

1 **Impact of snare injuries on parasite prevalence in wild**
2 **chimpanzees (*Pan troglodytes*)**

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15

16 **Abstract**

17 Many primate populations are severely threatened by human activity. Illegal hunting with
18 snares frequently causes fatal injuries and permanent mutilations in wild primates. Traumatic
19 injuries and stressful experiences can reduce the efficacy of the immune system to fight
20 parasitic infections. Snare-related changes in primate behaviour may also influence the
21 probability of exposure to parasites. We hypothesized that primates with permanent snare-
22 related injuries would have higher prevalence of intestinal parasites than control individuals.
23 We tested the relationship between snare injuries and the prevalence of intestinal parasites in
24 wild chimpanzees (*Pan troglodytes*) of Budongo forest, Uganda. We collected 487 faecal
25 samples from known individuals (70 control and 20 snare-injured chimpanzees) and used
26 flotation and sedimentation to isolate helminth eggs and an immune-chromatographic assay to
27 identify protozoan cysts. We found that the prevalence of Strongylida nematodes was
28 significantly higher in snare-injured chimpanzees than control individuals. In contrast, we
29 found no association between snare injuries and three other parasite taxa: *Ascaris*, cestode and
30 *Cryptosporidium parvum*. Our study suggests that snare-injured primates may have higher
31 exposure and/or be more susceptible to developing infections with helminth parasites than
32 control individuals. Future studies should investigate whether snare injuries influence parasite
33 prevalence in other species of wild primates.

34

35 **Keywords**

36 Conservation, helminths, intestinal parasites, *Pan troglodytes*, protozoa, snare injury

37 **Introduction**

38 Illegal snare setting is common in many parts of subtropical Africa, especially in areas where the
39 local population relies on wild game for their livelihood (National Environment Management
40 Authority, 2010; Tumusiime et al., 2010). In tropical forests in West and Central Africa, more
41 than 70 mammal species are hunted (Fa et al., 2005). Poachers usually target animal species
42 indiscriminately, and terrestrial primates are frequent victims, often with fatal consequences. The
43 use of snares is the preferred technique for many rural population because it is easy to set them in
44 a short period of time (Tumusiime et al., 2010).

45

46 When caught in a snare, animals usually react by trying to pull the snare from its substrate, which
47 causes the wire to tighten and to cut deeper into the flesh (Noss, 1998). Although larger primates
48 can separate the snare from the substrate, the wire often remains tightly wrapped around the
49 affected body part, causing obstruction in blood circulation, infections, necrosis, loss of function
50 and mutilation. Although not directly targeted, chimpanzees (*Pan troglodytes*) are sometimes
51 accidentally caught in snares, which can cause severe and permanent mutilations (Waller, 1995).
52 These injuries are a substantial problem in many chimpanzee communities. For example, in
53 Budongo Forest, Uganda, an estimated third of the entire chimpanzee population suffers from
54 some form of snare injury (Reynolds, 2005).

55

56 Snare injuries can affect chimpanzee behaviour in various ways. One study found that severely
57 disabled females spent more time in small parties than control females, probably to avoid food
58 competition and because they have more difficulty to follow larger travelling parties (Hermans,
59 2011). Another study found that female chimpanzees with snare injuries carried their offspring
60 less often than uninjured females, especially once the infants got older and heavier (Munn,
61 2006). In the same study, the severity of a snare injury was also negatively correlated with the

62 daily travel distance, suggesting that disabled mothers may be disadvantaged in their foraging
63 capacities. In addition, snare-injured mothers spent more time in trees than uninjured mothers,
64 suggesting that snare injuries influence climbing abilities. Snare injuries may also reduce the
65 ability of male chimpanzees to rise in social rank, which will negatively affect their mating
66 opportunities and their ability to compete for food (Smith, 1995). Finally, chimpanzees rely
67 heavily on their hands when processing food (Byrne and Stokes, 2002), suggesting that hand
68 injuries will reduce their ability to access food and require them to use suboptimal techniques,
69 such as using their mouth or feet to manipulate food items.

