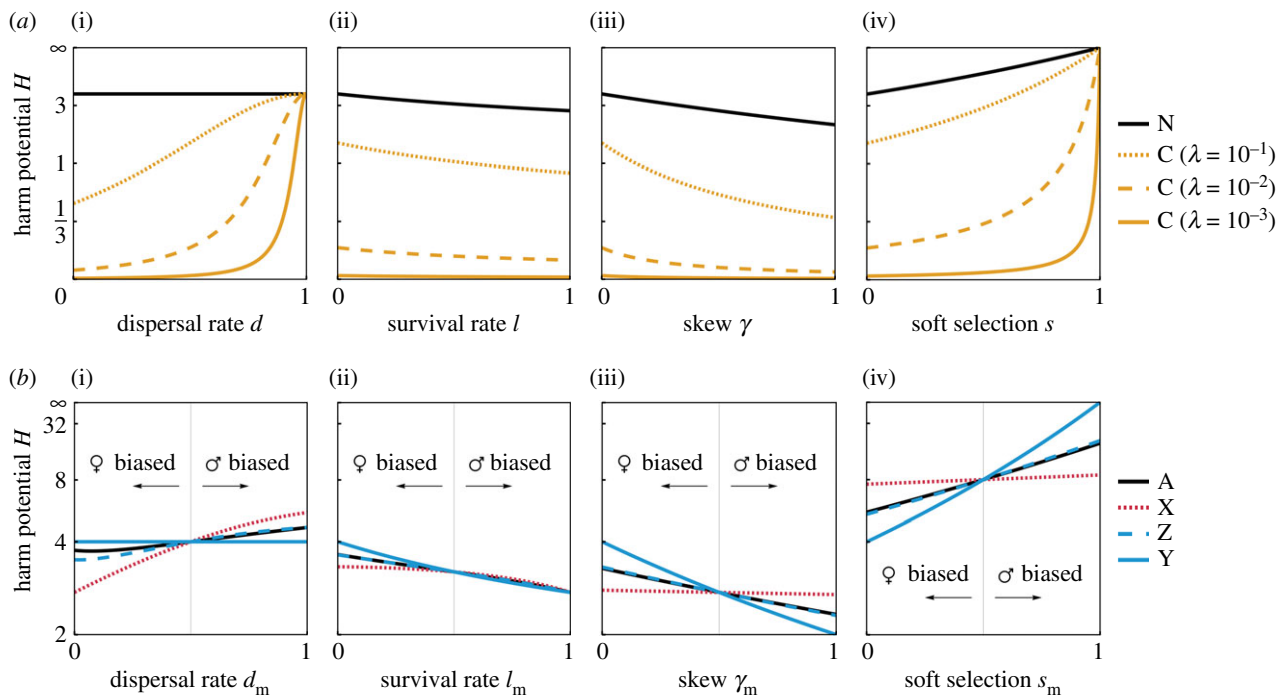


Figure 1. Demography modulates the potential for harm H differently across the genome. (a) Demographic factors modulate harm differently between nuclear (N: autosomes, X, Y, Z) and cytoplasmic genes (C), with differences dependent on the extent of paternal transmission λ . In panels (ii–iv), $d = 0.5$. (b) Sex differences in demographic parameters such as (i) dispersal ($d_f = 0.5$), (ii) survival rate ($l_f = 0.5$), (iii) reproductive skew ($\gamma_f = 0.5$) and (iv) population regulation ($s_f = 0.5$), uncouple the interests of nuclear genes with respect to male harm. In panels (ii–iv) $d_f = d_m = 0.5$. Across all panels $n = 5$. Full methods to recreate these plots can be found in the electronic supplementary material, S2. (Online version in colour.)

higher $c_{m \rightarrow f}$ and/or $c_{m \rightarrow m}$) will typically increase the potential for male harm while increasing female reproductive value (i.e. higher $c_{f \rightarrow f}$ and/or $c_{f \rightarrow m}$) will typically decrease the potential for male harm.

Treating reproductive value, relatedness and kin competition as open parameters is useful for conceptualizing the higher-level forces shaping male harm, and generating comparative results that apply across a wide range of settings [7,34,35]. However, many specific ecological, demographic and genetic factors of interest will modulate several of these parameters simultaneously. For example, assumptions about which genetic party controls the trait, and thus the underlying transmission genetics, will shape both relatedness and reproductive value, and dispersal patterns will alter both relatedness and the intensity of kin competition. To understand how such concrete ecological, demographic and genetic factors will impact upon male harm, we now move from this ‘open’, more general model to a series of ‘closed’, ecologically, demographically and genetically explicit ones [34,35], in which the intensity of kin competition, relatedness coefficients and reproductive values emerge as functions of population processes, life cycle and transmission genetics. Specifically, we now investigate how dispersal, overlapping generations, reproductive skew and the mechanism of population regulation impact the potential for male harm across different parts of the genome.

3. Population viscosity drives conflict between nuclear and cytoplasmic genes

We first investigate how limited dispersal modulates the potential for male harm by considering that a fraction $1 - d$ of juvenile males and females remain on their natal patch, while a fraction d disperse to other patches, prior to both

mating and reproduction (step 5). Lower dispersal increases relatedness between social partners (higher r), but also increases the intensity of competition between their offspring (higher a) [7,41]. For autosomes, as well as X, Y and Z chromosomes, we find that these two effects cancel exactly, such that the potential for male harm is completely independent of the dispersal rate and indeed is given by $H = n - 1$ across all of these genetic systems, where n is the number of males on the patch (figure 1a(i)). That the potential for male harm is the same for autosomes and sex chromosomes under full dispersal ($d = 1$) recovers Andrés & Morrow’s [44] result that there is no intragenomic conflict between these different portions of the genome in the absence of kin selection. Moreover, the invariance of the potential for male harm with respect to dispersal rate was shown previously for autosomes by Rankin [24] and Faria *et al.* [26]. Here we have shown that this invariance extends to the sex chromosomes such that, under the full range of dispersal rates ($0 \leq d \leq 1$), there is no intragenomic conflict with respect to male harm between the autosomes and sex chromosomes.

