

1 **Title:** Allometric growth in reef-building corals

2

3 **Authors:** Maria Dornelas¹, Joshua S. Madin², Andrew H. Baird³, and Sean R. Connolly^{3,4}

4 **Affiliations and emails:**

5 ¹Centre for Biological Diversity, Scottish Oceans Institute, University of St. Andrews, KY16

6 9TH, Scotland, UK, maadd@st-andrews.ac.uk

7 ²Department of Biological Sciences, Macquarie University, Sydney NSW 2109, Australia,

8 joshua.madin@mq.edu.au

9 ³ARC Centre of Excellence for Coral Reef Studies, James Cook University, Townsville QLD

10 4811, Australia, andrew.baird@jcu.edu.au

11 ⁴School of Marine and Tropical Biology, James Cook University, Townsville QLD 4811,

12 Australia, sean.connolly@jcu.edu.au

13

14

15

16 *The abstract is 141 words long, and in the main text is 3822. The manuscript cites 50*

17 *references, has 2 figures and 2 tables.*

18

19 Allometric growth in reef-building corals

20 **Abstract**

21 Predicting demographic rates is a critical part of forecasting the future of ecosystems under
22 global change. Here, we test if growth rates can be predicted from morphological traits for a
23 highly diverse group of colonial symbiotic organisms: scleractinian corals. We ask whether
24 growth is isometric or allometric among corals, and whether most variation in coral growth
25 rates occurs at the level of the species or morphological group. We estimate growth as change
26 in planar area for 11 species, across five morphological groups and over five years. We show
27 that coral growth rates are best predicted from colony size and morphology rather than
28 species. Coral size follows a power scaling law with a constant exponent of 0.91. Despite
29 being colonial organisms, corals have consistent allometric scaling in growth. This
30 consistency simplifies the task of projecting community responses to disturbance and climate
31 change.

32

33 **Key words:** allometry, scaling, traits, morphology, Scleractinia

34

35

36

37 **Introduction**

38 Predicting variation in life history is an essential element of ecology and evolution.
39 Combined demographic rates determine fitness, and therefore the effects of selection and
40 adaptation. Demographic rates are also critical to link population, community and ecosystem
41 ecology. Thus, developing robust predictors of demographic rates across entire assemblages
42 is a critical component of forecasting how global change, through its effects on species
43 diversity and composition, affects ecosystem function. For instance, how communities
44 recover from low densities following extreme disturbances, such as forest fires [1] or extreme
45 weather events [2], depends on differential growth rates among survivors and new recruits.
46 Here, we determine the best morphological predictors of growth rates of reef-building coral
47 colonies, the ecosystem engineers of one of the most threatened ecosystems on the planet [3].

48 Growth is a key demographic rate. From an organismal point of view, it measures
49 energetic and resource investment into somatic extension [4]. From a population perspective,
50 growth measures change in size, which in turn mediates changes in other demographic rates
51 (such as mortality and fecundity) [5]. From an ecosystem perspective, growth leads to
52 biomass production and creates habitat structure, and hence the contribution of the population
53 to multiple ecosystem functions [6]. Accurate predictions of growth rates are therefore useful
54 across ecological scales.

55 Most of what we know about growth comes from unitary organisms, which typically
56 have size dependent growth rates because of geometric and energetic constraints. In the most
57 extreme cases, somatic growth ceases once reproductive maturity is reached, a phenomenon
58 known as determinate growth [7]. In contrast, species with indeterminate growth partition
59 energy and resources between reproduction and growth throughout their life. There are
60 energetic reasons to expect growth to be size-dependent even for species with indeterminate

61 growth. For most organisms, metabolic rate scales with body mass [8]. Hence, the amount of
62 energy available for growth follows scaling laws, which have been hypothesized to arise out
63 of physical constraints associated with the fractal networks that distribute resources within
64 organisms [9]. Although the exact value of the scaling exponent, and its consistency across
65 organisms, has been hotly debated for nearly a century, it is always less than 1 (e.g. [10, 11])
66 indicating that growth slows down as organisms get bigger.

67 For colonial organisms, both the relationship between size and age [12], and the size
68 dependent allometric constraints that drive growth scaling laws, do not necessarily apply. In
69 fact, whether “body size” applies to the entire colony, or to the modules from which it is
70 comprised, depends on the physiological process and on the level of integration among
71 modules. For example, among social insects metabolic rate scales allometrically as a power
72 law of colony size, but isometrically among isolated worker groups that lack the social
73 integration of a colony [13]. Hence, allometric variation seems to be an indicator of
74 integration among modules in colonial organisms.

75 Among corals, growth is a function of the addition of modules to the colony and loss
76 of modules through partial mortality. Early work suggested rates of linear extension are
77 independent of colony size [14, 15]. For circular colonies, constant radial extension results in
78 a curvilinear relationship between sizes at different times, asymptotically approaching the
79 line of no-growth as the ratio of perimeter to area decreases with size. Moreover, there is also
80 evidence for complex allometries in energy acquisition, which affect energy available for
81 growth. On the one hand, resource acquisition is predicted to scale allometrically with body
82 size in corals as resource availability to interior modules is reduced because of active
83 depletion or passive deflection by modules on the exterior - similar to a self-shading effect
84 [16, 17]. On the other hand, process-based models of the effect of irradiance and gas
85 diffusion on photosynthesis and respiration predict that large colonies have a positive energy

86 balance over a broader range of flow and light conditions than small colonies [18, 19]. These
87 predictions are supported by positive correlations between predicted energy acquisition,
88 tissue biomass and reproductive output [18]. Clearly, more work is needed to test alternative
89 hypotheses regarding growth in reef corals. Here, we contrast support for the hypotheses that
90 scleractinian corals have isometric vs. allometric growth across a range of colony
91 morphologies.