70

71 Despite the obvious physical harm of a snare injury, the long-term fitness consequences for
72 injured chimpanzees remain poorly understood. In Budongo Forest (Uganda), there have been
73 two reports of death due to snares between 1990 and 2005, although the true mortality rate is
74 estimated to be much higher, around 2-3 fatalities per year (Reynolds, 2005). Snare injuries do
75 not seem to have an obvious negative impact on the physical condition of chimpanzees that
76 survive (Smith, 1995). However, one adult male of the Sonso community in Budongo Forest
77 suffered from almost total paralysis of both hands due to snare injuries and developed chronic
78 skin problems, presumably due to a lack of auto-grooming. (Hobaiter and Byrne, 2010).

79

80 Chimpanzees harbour numerous intestinal parasites that can affect their immune system (Abee et
81 al., 2012). Chimpanzees can acquire these parasites when they come into contact with
82 contaminated faecal matter. There is some circumstantial evidence to suggest that animals with
83 snare injuries might have higher exposure rates to intestinal parasites than healthy animals. For
84 example, snare-injured chimpanzees reuse old nests more often than control individuals
85 (Plumptre and Reynolds, 1997) and are therefore more likely to encounter parasite-
86 contaminated faecal matter. The ability of chimpanzees to fight intestinal parasites also depends

87 on the quality of their immune system (Coe, 2012). The efficacy of the immune system, in turn,
88 depends on nutrition, stress, and other factors. Snare-injured individuals may have lower quality
89 nutrition and higher stress levels, which will reduce their immune-competence to fight parasitic
90 infections.

91

92 We aimed to compare the prevalence of intestinal parasites between snare-injured and control
93 chimpanzees in two communities of Budongo Forest, Uganda. We predicted that increased
94 exposure and/or reduced immune-competence would result in chimpanzees with snare injuries
95 having higher prevalence of intestinal parasites than control chimpanzees.

96

97 **Materials and methods**

98 **Study area and subjects**

99 We conducted this study at the Budongo Conservation Field Station (BCFS), a Ugandan
100 NGO, in the Budongo Forest Reserve (latitude 1°37'N–2° 03'N, longitude 31°22'E–
101 31°46'E), Masindi District, North Western Uganda (Mwavu and Witkowski, 2008). Budongo
102 Forest is the largest natural forest in Uganda with an area of about 825 km² (Reynolds, 2005)
103 and a mean altitude of 1,100 m. This semi-deciduous tropical rainforest experiences a mean
104 annual rainfall of 1,600 mm. Peak precipitation occurs between March and May and between
105 September and November with a dry season from December to February. Temperatures vary
106 between 19°C and 32°C (Reynolds, 2005).

107

108 We obtained faecal samples from two chimpanzee communities, the Waibira group (partially
109 habituated; observations since 2011) and the Sonso group (fully habituated; observations
110 since 1990) over a period of four years. We collected samples from the Sonso community
111 between 2011 and 2014 and from the Waibira community from 2013 to 2014. We classified

112 the chimpanzees that had a morphological change due to a snare injury as “snare-injured”.
113 These morphological changes included clenched toes, hooked hands, missing hands, or
114 missing legs. We classified the chimpanzees that were never injured by a snare or that had
115 suffered a snare injury earlier in life, which did not result in a morphological disability as
116 “controls”. For both communities, we collected faecal samples opportunistically from known
117 individual chimpanzees using non-invasive sampling methods. Some chimpanzees were not
118 well habituated (particularly those in the Waibira community) so it was difficult to collect
119 many samples from these individuals. For the Sonso community, we were unable to obtain
120 samples when chimpanzees foraged on crops in human settlements adjacent to the forest
121 reserve due to the ethical guidelines of BCFS. As a result, we did not obtain a balanced
122 dataset for the different focal animals. To obtain an estimate of population-wide patterns, we
123 included faecal samples from all identified individuals, even if sample sizes were not
124 balanced. We placed all faecal samples in sterile containers and labelled them with the
125 individual’s name, sex, age, and date of defecation.

126

127 **Laboratory analysis**

128 We analysed all faecal samples at the BCFS field laboratory for a range of intestinal parasites
129 that are known to infect wild chimpanzees in the area (Barrows, 1996; Mugisha, 2004;
130 Zommers, 2010). We identified the following parasites: intestinal helminth worms, such as
131 Strongylid nematodes (Taylor et al., 2007), *Ascaris* nematodes, *Trichuris* nematodes and
132 cestodes. We lumped all cestodes, although this class includes many genera. We also
133 identified intestinal protozoan parasites, such as *Cryptosporidium parvum*, *Giardia lamblia*
134 and *Entamoeba histolytica*.