However, we find that this invariance does not extend to cytoplasmic elements (figure 1a(i); electronic supplementary material, S2.4). Assuming homoplasmy (i.e. that an individual’s cytoplasmic factors are clonally related), and denoting the probability that a cytoplasmic gene is paternally inherited by λ (analogous to the approach of [45]), then we find that this cancellation of increased local competition and increased relatedness only holds in the extremes of strict matrilineal ($\lambda = 0$), exact biparental ($\lambda = 0.5$) or strict patrilineal ($\lambda = 1$) inheritance. Outwith these three cases, the rate of dispersal modulates the potential for harm (figure 1a(ii)), and thus intragenomic conflicts can arise between cytoplasmic and nuclear genes. Under incompletely matrilineally biased inheritance ($0 < \lambda < 0.5$), lower dispersal is associated with

reduced potential for harm, and thus such cytoplasmic elements favour lower levels of harm than do nuclear ones. By contrast, under incompletely patrilineally biased inheritance ($0.5 < \lambda < 1$), lower dispersal is associated with increased potential for harm, such that these elements favour a higher level of harm than do nuclear genes, although in this case the magnitude of the conflict is much smaller.

4. Further demographic factors shape relatedness and kin competition

Above we have shown that, for the various nuclear genes investigated (autosomes, X, Y and Z), limiting the rate of dispersal has no impact upon the potential for harm, owing to the way in which the effect of increased relatedness is perfectly offset by the effect of increased competition. In natural populations, other demographic factors will also typically be present in conjunction with limited dispersal, and together these may shift the balance between relatedness and kin competition, consequently modulating the potential for harm. To investigate this, we consider three further factors: overlapping generations, reproductive skew and soft selection (see electronic supplementary material, S2.1–4 for details).

Allowing adults to survive (and maintain their breeding spots) between generations with probability l (i.e. at step 4 in the life cycle), we find that an increased rate of survival favours lower levels of harm (figure 1a(ii)). This occurs because higher survival increases relatedness between patchmates, but without altering the intensity of kin competition [46]. Consequently, the relatedness and competition effects of dispersal are decoupled in the context of overlapping generations, such that limiting the rate of dispersal leads to higher potential for male harm, but in a way that is exactly equal for autosomes and the various sex chromosomes. Similarly, we find that reproductive skew also decreases the potential for male harm (electronic supplementary material, table S2; figure 1a(iii)). If reproduction is skewed such that a few breeding adults contribute a disproportionate share of the offspring produced on the patch, then this will inflate the relatedness among patchmates while leaving the intensity of kin competition unchanged, thereby reducing the potential for harm. We integrate skew by defining a parameter γ , such that when $\gamma = 0$ all individuals enjoy the same fecundity under neutrality, while in the extreme of $\gamma = 1$, all juveniles share the same parents. We find that as the degree of skew increases, the potential for harm is reduced, and by exactly the same extent for autosomes and sex chromosomes. Decreasing patch size has a similar effect to increasing relatedness, reducing potential for harm (electronic supplementary material, table S2 and figure S2).

Finally, we consider the mechanism by which the population is kept constant in size, and the timing of this regulation step during the life cycle. In particular, we investigate the extent to which it occurs before versus after dispersal by allowing a proportion s of regulation to occur before the dispersal phase and a proportion $1 - s$ of regulation after. Scenarios in which complete regulation occurs before dispersal, such that between-patch differences in productivity are completely abolished, have been described as involving ‘soft selection’ ($s = 1$), whereas scenarios in which complete regulation occurs after dispersal, such that different patches may enjoy differences in productivity, have been described as involving ‘hard selection’ ($s = 0$) [47–49]. Up to now, we have

considered only hard selection ($s = 0$). As we allow the proportion of regulation before dispersal s to increase, so does the extent of kin competition a , and thus the potential for harm, with these effects equivalent for autosomes and sex chromosomes. In the limit of pure soft selection ($s = 1$), decreased female fecundity does not alter the net productivity of the patch, and thus increased harm is always favoured ($H = \infty$; figure 1a(iv); electronic supplementary material, table S2). A fuller description of this life cycle, as well as other approaches to implement the effects of soft selection [50], can be seen in electronic supplementary material, S2.

5. Sex-biased demography drives intragenomic conflict between nuclear genes

Previous work has shown that, for autosomal genes, sex-biased demography may uncouple the balance between relatedness and kin competition [43,51] and consequently may modulate the potential for male harm [26]. Moreover, given their sex-specific inheritance patterns, these effects may be expected to be manifest differently across autosomes and sex chromosomes, thereby potentially uncoupling their inclusive-fitness interests and driving intragenomic conflicts of interest. To investigate this, we allow for sex biases in the rate of dispersal and survival, and in the degree of reproductive skew and soft selection.

We find that sex-biased dispersal ($d_f \neq d_m$) leads to a divergence between the inclusive-fitness interests of autosomes and sex chromosomes (figure 1b(i)). Typically, under male-biased dispersal, the potential for male harm is greatest for X chromosomes and lowest for Y chromosomes ($H_X > H_Z > H_A > H_Y$). Conversely, under female-biased dispersal this ranking is usually reversed ($H_X < H_Z < H_A < H_Y$), and across these parameter values the Y chromosome remains invariant with respect to dispersal. However, this ranking of harm potential does not hold perfectly across all parameter values. For example, autosomes have the highest potential for male harm under high female-dispersal and low male-dispersal regimes. These complex patterns arise because sex-biased dispersal alters both the relatedness structure arising through matrilineal and patrilineal [27,52], and the intensity of competition experienced by daughters and sons. Owing to the sex-specific transmission patterns, these effects are felt differently by the different genomic elements. For instance, when male dispersal is low, kin competition is intense among sons relative to daughters, but patrilineal and matrilineal relatedness increase more evenly. This has a bigger harm-reducing effect for X chromosomes which are primarily transmitted through daughters (and thus experience a lesser increase in kin competition relative to the increase in relatedness).