92 Partial mortality has the potential to decouple colonies' metabolic capacity for growth
93 from their realized growth [12, 15]. In particular, the probability of injury increases in direct
94 proportion to colony size [12, 20], and hence, larger colonies exhibit lower rates of net colony
95 growth [14, 21]. Moreover, the high frequency and stochasticity of disturbances on coral
96 reefs might mean that temporal variation in demographic rates can overwhelm any other
97 signal [22]. Hence, we compare temporal variation in growth rates with inter species and inter
98 morphological group variation.

99 Trait-based ecology is a promising approach to understanding the drivers of variation
100 in demographic rates across ecological assemblages [23]. Rather than focusing on taxonomic
101 identity as a driver of variation, this approach aims to identify traits associated with variation
102 in response to environmental gradients, or in demographic rates. This approach is particularly
103 pertinent to the goal of estimating demographic rates in diverse assemblages [24]. Estimating
104 demographic rates individually for each species is logistically impossible for any but the
105 simplest assemblages, due to the number of parameters that require estimation, and the large
106 number of rare species [25], for which sufficient replication is difficult to obtain. Trait-based
107 demography is potentially an efficient solution to this problem, if easily measured traits can
108 be identified as proxies of demographic rates. However, the success of this approach depends
109 on the proportion of variation in demographic rates associated with traits vs. idiosyncratic
110 variation among and within species.

111 Trait variation across species is not independent, as it is the result of evolution.
112 Hence, some combinations of traits occur far more frequently than others, so that species (and
113 individuals) are typically organised in trait space into groups of similar traits and function
114 [26]. Assessing how much of the variation in demographic rates is explained by these trait-
115 functional groups is a parsimonious way of identifying whether predictive traits are likely to
116 be included in the group classification. It is also an efficient way to be able to scale from
117 ecological neighbourhoods to ecosystem functions, and predict how such functions respond
118 to global change [27]. This is an urgent task for coral reefs in particular, given the level of
119 threat to this ecosystem. Predicting growth rates across entire coral assemblages is critical to
120 be able to forecast recovery from disturbances.

121 Reef-building corals can be classified into morphological groups according to the
122 general shape of their colonies. Typical groups include massive, encrusting, foliaceous and
123 multiple types of branching forms that vary in the length, width and organization of branches.
124 These morphological groups are known to differ functionally in terms of contribution to reef
125 accretion [16] and habitat provision [28]. Hence, being able to predict production rates for
126 different morphological groups is critical to forecast changes in these different ecosystem
127 functions. The groups also differ in demographic rates, such as size specific mortality
128 schedules (e.g. [29]), and fecundity (e.g. [30]). Finally, colony morphologies differ markedly
129 in the degree of three-dimensionality, tissue penetration in skeleton, and surface to volume
130 ratios, all of which have been hypothesized to affect growth rates [16]. In accordance with a
131 trait-based approach, we hypothesize that most variation in demographic rates occurs
132 between morphological groups rather than between species of the same morphology.

133 Here, we ask how best to predict growth of reef corals. Specifically, we ask: 1) if
134 growth scales isometrically or allometrically with colony size; 2) whether scaling is similar
135 for all species, can be predicted from morphology or is species specific; and 3) how inter-

136 specific variation in growth rate compares to temporal intra-specific variation. Answers to
137 these questions are essential for predicting variation in growth rates across entire coral
138 assemblages, and hence for projecting recovery from disturbances, such as the 2016 mass-
139 bleaching event.

140

141 **Material and methods**

142 *Field data*

143 Coral growth can be quantified in different ways. We focus on quantifying change in
144 cover (or horizontal planar area), which is a metric of live coral used for example in global
145 biodiversity assessments [31]. Space and access to light are the main limiting resources for
146 which corals compete. Change in planar area is, thus, the most appropriate metric from a
147 coral demography perspective [32]. Three-dimensional surface area and volume are also
148 important metrics of size, as they correspond respectively to live tissue biomass and
149 deposited skeleton. These metrics are far more difficult to estimate *in situ*, especially non-
150 destructively. However, tight morphotype specific scaling relationships between these
151 metrics of size [33] suggest that planar area can serve as an effective proxy for three-
152 dimensional colony growth.

153 We estimated growth as change in planar area for 11 species of scleractinian corals,
154 across five morphological groups: tabular (*Acropora cytherea* and *A. hyacinthus*), arborescent
155 (*A. intermedia* and *A. robusta*), corymbose (*A. spathulata*, *A. millepora*, and *A. nasuta*),
156 digitate (*A. cf. digitifera* and *A. humilis*) and massive (*Goniastrea pectinata* and *G.*
157 *retiformis*). We tagged 30 colonies of each species with numbered cattle tags, or with metal
158 tags hammered into the substrate in the vicinity of the colonies. All colonies are distributed
159 along a 500 x 10 m area on the reef crest at Trimodal reef, between South and Palfrey islands
160 (14.699839°S, 145.448674°E), Lizard Island, Australia.

161 Each tagged colony was photographed from above with a two-dimensional scale plate
162 placed level with the surface of the colony. The angle of the camera was horizontal, and
163 distance from the colony was such that the entire colony was visible in the photograph.
164 Colonies were photographed yearly in November 2009, 2010, 2011, 2012 and 2013. The
165 images were corrected for barrel distortion, and the scale and outline of each colony were
166 digitised in ImageJ for estimation of planar area. Every year, dead or missing colonies were
167 replaced in order to maintain approximately 30 colonies per species. To minimize the effect
168 of observation error caused by slight differences in angle, distance from the camera or
169 positioning of the scale, 60% to 90% of the colonies were photographed twice to three times
170 independently every year, all photographs were digitized twice independently, and estimates
171 of area were averaged. In total we had 980 estimates of colony planar area, which are the
172 average of 2 to 9 replicate estimates for each of the 330 colonies (30 colonies x 11 species)
173 each year.