135

136 We used the RIDA®QUICK Cryptosporidium/ Giardia/ Entamoeba Combi Test (R-biopharm
137 AG, Germany), which is an antigen kit designed to detect protozoan parasites
138 (*Cryptosporidium parvum*, *Giardia lamblia*, *Entamoeba histolytica*). For each faecal sample,
139 we transferred ~ 50 mg of material into a tube without formalin and mixed with extraction
140 buffer, before inserting the test strip into the resulting solution. The analysis is based on an
141 immune-chromatographic assay, with parasite-specific antibodies on the strip indicating
142 parasite presence by specific colours. For all other parasites, we transferred samples into tubes
143 with 10% buffered formalin for later analysis. We used two techniques to isolate helminth
144 eggs: faecal flotation for light eggs using Sheather's solution (128 g of sugar in 100 ml of hot
145 water) and faecal sedimentation for heavy eggs (Gillespie, 2006). We visualized helminth
146 eggs using a light microscope (40x and 10x objective) and identified them using a taxonomic
147 key (Matsubayashi et al., 1965; Skrjabin et al., 1952). Parasite prevalence is typically defined
148 by looking for eggs in 1 gram of faecal matter. In this study, we used 3 grams of faecal matter
149 because the number of eggs was very low for some parasites.

150

151 **Statistical analysis**

152 We considered four helminth taxa for our study: Strongylid nematodes, *Ascaris* nematodes,
153 *Trichuris* nematodes and cestodes. We also considered three protozoan taxa: *Cryptosporidium*
154 *parvum*, *Giardia lamblia*, and *Entamoeba histolytica*. We analysed eight different parasite
155 response variables: helminth richness (based on the four helminth taxa) and seven parasite
156 prevalence variables (1 variable per taxon).

157

158 We defined the helminth richness as the number of helminth genera found in one faecal
159 sample. We defined the prevalence of a particular parasite taxon as the proportion of faecal
160 samples infected by that parasite taxon (binomial data: 0 = parasite absent, 1 = parasite

161 present). The analyses of the helminth richness and the four helminth prevalence variables
162 were based on the entire dataset (487 faecal samples from 90 individual chimpanzees)
163 whereas the prevalence of the three protozoa prevalence variables were based on a subset of
164 faecal samples that had been processed with respect to these variables (98 faecal samples
165 from 69 individual chimpanzees).

166

167 We used generalized linear mixed-effects (GLME) models to analyse these parasite response
168 variables as a function of the fixed and random factors of interest. We modelled helminth
169 richness and parasite prevalence using Poisson and binomial errors, respectively. The fixed
170 factors included: snare status (control, snare-injured), community membership (Sonso,
171 Waibira), sex (female, male), age class (adult, juvenile, subadult), season (dry, wet) and the
172 presence of road maintenance workers (absent, present). We included this last variable
173 because road workers visited the study area from November 2013 to February 2014 as part of
174 a project to reopen an old logging road for ecotourism. For snare status, we initially included
175 the degree of injury in our analysis, but the sample sizes of the different categories were too
176 low to carry out a meaningful analysis. We modelled chimpanzee identity as a random factor
177 to control for pseudo-replication since some individuals were sampled more than once. We
178 conducted the statistical analyses using R (R Core Team, 2014) (see the document titled
179 “S2_Script.docx” for the R code used). We used the `glmer()` function to run the GLME
180 models. For each parasite variable, we compared the full model (containing all the fixed
181 factors of interest) to the null model (containing none of the fixed factors of interest) to
182 determine the overall significance of the full model. To determine the statistical significance
183 of each fixed factor, we used log-likelihood ratio tests to compare the residual deviance
184 between the full model, which contained all fixed factors, and the reduced model, which was
185 missing the fixed factor of interest. We considered the results significant at an alpha level of

186 0.05. For three parasite taxa (*Trichuris*, *G. lamblia*, and *E. histolytica*), the number of infected
187 faecal samples was so low (6, 7, and 0, respectively) that the R software was unable to
188 calculate the parameter estimates for the models (see Table S1 in the document titled
189 “S3_table.docx”). We therefore did not present these parasite variables in the results section.