Under sex-biased survival ($l_f \neq l_m$), we find that sex chromosomes and autosomes once again diverge in their inclusive-fitness interests (figure 1b(ii)). With female-biased survival ($l_f > l_m$), relatedness is higher through matrilineal than patrilineal; and with male-biased survival ($l_f < l_m$), relatedness is higher through patrilineal than matrilineal [53]. Higher matrilineal relatedness has a greater impact upon those genomic elements for which a greater fraction are maternally inherited and thus the potential for harm is highest for Y chromosomes and lowest for X chromosomes ($H_Y > H_Z > H_A > H_X$). The reverse is true when there is higher patrilineal relatedness, with harm lowest for Y chromosomes and highest for X chromosomes ($H_Y < H_Z < H_A < H_X$). The same qualitative

pattern also obtains under sex differences in reproductive skew ($\gamma_f \neq \gamma_m$; figure 1b(iii)). When skew is higher in females ($\gamma_f > \gamma_m$) then there is higher matrilineal relatedness, and thus harm is highest for Y chromosomes and lowest for X chromosomes ($H_Y > H_Z > H_A > H_X$), and when skew is higher in males ($\gamma_f < \gamma_m$), then there is higher patrilineal relatedness, and thus harm is lowest for Y chromosomes and highest for X chromosomes ($H_Y < H_Z < H_A < H_X$). Similarly, sex biases in the number of breeders shapes relatedness. If there are more male than female breeders ($n_f < n_m$), then relatedness is higher through matrilineal than patrilineal, while if there are more female than male breeders ($n_f > n_m$), then relatedness is higher through patrilineal than matrilineal, with similar consequences as before on the potential for harm.

Finally, we find that the inclusive-fitness interests of autosomes and sex chromosomes also diverge as a consequence of sex-biased soft selection ($s_f \neq s_m$; figure 1b(iv)). If females experience a higher degree of soft selection, $s_f > s_m$, then kin competition is more intense among daughters than among sons. Conversely, if males experience a higher degree of soft selection, $s_f < s_m$, then the reverse obtains. Greater competition between same-sex relatives promotes harm more for those elements which achieve relatively higher reproductive value through that sex. Accordingly, when the degree of soft selection is greater in females then the potential for harm is lowest for Y chromosomes and highest for X chromosomes ($H_Y < H_Z < H_A < H_X$), and when the degree of soft selection is greater in males, then the potential for harm is highest for Y chromosomes and lowest for X chromosomes ($H_Y > H_Z > H_A > H_X$). Alongside these potentials for harm, we also analyse an example with specific male and female fecundity functions in electronic supplementary material, §4, explicitly solving for the optimum harm value across different loci (electronic supplementary material, figures S3–S5).

6. Discussion

Male harming traits have been described across a wide range of taxa [4], from traumatic insemination of bed bugs [54], and grasping appendages of water striders [55,56], to proteins in the ejaculates of flatworms [57], and tomiodonts of painted turtles [58]. Recent theory has shown how kin selection may curb the worst excesses of such male harm [24–27,29] and has been supported empirically in a growing range of taxa, including arachnids, birds and insects [12–21,23,30]. We have built upon this theory to show how aspects of demography may shape the potential for male harm differently across different parts of the genome, yielding novel predictions as to how intragenomic conflicts may emerge over such traits, where male harm loci are likely to be enriched, and how these patterns are expected to vary across different populations and species.

In particular, we have found that cytoplasmic genes may favour distinct levels of harm to their nuclear counterparts. As matrilineal inheritance of both cytoplasmic genes and other endosymbionts is the norm across the animal and plant kingdoms [59], our analysis suggests that these elements tend to favour lower levels of male harm than do nuclear genes, generating potentially intense intragenomic conflicts over such traits. One particular arena of conflict may be over sperm competitiveness, as while competitive sperm may provide a benefit to the focal male, they may also impose fecundity costs for

females, for example through zygote inviability owing to polyspermy [60,61]. Given the central role mitochondria play in sperm physiology, there may be a large scope for conflict in this context. Indeed, while mitochondrial alleles contributing to variation in sperm performance are typically assumed to be the products of drift [62–64] (i.e. ‘mother’s curse’ [65,66]), in viscous populations such alleles may be positively selected if they reduce the fitness costs imposed upon interacting female kin (this is a negative variant of the argument made by [67]). It also mirrors the evolution of male-killing symbionts in various arthropod groups [68]; in both cases, alleles which decrease a male’s fitness may improve the fitness of his female relatives, by either reducing the extent of male harm or decreasing the intensity of juvenile competition for resources.

Although matrilineal inheritance is the norm for cytoplasmic elements, various exceptions—such as the doubly uniparental inheritance of bivalve molluscs [69], paternal transmission of mitochondria in cucumbers and sequoias [70] and paternal transmission of symbionts in mosquitos [71], leafhoppers [72] and tsetse flies [73]—provide exciting avenues for further empirical testing, with non-matrilineally inherited genes expected to exhibit greater harm than those with strict matrilineal inheritance. Although the above examples are somewhat speculative, one example which may be more amenable to experimental investigation is the obligate vertically transmitted rhabdovirus sigma. This is biparentally transmitted in *Drosophila melanogaster* [74], and experiments have shown that males infected with sigma appear to have increased mating success [75], although the mechanism of action and direct cost to females (if any) is unclear. Given that sigma viruses infect other arthropods and appear to show similar transmission patterns [76], this system may be amenable for comparisons across different transmission patterns and demographic scenarios, as well as to experimental manipulation of these factors. Moreover, although we have made a conceptual distinction between nuclear versus cytoplasmic genes, there are nonetheless nuclear genes whose inheritance patterns more closely match those of cytoplasmic factors, and to which our predictions for cytoplasmic genes may readily apply. For instance, the germline-restricted chromosome in zebra finches is maternally transmitted, with rare occurrences of paternal transmission [77,78]. Depending on which tissues such genes are present in, and the extent of their expression, then these too may have the potential to modulate male harm and come into conflict with other genes inhabiting the same nuclei.