174 *Analysis*

175 We used ordinary least squares to fit linear models to estimates of planar area at one
176 point in time as a function of its planar area the year before, species, morphological group,
177 year and combinations of the above as outlined below. On a log-log plot of area at time t as a
178 function of area at time $t-1$, a line with slope of 1 and intercept of 0 corresponds to no change
179 in area. Colonies above this no growth line increase in size, and colonies below it decrease in
180 size between time points. A line of slope 1 and non-zero intercept indicates isometric growth:
181 colonies grow or shrink by a fixed proportion of their current colony size. A slope of more
182 than 1 indicates accelerating growth, and a slope of less than 1 decelerating growth. For lines
183 with slope less than 1 (and positive intercepts) once the line intercepts the unity line, colonies
184 start shrinking on average. Model selection was done using Akaike's Information Criterion

185 (AICc). Analyses were conducted using the “lm” and “lmer” [34] functions in the statistical
186 programming language R [35].

187 Our main goal was to find a model that best predicts change in planar area, and hence
188 we compare the fit of different combinations of predictor variables. Specifically, to test
189 whether growth rates change as a function of colony size, we fitted models with size the
190 previous year as a predictor variable, and as an offset. We also compared models with and
191 without morphological group or species to determine which level of differences best predicts
192 change in planar area. Because our sampling design included repeated measurements of the
193 same colonies, we also compared models with and without colony as a random effect
194 (Supplementary Material). We assessed the robustness of our results to functional group
195 classification (see Supplementary Material).

196 Change in planar area integrates colony growth with partial mortality. To attempt to
197 disentangle ability to grow from ability to withstand such conditions, we used quantile
198 models using the rq function in package quantreg [36]. Specifically, we hypothesize that the
199 95th quantile function reflects the limits of how much colonies can grow, and the 5th quantile
200 of the susceptibility to partial mortality. We fit quantile regressions through the median as
201 well, to check the robustness of inferences from the OLS regression to skew and
202 heterogeneity of variances in the residual variation.

203 Also, we considered temporal variation by comparing models that included year and
204 all possible interactions with morphological group and species. We asked how between group
205 variation in change in planar area compares to between year variation, and whether temporal
206 variation affects all groups similarly or differently.

207

208 **Results**

209 A model that includes size the previous year as a predictor of size in the current year
210 was always selected regardless of other variables included (Table 1). All models fitted, with
211 the exception of the quantile regression for the 0.05 quantile, had a positive slope of less than
212 one, and positive intercepts (Table 2). That is, irrespective of which model is fitted, growth as
213 a proportion of colony size decreases with increasing size, and at some point intercepts the
214 unity line so that, on average, colonies stop growing (Figure 1).

215 In a comparison among models where parameters vary as a function of species,
216 morphological group, year, and combinations of these predictor variables, the model with the
217 lowest AICc has a common slope but different intercepts for the interaction of each
218 morphological group and year (Table 1). This result is robust to using colony as a random
219 effect in the models (Supplementary Material).

220 A comparison of R^2 values between models with species or with morphological
221 groups as predictors shows that variation between species within morphological groups is
222 minimal (e.g. best OLS model with equivalent species level model has a 0.11% difference,
223 Table 3). The distribution of residuals for this model (Figure 1) indicate that negative
224 residuals are more spread than positive residuals, and further justify the quantile regression
225 analysis. The residuals also identify three potential outliers, hence all analyses were repeated
226 after removing these outliers with no effect on the results.

227 The ranking in growth rate for the different morphological groups (which is a function
228 of the intercept in these models, because the slope is constant) changed between quantiles
229 (Figure 2). For the 95th quantile, the ranking from lowest to highest is massive, digitate,
230 corymbose, arborescent, encrusting arborescent, and tabular. For the median quantile,
231 representing a trade-off between positive and negative change in size, the ranking becomes
232 corymbose, encrusting arborescent, arborescent, massive, digitate and tabular. For the 5th

233 quantile, the ranking is encrusting arborescent, corymbose, tabular, arborescent, digitate and
234 massive. Parameter estimates for these three models are presented in Table 2.

235 There are also clear differences in inter-annual variability among morphological
236 groups. All groups, with the exception of massive colonies, had a decrease in both the ability
237 to grow and median growth within the study period (Table 2). Massive colonies had very low
238 inter-annual variability in both the OLS and the three quantile regression models (Figures 1
239 and 2). All groups, with the exception of encrusting arborescents, also had increasing
240 reductions in size over the study period. Encrusting arborescents decreased their losses, but
241 were also the group with the highest losses overall (Table 2, Figure 2).

242

243 **Discussion**

244 Coral growth rates scale with size according to a power law with constant slope across
245 all species. This result is clearly indicative of allometric growth among corals: if growth were
246 isometric, size in successive years would be proportional to size in previous years, yielding a
247 slope of one in log-log space. Allometric growth is consistent with the hypothesis of colony-
248 level integration among modules. Integration can take the form of energy or nutrient sharing
249 among modules, and/or it can be a consequence of geometric constraints associated with
250 colony morphology. The identical estimated slope across morphological groups implies that
251 constraints placed by size on growth are highly consistent across these morphological groups
252 in reef-building corals. For all species in our study, colonies increase size up to a point when,
253 on average, they stop growing.

254 Colonial organisms can theoretically grow indefinitely by adding modules to their
255 colony, but in practice there are geometric and structural constraints to how modules are
256 added. Proportional growth rates are expected to decrease with size because of geometric
257 constraints in how corals add cover [32, 37]. Corals increase in planar area by adding area at

258 the periphery of the colony. Because the ratio of perimeter to area decreases with size, unless
259 radial increments increase with size, proportional growth rate necessarily decreases with size.
260 Perimeter increases with area according to functions that depend on the shape of the colony,
261 but that, typically, asymptotically approach the line of zero growth. Constant radial increment
262 thus corresponds to allometric growth.