We have also shown that while the potential for harm is constant across the nuclear genes under sex-neutral demography, population viscosity in concert with sex-biased demography generates differences in how male harm loci evolve on autosomes and sex chromosomes. This shares conceptual similarities with how the potential for altruistic behaviour remains invariant across diploidy and haplodiploidy under sex-neutral dispersal but diverges under sex-biased dispersal [41,51,79]. This yields predictions about where male harm loci should be enriched across the genome, and how such patterns will depend on both the extent and direction of sex biases in demography. Currently, these predictions are challenging to test, as the genetics of many male harm traits is still poorly understood [32]. As Rowe *et al.* point out [32], there are cases where the phenotypic interactions are well understood, but the genetics is not, and cases where the genetics is well understood, but the phenotypic interactions are not. Nonetheless, there are an increasing number of examples that span this

gap, including genes underpinning the morphological aspects of grasping behaviours in water striders [80,81], metabolic genes associated with siring success in bulb mites [82–84], and gamete-recognition proteins in abalone and sea urchins [85,86]. Alongside these specific examples, there are classes of male harm traits that may be particularly amenable to large-scale genetic analysis. For example, sperm fluid proteins (Sfps) currently represent among the most-successful syntheses of the genetics and ecology of sexual conflict [87]. Though even here we lack detailed knowledge of what many of these proteins do, and whether or not they are definitively involved in sexual conflict. For instance, while the *Drosophila melanogaster* seminal proteome is well-characterized in comparison with many others, we still only have a good functional understanding of about 10% of its constituent proteins [88]. As the proteomes of more species have begun to be characterized, and those proteomes functionally described, then hopefully these within-genome comparisons will be increasingly tractable.

In addition to studies of natural populations, experimental evolution offers ways to artificially generate particular population structures and thereby investigate their effects on male harm. Previously, such approaches have been used in bulb mites [17], spider mites [18] and seed beetles [20,23]. While spider mites are arrhenotokous and lack sex chromosomes, both bulb mites (XO) and seed beetles (XY) have sex chromosomes and therefore may be systems in which the predictions we have outlined could be most effectively tested. Additionally, as well as manipulating population structures, previous studies in *D. melanogaster* have used balancer chromosomes to manipulate the inheritance patterns experienced by X chromosomes, enforcing either strictly matrilineal or strictly patrilineal inheritance (e.g. [89,90]). Accordingly, combined with manipulated population structures, one could effectively force Y-chromosomal inheritance on the X, and thus partially control for the historical gene content of these different chromosomes, as well as enabling experimental exploration of the intervening parameter space.

Even if individual trait levels are dominated by the interests of autosomal genes—as might be expected if they are the largest genomic fraction [91–93]—intra-genomic conflicts may still be expected to escalate and lead to differences in the abundance of ‘harm-promoting’ and ‘harm-inhibiting’ loci across different portions of the genome [27,94]. Population crosses can provide one method to identify these signatures of intra-genomic conflict, by creating ‘imbalanced’ genomes with a relative abundance or paucity of harm-promoting or harm-inhibiting loci (see electronic supplementary material, S4) [27]. For instance, evidence of conflicts between maternal-origin and paternal-origin genes have been found by performing reciprocal crosses in flowering plants [95–99], mammals [100–102] and insects [103–105]. Similar approaches may be used to uncover the intra-genomic conflicts we have outlined here. In electronic supplementary material, figure S6, we present two examples of the phenotypes predicted for a cross between two populations with an XO sex-determination system. If these two populations differ in either the direction or intensity of conflict, then crosses between them are expected to lead to extreme phenotypes as the delicate balance between competing genomic factions is disrupted, with reciprocal directions of cross expected to lead to opposite phenotypes. Given the extreme phenotypes that are expected to arise from population crosses, this may be a further mechanism by which intra-genomic conflict contributes to hybrid inviability and hence speciation [106].

Our analysis has focused on cases where males reduce the immediate fecundity of interacting females, but population structure is likely to uncouple the interests of different genomic factions in relation to a wider set of both inter- and intra-sexual social traits. This may include other forms of male harm, for example whereby harm reduces longevity and/or future fecundity or reduces fecundity unevenly across the females in the group [28,107]. While some of these effects are already incorporated in our model—for instance, when there is no survival of females between generations ($l_f = 0$), a reduction in female reproductive success could be interpreted as owing to loss of fecundity or alternatively premature death before the completion of reproductive effort—more generally these different assumptions about the ecology of harm will probably alter results, at least quantitatively if not qualitatively. Moreover, we have only investigated the selective pressures shaping male harm, and thus have not considered possible coevolutionary dynamics between male harm and female resistance. Previous analyses of these coevolutionary processes have typically focused on autosomal inheritance (e.g. [24,26,27,108,109]) but the inclusion of other genomic elements may well lead to divergent results, because—as we have shown—these elements may have distinct evolutionary interests. Coevolutionary dynamics will therefore probably be sensitive to assumptions about demography, but also to how control over the phenotype is dispersed across the genome and the relative phenotypic power these genomic elements have in males and females. Recent theoretical and empirical work in *D. melanogaster* has shown that sexually antagonistic coevolution of the sex chromosomes may also play an important role in speciation [31], and thus we may expect the intra-genomic conflicts that we have outlined here to further contribute to the origin of species [106].

To conclude, we have shown how ecological and demographic processes—and, in particular, their sex-specific aspects—may differentially mould male harm across the various inheritance systems that coexist within individual genomes. With differences in the flow of genes from mothers and fathers to daughters and sons, differences in relatedness to social partners and the intensity of kin competition may emerge, igniting conflicts of interest between autosomes, sex chromosomes and cytoplasmic elements over male harm. As knowledge of the molecular basis of sexual conflict grows—from flies and water striders to abalone and sea urchins—these models may help guide the design of future experiments and aid in the interpretation of data collected from natural populations.

Data accessibility. The data are provided in the electronic supplementary material [110].

Authors' contributions. T.J.H.: conceptualization, formal analysis, investigation, methodology, project administration, validation, visualization, writing—original draft, writing—review and editing; A.G.: conceptualization, funding acquisition, methodology, resources, supervision, writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Competing interests. The authors declare no conflicts of interest.

Funding. T.J.H. is supported by a PhD scholarship funded by the School of Biology, University of St Andrews. A.G. is supported by a Natural Environment Research Council Independent Research Fellowship (grant no. NE/K009524/1) and a European Research Council Consolidator (grant no. 771387).

Acknowledgements. We thank G. Faria, V. Litzke, J. Rayner, M. Ritchie, D. Shuker and K. Stucky for helpful discussion, and two anonymous reviewers for constructive comments.

References

- Trivers R. 1972 Parental investment and sexual selection. In *Sexual selection and the descent of man* (ed. BG Campbell), pp. 136–179. Chicago, IL: Aldine Publishing Company.
- Parker GA. 1979 Sexual selection and sexual conflict. *Sex. Sel. Reprod. Compet. insects* **123**, 166.
- Charnov EL. 1979 Simultaneous hermaphroditism and sexual selection. *Proc. Natl Acad. Sci. USA* **76**, 2480–2484. (doi:10.1073/pnas.76.5.2480)
- Arnqvist G, Rowe L. 2005 *Sexual conflict*. Princeton, NJ: Princeton University Press.
- Hamilton WD. 1964 The genetical evolution of social behaviour. I. *J. Theor. Biol.* **7**, 1–16. (doi:10.1016/0022-5193(64)90038-4)
- Hamilton WD. 1964 The genetical evolution of social behaviour. II. *J. Theor. Biol.* **7**, 17–52. (doi:10.1016/0022-5193(64)90039-6)
- Frank SA. 1998 *Foundations of social evolution*. Princeton, NJ: Princeton University Press.
- Rousset F. 2004 *Genetic structure and selection in subdivided populations (MPB-40)*. Princeton, NJ: Princeton University Press.
- Bourke A. 2011 *Principles of social evolution*. Oxford, UK: Oxford University Press.
- Bourke AFG. 2009 The kin structure of sexual interactions. *Biol. Lett.* **5**, 689–692. (doi:10.1098/rsbl.2009.0146)
- Pizzari T, Gardner A. 2012 The sociobiology of sex: inclusive fitness consequences of inter-sexual interactions. *Phil. Trans. R. Soc. B* **367**, 2314–2323. (doi:10.1098/rstb.2011.0281)
- Hollis B, Kawecki TJ, Keller L. 2015 No evidence that within-group male relatedness reduces harm to females in *Drosophila*. *Ecol. Evol.* **5**, 979–983. (doi:10.1002/ece3.1417)
- Chippindale AK, Berggren M, Alpern JHM, Montgomerie R. 2015 Does kin selection moderate sexual conflict in *Drosophila*? *Proc. R. Soc. B* **282**, 20151417. (doi:10.1098/rspb.2015.1417)
- Martin ES, Long TAF. 2015 Are flies kind to kin? The role of intra- and inter-sexual relatedness in mediating reproductive conflict. *Proc. R. Soc. B* **282**, 20151991. (doi:10.1098/rspb.2015.1991)
- Le Page S, Sepil I, Flinham E, Pizzari T, Carazo P, Wigby S. 2017 Male relatedness and familiarity are required to modulate male-induced harm to females in *Drosophila*. *Proc. R. Soc. B* **284**, 20170441. (doi:10.1098/rspb.2017.0441)
- Tan CKW, Doyle P, Bagshaw E, Richardson DS, Wigby S, Pizzari T. 2017 The contrasting role of male relatedness in different mechanisms of sexual selection in red junglefowl. *Evolution (NY)* **71**, 403–420. (doi:10.1111/evo.13145)
- Łukasiewicz A, Szubert-Kruszyńska A, Radwan J. 2017 Kin selection promotes female productivity and cooperation between the sexes. *Sci. Adv.* **3**, e1602262. (doi:10.1126/sciadv.1602262)
- Rodrigues L, Torralba Sáez M, Alpedrinha J, Lefèvre S, Brengues M, Magalhães S, Duncan AB. 2021 Consequences of population structure for sex allocation and sexual conflict. *J. Evol. Biol.* **34**, 525–536. (doi:10.1111/jeb.13755)
- Lymbery SJ, Simmons LW. 2017 Males harm females less when competing with familiar relatives. *Proc. R. Soc. B* **284**, 20171984. (doi:10.1098/rspb.2017.1984)
- Lymbery SJ, Wyber B, Tomkins JL, Simmons LW. 2020 No evidence for divergence in male harmfulness or female resistance in response to changes in the opportunity for dispersal. *J. Evol. Biol.* **33**, 966–978. (doi:10.1111/jeb.13628)
- Berg EC, Lind MI, Monahan S, Bricout S, Maklakov AA. 2019 Kin but less than kind: within-group male relatedness does not increase female fitness in seed beetles. *Proc. R. Soc. B* **286**, 20191664. (doi:10.1098/rspb.2019.1664)
- Pizzari T, Biernaskie JM, Carazo P. 2015 Inclusive fitness and sexual conflict: how population structure can modulate the battle of the sexes. *Bioessays* **37**, 155–166. (doi:10.1002/bies.201400130)
- Rodriguez-Exposito E, Garcia-Gonzalez F. 2021 Metapopulation structure modulates sexual antagonism. *Evol. Lett.* **5**, 344–358. (doi:10.1002/evl3.244)
- Rankin DJ. 2011 Kin selection and the evolution of sexual conflict. *J. Evol. Biol.* **24**, 71–81. (doi:10.1111/j.1420-9101.2010.02143.x)
- Wild G, Pizzari T, West SA. 2011 Sexual conflict in viscous populations: the effect of the timing of dispersal. *Theor. Popul. Biol.* **80**, 298–316. (doi:10.1016/j.tpb.2011.09.002)
- Faria GS, Varela SAM, Gardner A. 2015 Sex-biased dispersal, kin selection and the evolution of sexual conflict. *J. Evol. Biol.* **28**, 1901–1910. (doi:10.1111/jeb.12697)
- Faria GS, Varela SAM, Gardner A. 2017 Sexual selection modulates genetic conflicts and patterns of genomic imprinting. *Evolution (NY)* **71**, 526–540. (doi:10.1111/evo.13153)
- Faria GS, Gardner A, Carazo P. 2020 Kin discrimination and demography modulate patterns of sexual conflict. *Nat. Ecol. Evol.* **4**, 1141–1148. (doi:10.1038/s41559-020-1214-6)
- Iritani R. 2020 Gametophytic competition games among relatives: when does spatial structure select for facilitativeness or competitiveness in pollination? *J. Ecol.* **108**, 1–13. (doi:10.1111/1365-2745.13282)
- Carazo P, Tan CKW, Allen F, Wigby S, Pizzari T. 2014 Within-group male relatedness reduces harm to females in *Drosophila*. *Nature* **505**, 672–675. (doi:10.1038/nature12949)
- Lund-Hansen KK, Olito C, Morrow EH, Abbott JK. 2021 Sexually antagonistic coevolution between the sex chromosomes of *Drosophila melanogaster*. *Proc. Natl Acad. Sci. USA* **118**, e2003359118. (doi:10.1073/pnas.2003359118)
- Rowe L, Chenoweth SF, Agrawal AF. 2018 The genomics of sexual conflict. *Am. Nat.* **192**, 274–286. (doi:10.1086/698198)
- Rautiala P, Gardner A. 2016 Intragenomic conflict over soldier allocation in polyembryonic parasitoid wasps. *Am. Nat.* **187**, E106–E115. (doi:10.1086/685082)
- Gardner A, West SA. 2006 Demography, altruism, and the benefits of budding. *J. Evol. Biol.* **19**, 1707–1716. (doi:10.1111/j.1420-9101.2006.01104.x)
- Cooper GA, Levin SR, Wild G, West SA. 2018 Modeling relatedness and demography in social evolution. *Evol. Lett.* **2**, 260–271. (doi:10.1002/evl3.69)
- Fisher RA. 1930 *The genetical theory of natural selection*. London, UK: The Clarendon Press.
- Taylor PD. 1990 Allele-frequency change in a class-structured population. *Am. Nat.* **135**, 95–106.
- Grafen A. 2006 A theory of Fisher's reproductive value. *J. Math. Biol.* **53**, 15–60. (doi:10.1007/s00285-006-0376-4)
- Taylor PD, Frank SA. 1996 How to make a kin selection model. *J. Theor. Biol.* **180**, 27–37. (doi:10.1006/jtbi.1996.0075)
- Taylor PD. 1996 Inclusive fitness arguments in genetic models of behaviour. *J. Math. Biol.* **34**, 654–674. (doi:10.1007/BF02409753)
- Taylor PD. 1992 Altruism in viscous populations—an inclusive fitness model. *Evol. Ecol.* **6**, 352–356. (doi:10.1007/BF02270971)
- Pepper JW. 2000 Relatedness in trait group models of social evolution. *J. Theor. Biol.* **206**, 355–368. (doi:10.1006/jtbi.2000.2132)
- Gardner A. 2010 Sex-biased dispersal of adults mediates the evolution of altruism among juveniles. *J. Theor. Biol.* **262**, 339–345. (doi:10.1016/j.jtbi.2009.09.028)
- Andrés JA, Morrow EH. 2003 The origin of interlocus sexual conflict: is sex-linkage important? *J. Evol. Biol.* **16**, 219–223. (doi:10.1046/j.1420-9101.2003.00525.x)
- Kuijper B, Lane N, Pomiankowski A. 2015 Can paternal leakage maintain sexually antagonistic polymorphism in the cytoplasm? *J. Evol. Biol.* **28**, 468–480. (doi:10.1111/jeb.12582)
- Taylor PD, Irwin AJ. 2000 Overlapping generations can promote altruistic behavior. *Evolution (NY)* **54**, 1135–1141. (doi:10.1111/j.0014-3820.2000.tb00549.x)
- Wallace B. 1968 Polymorphism, population size, and genetic load. In *Population biology and evolution* (ed. RC Lewontin), pp. 87–108. Syracuse, NY: Syracuse University Press.
- Christiansen FB. 1975 Hard and soft selection in a subdivided population. *Am. Nat.* **109**, 11–16.
- Wade MJ. 1985 Soft selection, hard selection, kin selection, and group selection. *Am. Nat.* **125**, 61–73. (doi:10.1086/284328)
- Débarre F, Gandon S. 2011 Evolution in heterogeneous environments: between soft and hard selection. *Am. Nat.* **177**, E84–E97. (doi:10.1086/658178)