263 Net growth is a function not only of the ability to grow, but also of partial mortality.
264 This trade-off is reflected in the changed rankings observed in the median quantile regression,
265 where most morphological groups have similar growth rates, and the lowest quantile
266 regression, where the rankings are largely reversed. While exponent estimates for maximum,
267 median and mean growth are all indicative of allometric scaling, the exponent estimate for
268 the approximate lower bound of net (negative) growth (5th percentile) is not significantly
269 different from one. This indicates that partial mortality is a fixed fraction of colony area,
270 independent of colony size, a finding that is consistent with previous reports that the
271 probability of injury increases proportionately with colony size [12, 20]. Because growth
272 decreases with colony size, and partial mortality is isometric, it follows that, at a threshold
273 colony size, partial mortality will cancel growth, and colonies above this threshold will tend
274 to shrink, on average.

275 In addition to the geometric constraints, there are also energetic colony level
276 constraints to growth in corals. Energy acquisition is predicted to scale allometrically in
277 corals [17]. Additionally, larger colonies may invest a disproportionate portion of available
278 energy to maintenance or reproduction, reducing energy available for growth [38-41].
279 Similarly to unitary organisms, both geometric and energetic constraints are likely involved
280 in determining growth rates in colonial organisms, and are indicative of integration among
281 the modules of the colony. Specifically, the geometric constraints we discuss arise out of
282 partitioning of tasks among modules, a key characteristic of colony integration, with growing

283 modules situated at the periphery of the colony. Colony level energetic constraints are
284 indicative of energy partitioning among modules.

285 We found that variation in size dependent growth is best predicted by morphological
286 group rather than species. This is consistent with previous studies showing that colony
287 morphology is associated with coral adaptive strategies [42]. In addition, there is an extensive
288 literature reporting marked differences in life history rates among different morphological
289 groups of coral reef invertebrates [43-46]. For example, [37] report a 20-fold difference in
290 growth rates between branching and massive corals, [47] report a 30-fold difference in
291 skeletal extension among branching vs. encrusting colonies. Nonetheless, at some scale
292 phylogeny is likely to be important. For example, family in addition to morphology was an
293 important predictor of growth rates across the order Scleractinia [24]. Future studies
294 increasing the taxonomic breadth of growth estimates will be critical to further test the
295 generality of our common scaling exponent.

296 Finally, it is interesting to note that inter-annual variation was generally greater than
297 inter-group variation, but that the degree of these discrepancies differed among the groups.
298 Specifically, robust slow growing morphologies such as massive and digitate had the least
299 temporal variation, whereas the more fragile fast growing morphologies such as tables and
300 arborescent had the highest (Figure 1 and 2). The shifts in rankings between good and bad
301 years for fast and slow growers highlight the long recognised need for long-term studies for
302 robust estimates of coral growth rates. In combination, the shifts in rankings for different
303 quantiles of growth and the patterns of temporal variation support the existence of a trade-off
304 in robustness and ability to grow in corals. The morphological groups most likely to be able
305 to withstand disturbances are also the slowest to grow. If disturbances become too frequent or
306 too extreme in magnitude the ability of reefs to recover will be compromised.

307 Tight scaling relationships between 2D and 3D metrics of size [33] suggest it is
308 possible to convert growth across different metrics. Thus our estimates of growth rates,
309 coupled with morphotype specific scaling relationships between volume, planar, and surface
310 area [33], allow estimating not only how coral cover changes, but also reef production. Both
311 mortality [45] and fecundity [30] are best modelled at morphological group level. Therefore,
312 combining geometric conversions with assemblage size structure information will allow for a
313 range of estimates about reef-scale functional processes, such as reef production (growth),
314 erosion (mortality) and the reproductive output (fecundity) for metapopulation models.

315 The importance of colony size as a predictor of growth has implications for
316 monitoring reef coral communities and predicting their responses to global change.
317 Modelling of demographic change in coral populations and communities should incorporate
318 allometric scaling of size. Current results suggest a constant scaling exponent of 0.91, but this
319 value and its generality may be revised as more data accumulates. Both mortality [45] and
320 fecundity [30] are also size dependent, which is consistent with the classical approach of size
321 dependent demography for corals [48]. An important implication of this result is that constant
322 growth underestimates rapid growth of small colonies, typical of early successional stages
323 following a disturbance, and overestimates growth of large colonies. As global change
324 increases the frequency and intensity of episodic disturbances size structures will shift
325 downwards [49], and this will alter how fast remaining individuals are able to re-grow.

326 The most commonly reported metric of reef health is coral cover, but in isolation it
327 does not allow forecasting change in ecosystem function or recovery. Size distributions and
328 morphological group abundances emerge as two essential variables that are easy to quantify,
329 and should be prioritised in coral reef monitoring. In combination these variables also allow
330 predicting habitat function changes in the reef, as habitat complexity is determined by which
331 coral morphotypes are present, and fish communities reflect habitat complexity. Projections

332 of how coral cover will respond to environmental change often use coral cover as a state
333 variable, and assume that growth is a fixed proportion of existing cover (i.e. that it is
334 isometric) [50, 51]. The allometry of growth that we documented here means that shifts in
335 size structure will lead to systematic changes in the proportional growth in cover of
336 population, which such models will not capture. Predictions about community shifts under
337 global change will need to consider differences in susceptibility, recovery potential, and their
338 interaction.

339

340

341 **Data accessibility:** Data used in this manuscript is included as supplementary material.

342

343 **Competing interests:** We have no competing interests.

344

345 **Author Contributions:** All authors designed the study and collected the data. MD analysed
346 the data and wrote the first draft. All authors contributed substantially to revisions.