51. Johnstone RA, Cant MA. 2008 Sex differences in dispersal and the evolution of helping and harming. *Am. Nat.* **172**, 318–330. (doi:10.1086/589899)
52. Haig D. 2000 Genomic imprinting, sex-biased dispersal, and social behavior. *Ann. NY Acad. Sci.* **907**, 149–163. (doi:10.1111/j.1749-6632.2000.tb06621.x)
53. Van Cleve J, Feldman MW, Lehmann L. 2010 How demography, life history, and kinship shape the evolution of genomic imprinting. *Am. Nat.* **176**, 440–455. (doi:10.1086/656277)
54. Tataric NJ, Cassis G, Siva-Jothy MT. 2014 Traumatic insemination in terrestrial arthropods. *Annu. Rev. Entomol.* **59**, 245–261. (doi:10.1146/annurev-ento-011613-162111)
55. Arnqvist G, Rowe L. 2002 Correlated evolution of male and female morphologies in water striders. *Evolution (NY)* **56**, 936–947. (doi:10.1111/j.0014-3820.2002.tb01406.x)
56. Perry JC, Rowe L. 2012 Sexual conflict and antagonistic coevolution across water strider populations. *Evolution (NY)* **66**, 544–557. (doi:10.1111/j.1558-5646.2011.01464.x)
57. Patlar B, Weber M, Temizyürek T, Ramm SA. 2020 Seminal fluid-mediated manipulation of post-mating behavior in a simultaneous hermaphrodite. *Curr. Biol.* **30**, 143–149.e4. (doi:10.1016/j.cub.2019.11.018)
58. Moldovan PD, Brooks RJ, Litzgus JD. 2020 Sex, shells, and weaponry: coercive reproductive tactics in the painted turtle, *Chrysemys picta*. *Behav. Ecol. Sociobiol.* **74**, 142. (doi:10.1007/s00265-020-02926-w)
59. Birky CW. 2001 The inheritance of genes in mitochondria and chloroplasts: laws, mechanisms, and models. *Annu. Rev. Genet.* **35**, 125–148. (doi:10.1146/annurev.genet.35.102401.090231)
60. Frank S. 2000 Sperm competition and female avoidance of polyspermy mediated by sperm-egg biochemistry. *Evol. Ecol. Res.* **2**, 613–625.
61. Edward DA, Stockley P, Hosken DJ. 2015 Sexual conflict and sperm competition. *Cold Spring Harb. Perspect. Biol.* **7**, a017707. (doi:10.1101/cshperspect.a017707)
62. Padua MV, Zeh DW, Bonilla MM, Zeh JA. 2014 Sisters' curse: sexually antagonistic effects constrain the spread of a mitochondrial haplogroup superior in sperm competition. *Proc. R. Soc. B* **281**, 20141686. (doi:10.1098/rspb.2014.1686)
63. Feng GF, Zhang J, Feng LM, Shen NX, Li LJ, Zhu YM. 2013 Mitochondrial DNA haplogroup associated with sperm motility in the Han population. *Asian J. Androl.* **15**, 630–633. (doi:10.1038/aja.2013.83)
64. Smith S, Turbill C, Suchentrunk F. 2010 Introducing mother's curse: low male fertility associated with an imported mtDNA haplotype in a captive colony of brown hares. *Mol. Ecol.* **19**, 36–43. (doi:10.1111/j.1365-294X.2009.04444.x)
65. Frank SA, Hurst LD. 1996 Mitochondria and male disease. *Nature* **383**, 224. (doi:10.1038/383224a0)
66. Gemmill NJ, Metcalf VJ, Allendorf FW. 2004 Mother's curse: the effect of mtDNA on individual fitness and population viability. *Trends Ecol. Evol.* **19**, 238–244. (doi:10.1016/j.tree.2004.02.002)
67. Wade MJ, Brandvain Y. 2009 Reversing mother's curse: selection on male mitochondrial fitness effects. *Evolution (NY)* **63**, 1084–1089. (doi:10.1111/j.1558-5646.2009.00614.x)
68. Hurst GDD, Frost CL. 2015 Reproductive parasitism: maternally inherited symbionts in a biparental world. *Cold Spring Harb. Perspect. Biol.* **7**, a017699. (doi:10.1101/cshperspect.a017699)
69. Ghiselli F, Gomes-dos-Santos A, Adema CM, Lopes-Lima M, Sharbrough J, Boore JL. 2021 Molluscan mitochondrial genomes break the rules. *Phil. Trans. R. Soc. B* **376**, 2020.0159. (doi:10.1098/rstb.2020.0159)
70. Breton S, Stewart DT. 2015 Atypical mitochondrial inheritance patterns in eukaryotes. *Genome* **58**, 423–431. (doi:10.1139/gen-2015-0090)
71. Damiani C *et al.* 2008 Paternal transmission of symbiotic bacteria in malaria vectors. *Curr. Biol.* **18**, R1087–R1088. (doi:10.1016/j.cub.2008.10.040)
72. Watanabe K, Yukuhiro F, Matsuura Y, Fukatsu T, Noda H. 2014 Intrasperm vertical symbiont transmission. *Proc. Natl Acad. Sci. USA* **111**, 7433–7437. (doi:10.1073/pnas.1402476111)
73. De Vooght L, Caljon G, Van Hees J, Van Den Abbeele J. 2015 Paternal transmission of a secondary symbiont during mating in the viviparous tsetse fly. *Mol. Biol. Evol.* **32**, 1977–1980. (doi:10.1093/molbev/msv077)
74. Longdon B, Jiggins FM. 2012 Vertically transmitted viral endosymbionts of insects: do sigma viruses walk alone? *Proc. R. Soc. B* **279**, 3889–3898. (doi:10.1098/rspb.2012.1208)
75. Rittschof CC, Pattanaik S, Johnson L, Matos LF, Brusini J, Wayne ML. 2013 Sigma virus and male reproductive success in *Drosophila melanogaster*. *Behav. Ecol. Sociobiol.* **67**, 529–540. (doi:10.1007/s00265-012-1472-7)
76. Longdon B *et al.* 2017 Vertically transmitted rhabdoviruses are found across three insect families and have dynamic interactions with their hosts. *Proc. R. Soc. B* **284**, 20162381. (doi:10.1098/rspb.2016.2381)
77. Pigozzi MI, Solari AJ. 2005 The germ-line-restricted chromosome in the zebra finch: recombination in females and elimination in males. *Chromosoma* **114**, 403–409. (doi:10.1007/s00412-005-0025-5)
78. Pei Y *et al.* 2021 Occasional paternal inheritance of the germline-restricted chromosome in songbirds. bioRxiv, 2021.01.28.428604. (doi:10.1101/2021.01.28.428604)
79. Johnstone RA, Cant MA, Field J. 2012 Sex-biased dispersal, haplodiploidy and the evolution of helping in social insects. *Proc. R. Soc. B* **279**, 787–793.
80. Khila A, Abouheif E, Rowe L. 2012 Function, developmental genetics, and fitness consequences of a sexually antagonistic trait. *Science* **336**, 585–589. (doi:10.1126/science.1217258)
81. Crumière AJJ, Khila A. 2019 Hox genes mediate the escalation of sexually antagonistic traits in water striders. *Biol. Lett.* **15**, 20180720. (doi:10.1098/rsbl.2018.0720)
82. Konior M, Radwan J, Kołodziejczyk M, Keller L. 2006 Strong association between a single gene and fertilization efficiency of males and fecundity of their mates in the bulb mite. *Proc. R. Soc. B* **273**, 309–314. (doi:10.1098/rspb.2005.3302)
83. Skwierzyńska AM, Plesnar-Bielak A. 2018 Proximate mechanisms of the differences in reproductive success of males bearing different alleles of Pgdh—a gene involved in a sexual conflict in bulb mite. *J. Evol. Biol.* **31**, 657–664. (doi:10.1111/jeb.13250)
84. Plesnar-Bielak A, Skwierzyńska AM, Radwan J. 2020 Sexual and ecological selection on a sexual conflict gene. *J. Evol. Biol.* **33**, 1433–1439. (doi:10.1111/jeb.13680)
85. Clark NL, Gasper J, Sekino M, Springer SA, Aquadro CF, Swanson WJ. 2009 Coevolution of interacting fertilization proteins. *PLoS Genet.* **5**, e1000570. (doi:10.1371/journal.pgen.1000570)
86. Levitan DR, Buchwalter R, Hao Y. 2019 The evolution of gametic compatibility and compatibility groups in the sea urchin *Mesocentrotus franciscanus*: an avenue for speciation in the sea. *Evolution (NY)* **73**, 1428–1442. (doi:10.1111/evo.13766)
87. Sirot LK, Wong A, Chapman T, Wolfner MF. 2015 Sexual conflict and seminal fluid proteins: a dynamic landscape of sexual interactions. *Cold Spring Harb. Perspect. Biol.* **7**, a017533.
88. Wigby S, Brown NC, Allen SE, Misra S, Sitnik JL, Sepil I, Clark AG, Wolfner MF. 2020 The *Drosophila* seminal proteome and its role in postcopulatory sexual selection: *Drosophila* seminal fluid proteins. *Phil. Trans. R. Soc. B* **375**, 20200072. (doi:10.1098/rstb.2020.0072rstb20200072)
89. Lund-Hansen KK, Abbott JK, Morrow EH. 2020 Feminization of complex traits in *Drosophila melanogaster* via female-limited X chromosome evolution. *Evolution (NY)* **74**, 2703–2713. (doi:10.1111/evo.14021)
90. Abbott JK, Chippindale AK, Morrow EH. 2020 The microevolutionary response to male-limited X-chromosome evolution in *Drosophila melanogaster* reflects macroevolutionary patterns. *J. Evol. Biol.* **33**, 738–750. (doi:10.1111/jeb.13618)
91. Leigh EG. 1971 *Adaptation and diversity*. San Francisco, CA: Freeman, Cooper and Company.
92. Gardner A, Ross L. 2014 Mating ecology explains patterns of genome elimination. *Ecol. Lett.* **17**, 1602–1612. (doi:10.1111/ele.12383)
93. Scott TW, West SA. 2019 Adaptation is maintained by the parliament of genes. *Nat. Commun.* **10**, 5163. (doi:10.1038/s41467-019-13169-3)
94. Haig D. 1996 Placental hormones, genomic imprinting, and maternal-fetal communication. *J. Evol. Biol.* **9**, 357–380. (doi:10.1046/j.1420-9101.1996.9030357.x)
95. Rebernick CA, Lafon-Placette C, Hatorangan MR, Slotte T, Köhler C. 2015 Non-reciprocal interspecies hybridization barriers in the *Capsella* genus are