347

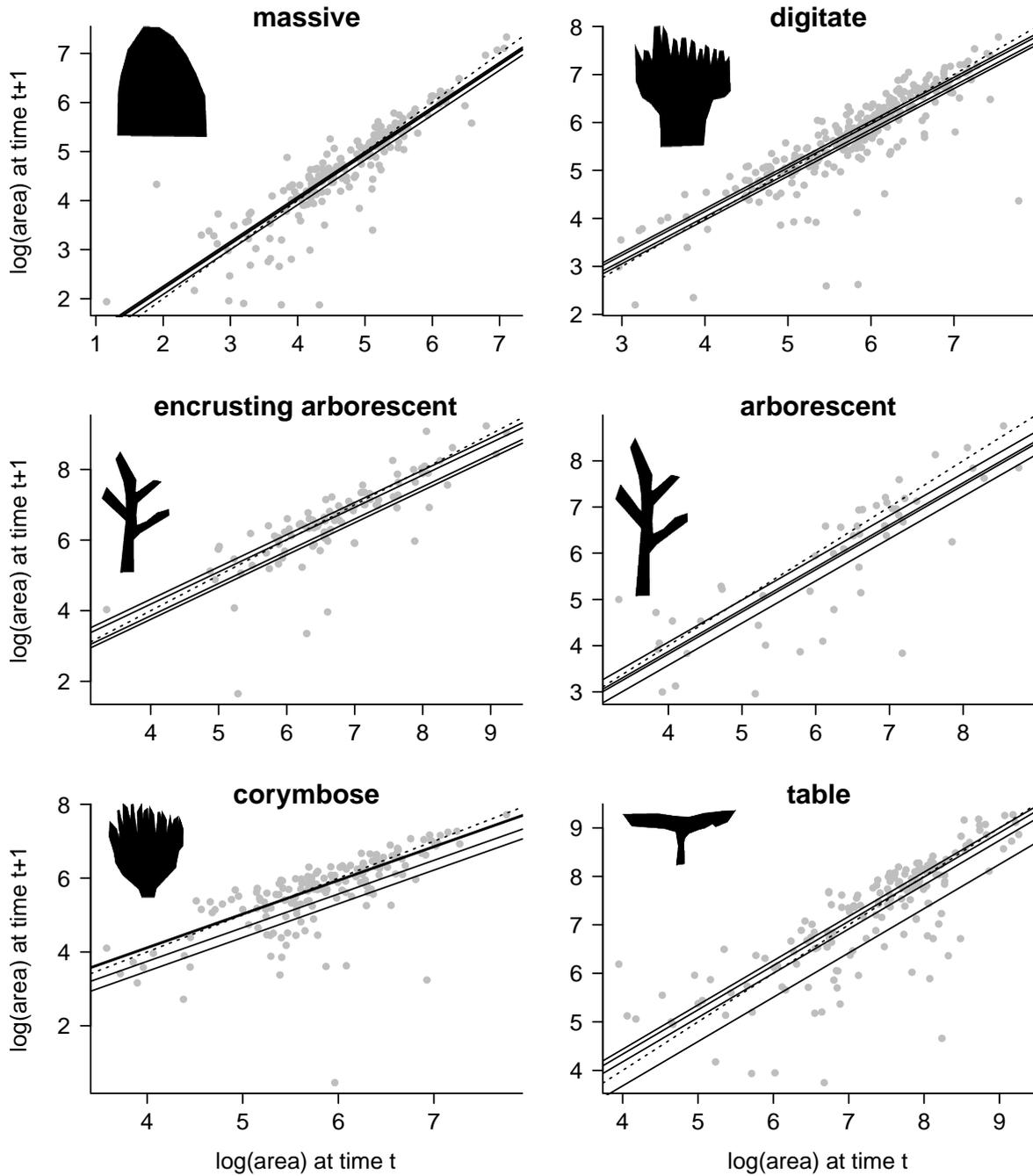
348 **Acknowledgments:** We are grateful to Mizue Hissano, Loic Thibault, Maddeline Emms,
349 Miguel Barbosa, Erin Graham, Dominique McGowan, Viv Cumbo, Emily Howels, Shane
350 Blowes, Marcela Diaz, Mariana Alvarez Noriega for help collecting the data.

351

352 **Funding:** We thank the ARC Centre of Excellence for Coral Reef Studies and the Australian
353 Research Council for fellowship and research support. We are also grateful to the Scottish
354 Funding Council (MASTS, grant reference HR09011) and to the ERC project bioTIME.