- established in the endosperm. *PLoS Genet.* **11**, e1005295. (doi:10.1371/journal.pgen.1005295)
96. Lafon-Placette C *et al.* 2017 Endosperm-based hybridization barriers explain the pattern of gene flow between *Arabidopsis lyrata* and *Arabidopsis arenosa* in Central Europe. *Proc. Natl Acad. Sci. USA* **114**, E1027–E1035. (doi:10.1073/pnas.1615123114)
97. Florez-Rueda AM, Paris M, Schmidt A, Widmer A, Grossniklaus U, Städler T. 2016 Genomic imprinting in the endosperm is systematically perturbed in abortive hybrid tomato seeds. *Mol. Biol. Evol.* **33**, 2935–2946. (doi:10.1093/molbev/msw175)
98. Coughlan JM, Brown MW, Willis JH. 2020 Patterns of hybrid seed inviability in the *Mimulus guttatus* sp. complex reveal a potential role of parental conflict in reproductive isolation. *Curr. Biol.* **30**, 83–93.
99. He H, Yokoi S, Tezuka T. 2020 A high maternal genome excess causes severe seed abortion leading to ovary abscission in *Nicotiana* interploidy-interspecific crosses. *Plant Direct* **4**, e00257. (doi:10.1002/pld3.257)
100. Brekke TD, Good JM. 2014 Parent-of-origin growth effects and the evolution of hybrid inviability in dwarf hamsters. *Evolution (NY)* **68**, 3134–3148. (doi:10.1111/evo.12500)
101. Brekke TD, Henry LA, Good JM. 2016 Genomic imprinting, disrupted placental expression, and speciation. *Evolution (NY)* **70**, 2690–2703. (doi:10.1111/evo.13085)
102. Vrana PB, Guan XJ, Ingram RS, Tilghman SM. 1998 Genomic imprinting is disrupted in interspecific *Peromyscus* hybrids. *Nat. Genet.* **20**, 362–365. (doi:10.1038/3833)
103. Galbraith DA, Kocher SD, Glenn T, Albert I, Hunt GJ, Strassmann JE, Queller DC, Grozinger CM. 2016 Testing the kinship theory of intragenomic conflict in honey bees (*Apis mellifera*). *Proc. Natl Acad. Sci. USA* **113**, 1020–1025. (doi:10.1073/pnas.1516636113)
104. Kocher SD *et al.* 2015 A search for parent-of-origin effects on honey bee gene expression. *G3 Genes|Genomes|Genetics* **5**, 1657–1662. (doi:10.1534/g3.115.017814)
105. Smith NMA *et al.* 2020 Paternally-biased gene expression follows kin-selected predictions in female honey bee embryos. *Mol. Ecol.* **29**, 1523–1533. (doi:10.1111/mec.15419)
106. Crespi B, Nosil P. 2013 Conflictual speciation: species formation via genomic conflict. *Trends Ecol. Evol.* **28**, 48–57. (doi:10.1016/j.tree.2012.08.015)
107. Long TAF, Pischedda A, Stewart AD, Rice WR. 2009 A cost of sexual attractiveness to high-fitness females. *PLoS Biol.* **7**, e1000254. (doi:10.1371/journal.pbio.1000254)
108. Gavrilets S, Hayashi TI. 2005 Speciation and sexual conflict. *Evol. Ecol.* **19**, 167–198. (doi:10.1007/s10682-004-7916-4)
109. Hayashi TI, Vose M, Gavrilets S. 2007 Genetic differentiation by sexual conflict. *Evolution (NY)* **61**, 516–529. (doi:10.1111/j.1558-5646.2007.00059.x)
110. Hitchcock TJ, Gardner A. 2021 Sex-biased demography modulates male harm across the genome. Figshare.