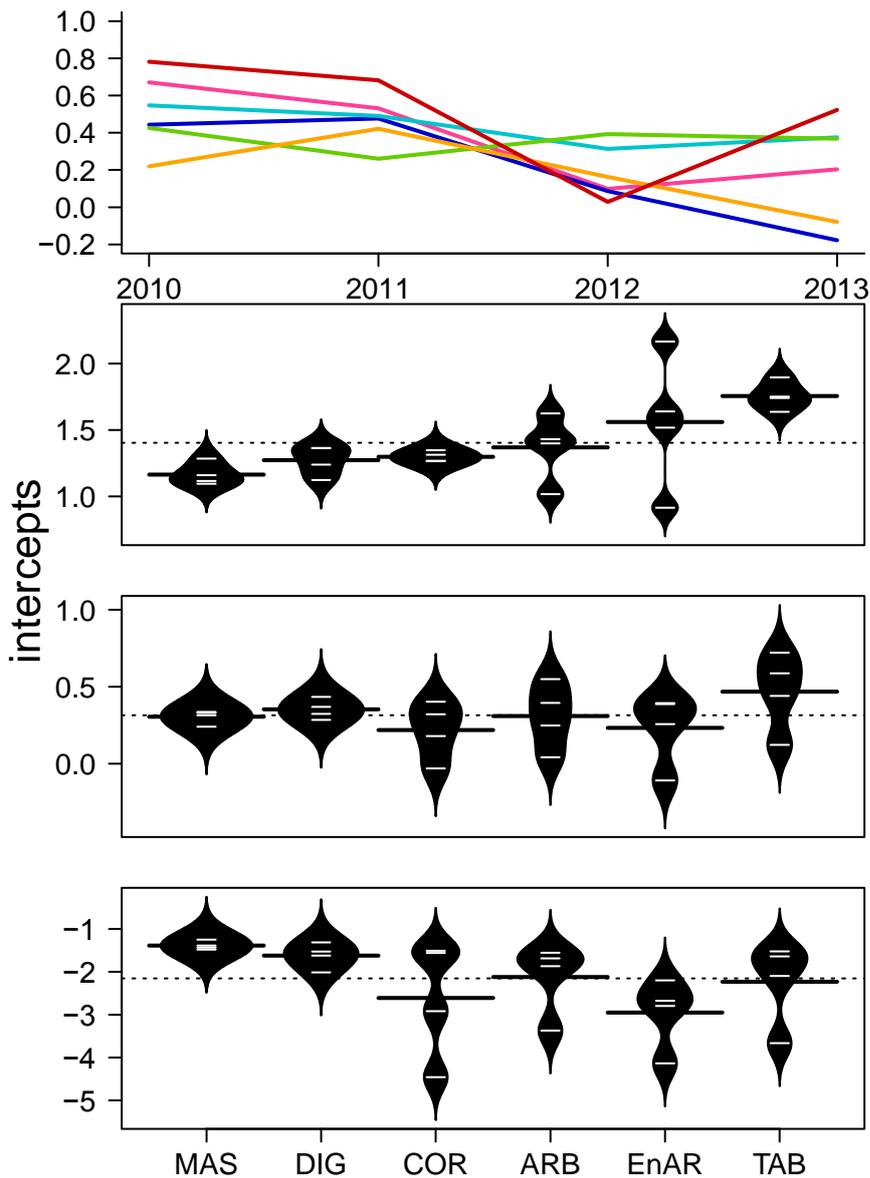
355

Figure captions



358
359
360
361
362
363

Figure 1 – Size at time t+1 as a function of size at t for each morphological group as per panel label. Circles represent data, full lines the best fitted model as per table 3 (one line for each year) and dotted line the no-growth line (when size is unchanged between one year and the next).



364
 365 **Figure 2 – Comparison of temporal variation in mean, maximum, median and**
 366 **minimum growth. Top:** OLS estimates of intercepts for best model as per Table 3, for each
 367 year and each morphological group. The red line represents tabular colonies, orange
 368 arborescent, green massive, turquoise digitate, blue corymbose and pink encrusting
 369 arborescent). Intercepts across years for the 95th (**second from top**), 50th (**third from top**),
 370 and 5th (**bottom**) quantile regressions of size at time t as a function of size at time t-1 and
 371 morphological group (as per best model in table 3). Estimates are plotted as white lines in the
 372 centre of each black polygon, which represents a normal density curve around the estimates
 373 and is intercepted by a black line representing the mean for each morphological group. The
 374 dotted line represents the mean across all morphological groups.
 375

376
377

Table 1 – Model selection for the effect of time, species and morphological group

model	df	AICc	adjusted R ²
$\log(\text{area}_{t+1}) \sim \text{offset}(\log(\text{area}_t))$	2	1971.377	
$\log(\text{area}_{t+1}) \sim \text{offset}(\log(\text{area}_t)) + \text{group}$	7	1964.730	0.7985
$\log(\text{area}_{t+1}) \sim \text{offset}(\log(\text{area}_t)) + \text{year}$	5	1950.108	0.8081
$\log(\text{area}_{t+1}) \sim \text{offset}(\log(\text{area}_t)) + \text{species}$	12	1971.011	0.7976
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t)$	3	1955.052	0.7807
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) + \text{group}$	8	1946.720	0.7837
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) + \log(\text{area}_t):\text{group}$	8	1947.799	0.7833
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) * \text{group}$	13	1955.384	0.7833
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) + \text{species}$	13	1952.889	0.7835
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) + \log(\text{area}_t):\text{species}$	13	1953.215	0.7833
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t)*\text{species}$	23	1957.555	0.7846
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) + \text{year}$	6	1927.974	0.7895
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) + \log(\text{area}_t):\text{year}$	6	1919.232	0.7911
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t)*\text{year}$	9	1914.654	0.7915
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) + \text{year} + \text{group}$	11	1917.721	0.7927
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) + \text{year} * \text{group}$	26	1908.040	0.7986
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) * \text{group} * \text{year}$	49	1910.305	0.7921
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) + \text{year} + \text{species}$	16	1923.726	0.7926
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) + \text{year} * \text{species}$	46	1924.281	0.7997
$\log(\text{area}_{t+1}) \sim \log(\text{area}_t) * \text{species} * \text{year}$	89	1929.261	0.8074

378
379

380
381
382
383
384
385

Table 2- Parameter estimates (mean intercepts across years) for the OLS and quantile regression models 95th, 50th and 5th quantile regressions of planar area at time t as a function of planar area at time t-1 and morphological group.

	OLS	95th	50th	5th
α	0.913	0.851	0.944	1.111
$\beta_{\text{arborescent}}$	0.375	1.368	0.308	-2.121
$\beta_{\text{encrustingarborescent}}$	0.181	1.559	0.232	-2.952
$\beta_{\text{corymbose}}$	0.207	1.298	0.218	-2.611
β_{digitate}	0.431	1.272	0.352	-1.619
β_{massive}	0.361	1.163	0.305	-1.388
β_{tabular}	0.503	1.756	0.467	-2.231

386
387
388

389 **References**

390
391

- 392 [1] He, H.S. & Mladenoff, D.J. 1999 Spatially explicit and stochastic simulation of forest -
393 landscape fire disturbance and succession. *Ecology* **80**, 81-99.
- 394 [2] Jentsch, A. & Beierkuhnlein, C. 2008 Research frontiers in climate change: effects of
395 extreme meteorological events on ecosystems. *Comptes Rendus Geoscience* **340**, 621-628.
- 396 [3] Hughes, T.P., Baird, A.H., Bellwood, D.R., Card, M., Connolly, S.R., Folke, C.,
397 Grosberg, R., Hoegh-Guldberg, O., Jackson, J.B.C., Kleypas, J., et al. 2003 Climate change,
398 human impacts, and the resilience of coral reefs. *Science* **301**, 929-933.
- 399 [4] Kooijman, S.A.L.M. 2010 *Dynamic energy budget theory for metabolic organisation*,
400 Cambridge university press.
- 401 [5] Caswell, H. 2001 *Matrix population models*, Wiley Online Library.
- 402 [6] Vile, D., Shipley, B. & Garnier, E. 2006 Ecosystem productivity can be predicted from
403 potential relative growth rate and species abundance. *Ecology letters* **9**, 1061-1067.
- 404 [7] Yeager, A. 1927 Determinate growth in the tomato. *Journal of Heredity* **18**, 263-265.
- 405 [8] Kleiber, M. 1932 Body size and metabolism. *Hilgardia* **6**, 315-332.
- 406 [9] West, G.B., Brown, J.H. & Enquist, B.J. 2001 A general model for ontogenetic growth.
407 *Nature* **413**, 628-631.
- 408 [10] Damuth, J. 2001 Scaling of growth: plants and animals are not so different. *Proceedings*
409 *of the National Academy of Sciences* **98**, 2113-2114.
- 410 [11] Glazier, D.S. 2005 Beyond the '3/4 - power law' : variation in the intra - and
411 interspecific scaling of metabolic rate in animals. *Biological Reviews* **80**, 611-662.
- 412 [12] Hughes, T.P. & Jackson, J.B.C. 1980 Do corals lie about their age? Some demographic
413 consequences of partial mortality, fission and fusion. *Science* **209**, 713-715.
- 414 [13] Waters, J.S., Holbrook, C.T., Fewell, J.H. & Harrison, J.F. 2010 Allometric scaling of
415 metabolism, growth, and activity in whole colonies of the seed - harvester ant
416 *Pogonomyrmex californicus*. *The American Naturalist* **176**, 501-510.
- 417 [14] Connell, J.H. 1973 Population ecology of reef-building corals. In *Biology and Geology*
418 *of coral reefs* (eds. G.P. Jones & Endean), pp. 205-245.
- 419 [15] Hughes, T.P. & Jackson, J.B.C. 1985 Population Dynamics and Life Histories of
420 Foliose Corals. *Ecological Monographs* **55**, 142-166. (doi:10.2307/1942555).
- 421 [16] Buddemeier, R.W. & Kinzie, R.A. 1976 Coral Growth. *Oceanography and Marine*
422 *Biology: An Annual Review* **14**, 183-225.
- 423 [17] Kim, K. & Lasker, H. 1998 Allometry of resource capture in colonial cnidarians and
424 constraints on modular growth. *Functional Ecology* **12**, 646-654.
- 425 [18] Hoogenboom, M.O. & Connolly, S.R. 2009 Defining fundamental niche dimensions of
426 corals: synergistic effects of colony size, light, and flow. *Ecology* **90**, 767-780.
- 427 [19] Jokiel, P.L. & Morrissey, J.I. 1986 Influence of size on primary production in the reef
428 coral *Pocillopora damicornis* and the macroalga *Acanthophora spicifera*. *Marine Biology* **91**,
429 15-26.
- 430 [20] Jackson, J.B.C. 1979 Morphological strategies of sessile animals. In *Biology and*
431 *Systematics of colonial organisms* (eds. G. Larwood & B.R. Rosen), pp. 499-555. London,
432 Academic Press.
- 433 [21] Hughes, T.P. 1987 Skeletal density and growth form of corals. *Marine Ecology Progress*
434 *Series* **35**, 259-266.
- 435 [22] Hughes, T.P. & Connell, J.H. 1999 Multiple stressors on coral reefs: A long-term
436 perspective. *Limnology & Oceanography* **44**, 932-940.
- 437 [23] McGill, B.J., Enquist, B.J., Weiher, E. & Westoby, M. 2006 Rebuilding community
438 ecology from functional traits. *Trends In Ecology & Evolution* **21**, 178-185.

439 [24] Madin, J.S., Hoogenboom, M.O., Connolly, S.R., Darling, E.S., Falster, D.S., Huang, D.,
440 Keith, S.A., Mizerek, T., Pandolfi, J.M. & Putnam, H.M. 2016 A trait-based approach to
441 advance coral reef science. *Trends in ecology & evolution*.
442 [25] McGill, B.J., Etienne, R.S., Gray, J.S., Alonso, D., Anderson, M.J., Benecha, H.K.,
443 Dornelas, M., Enquist, B.J., Green, J.L. & He, F. 2007 Species abundance distributions:
444 moving beyond single prediction theories to integration within an ecological framework.
445 *Ecology letters* **10**, 995-1015.
446 [26] Raunkiaer, C. 1934 The life forms of plants and statistical plant geography; being the
447 collected papers of C. Raunkiaer. *The life forms of plants and statistical plant geography;*
448 *being the collected papers of C. Raunkiaer*.
449 [27] Díaz, S. & Cabido, M. 1997 Plant functional types and ecosystem function in relation to
450 global change. *Journal of vegetation science* **8**, 463-474.
451 [28] Graham, N.A.J., Wilson, S.K., Jennings, S., Polunin, N.V.C. & Bijoux, J.P. 2006
452 Dynamic fragility of oceanic coral reef ecosystems. *Proceedings of the National Academy of*
453 *Science* **103**, 8425-8429.
454 [29] Madin, J.S., Baird, A.H., Dornelas, M. & Connolly, S.R. 2014 Mechanical vulnerability
455 explains size - dependent mortality of reef corals. *Ecology letters* **17**, 1008-1015.
456 [30] Álvarez - Noriega, M., Baird, A.H., Dornelas, M., Madin, J.S., Cumbo, V.R. &
457 Connolly, S.R. 2016 Fecundity and the demographic strategies of coral morphologies.
458 *Ecology*.
459 [31] Butchart, S.H.M., Walpole, M., Collen, B., van Strien, A.J., Scharlemann, J.W.,
460 Almond, R.E.A., Baillie, J.E.M., Bomhard, B., Brown, C., Bruno, J., et al. 2010 Global
461 Biodiversity: Indicators of Recent Declines. *Science*, 1164-1168.
462 [32] Pratchett, M.S., Anderson, K.D., Hoogenboom, M.O., Widman, E., Baird, A.H.,
463 Pandolfi, J.M., Edmunds, P.J. & Lough, J.M. 2015 Spatial, temporal and taxonomic variation
464 in coral growth - implications for the structure and function of coral reef ecosystems.
465 *Oceanography and Marine Biology: an Annual Review* **53**, 215-295.
466 [33] House, J.E., Bidaut, L.M., Christie, A.P., Pizarro, O. & Dornelas, M. 2016 Moving to
467 3D: relationships between coral planar area, surface area and volume. *PeerJ Preprints* **4**,
468 e2443v2441.
469 [34] Bates, D., Maechler, M., Bolker, B. & Walker, S. 2014 lme4: Linear mixed-effects
470 models using Eigen and S4. *ArXiv e-print*. (doi:URL: <http://arxiv.org/abs/1406.5823>).
471 [35] Team, R.D.C. 2013 R: A language and environment for statistical computing. (Vienna,
472 Austria, R Foundation for Statistical Computing.
473 [36] Koenker, R. 2015 quantreg: Quantile Regression. R package version 5.11.
474 <http://CRAN.R-project.org/package=quantreg>.
475 [37] Gladfelter, E.H., Monahan, R., K. & Gladfelter, W.B. 1978 Growth rates of five reef
476 building corals in the northeastern Caribbean. *Bulletin of Marine Science* **28**, 728-734.
477 [38] Anthony, K.R.N., Connolly, S.R. & Willis, B.L. 2002 Comparative analysis of energy
478 allocation to tissue and skeletal growth in corals. *Limnology and Oceanography* **47**, 1417-
479 1429.
480 [39] Barnes, D. & Lough, J. 1993 On the nature and causes of density banding in massive
481 coral skeletons. *Journal of Experimental Marine Biology and Ecology* **167**, 91-108.
482 [40] Rinkevich, B. 1996 Do reproduction and regeneration in damaged corals compete for
483 energy allocation? *Marine Ecology Progress Series* **143**, 297-302.
484 [41] Rodrigues, L.J. & Grottoli, A.G. 2007 Energy reserves and metabolism as indicators of
485 coral recovery from bleaching. *Limnology and Oceanography* **52**, 1874-1882.
486 [42] Darling, E.S., Alvarez-Filip, L., Oliver, T.A., McClanahan, T.R. & Côté, I.M. 2012
487 Evaluating life-history strategies of reef corals from species traits. *Ecology Letters*. (doi:DOI:
488 10.1111/j.1461-0248.2012.01861.x).

- 489 [43] Jackson, J.B.C. & Hughes, T.P. 1985 Adaptive strategies of coral-reef invertebrates:
490 coral reef environments that are regularly disturbed by storms and by predation often favor
491 the very organisms most susceptible to damage by these processes. *American Scientist* **73**,
492 265-274.
- 493 [44] Hughes, T.P. 1987 Herbivory on coral reefs: community structure following mass
494 mortalities of sea urchins. *Journal of Experimental Marine Biology and Ecology* **113**, 39-59.
- 495 [45] Madin, J.S., Baird, A.H., Dornelas, M. & Connolly, S.R. 2014 Mechanical vulnerability
496 explains size-dependent mortality of reef corals. *Ecology Letters*, n/a-n/a.
497 (doi:10.1111/ele.12306).
- 498 [46] Marshall, P.A. & Baird, A.H. 2000 Bleaching of corals on the Great Barrier Reef:
499 differential susceptibilities among taxa. *Coral Reefs* **19**, 155-163.
- 500 [47] Morgan, K. & Kench, P. 2012 Skeletal extension and calcification of reef-building
501 corals in the central Indian Ocean. *Marine environmental research* **81**, 78-82.
- 502 [48] Hughes, T.P. 1984 Population dynamics based on individual size rather than age: a
503 general model with a reef coral example. *The American Naturalist* **123**, 778-795.
- 504 [49] Hughes, T.P. 2000 Recruitment failure, life histories, and long term decline of Caribbean
505 corals. *Ecology* **81**, 2250-.
- 506 [50] Mumby, P.J., Hastings, A. & Edwards, H.J. 2007 Thresholds and the resilience of
507 Caribbean coral reefs. *Nature* **450**, 98-101.
- 508 [51] Weijerman, M., Fulton, E.A., Kaplan, I.C., Gorton, R., Leemans, R., Mooij, W.M. &
509 Brainard, R.E. 2015 An integrated coral reef ecosystem model to support resource
510 management under a changing climate. *PloS one* **10**, e0144165.
- 511
512
513