190

191 **Results**

192 We collected a total of 487 faecal samples from chimpanzees in the Waibira and the Sonso
193 communities. There were 366 and 121 faecal samples from 70 control and 20 snare-injured
194 chimpanzees, respectively. Of the 90 individuals in this study, 36 belonged to the Waibira
195 community (9 with snare injuries) and 54 to the Sonso community (11 with snare injuries).

196

197 For each of the five parasite variables for which there were enough infected faecal samples to
198 run the analyses, the full model was statistically significant when compared to the null model
199 (Table 1). There was a significant association between snare status and the prevalence of
200 Strongylida (Tables 2 and 3; Figure 1). If we consider an adult female chimpanzee in the
201 Sonso community in the dry season in the absence of road workers as the reference point, the
202 probability that a faecal sample was infected with Strongylida was 81.7% for a control
203 individual and 91.8% for a snare-injured individual. There were no significant differences
204 between snare-injured and control chimpanzees for the other parasite variables: helminth
205 richness, prevalence of *Ascaris*, prevalence of cestodes, and the prevalence of
206 *Cryptosporidium parvum* (Tables 2 and 3).

207

208 Other ecological factors that had effects on the parasitic prevalence variables were the
209 chimpanzee community, the season, and the presence of road workers (Tables 2 and 3). The
210 chimpanzees in the fully habituated Sonso community had a higher prevalence for *Ascaris*

211 and cestodes than the Waibira community. Parasite prevalence was higher in the dry season
212 than the wet season for *C. parvum*. The presence of road workers decreased the following
213 four parasite variables: helminth richness, the prevalence of *Ascaris*, the prevalence of *C.*
214 *parvum* and the prevalence of Strongylida. There was no effect of sex and the class of age on
215 the parasite variables (Tables 2 and 3).

216

217 **Discussion**

218 We found that snare-injured chimpanzees had significantly higher prevalence of Strongylida
219 nematodes than control individuals. Snare injuries had no impact on the helminth richness and
220 the prevalence of *Ascaris*, cestodes and *Cryptosporidium parvum*. The most interesting result
221 of this study was that snare-injured chimpanzees had a higher prevalence of Strongylida than
222 control individuals. There are at least two possible explanations for this result. First, snare-
223 injured individuals have higher exposure to Strongylida parasites than control individuals.
224 Second, snare injuries facilitate the proliferation of these parasites because of a weakened
225 immune system.

226

227 The first hypothesis suggests that snare-injured chimpanzees have higher exposure to
228 helminth parasites than control individuals. Many helminth parasites (e.g. hookworms) have
229 infective stages that penetrate the feet and hands of their chimpanzee hosts. Exposure rates
230 should therefore be correlated with time spent on the ground (Zommers et al., 2013).

231 However, a previous study found that snare-injured individuals spend less time on the ground
232 than control individuals (Munn, 2006). In addition, snare-injured individuals spent more time
233 in small groups (Hermans, 2011) where exposure to ground-dwelling helminth parasites is
234 presumably lower than in large groups. These observations suggest that snare-injured
235 individuals do not have higher exposure rates to ground-dwelling helminth parasites than not

236 snared-injured individuals. However, snare-injured chimpanzees reuse old nests more often
237 than control individuals (Plumptre and Reynolds, 1997) and this behaviour may enhance their
238 probability to encounter parasite-contaminated faecal matter. Thus, it is currently not clear
239 whether snare-injured chimpanzees are more or less likely to encounter helminth parasites
240 than control individuals.

241

242 The second hypothesis suggests that snare injuries directly facilitate intestinal parasite
243 infections. The skin is an important defence against pathogens (Janeway et al., 2001) and
244 snare-induced skin injuries trigger inflammatory responses to heal the wound and to kill
245 invading microbial pathogens (Davis, 2008). Severe snare injuries may thus compromise the
246 immune system and, as a consequence, facilitate intestinal infections due to trade-offs
247 between the different components of the immune response (Sadd and Schmid-Hempel, 2009).
248 The efficacy of the immune system also depends on nutrition, stress, and other factors. Snare-
249 injured individuals may have lower quality nutrition and higher stress levels, which would
250 reduce their immune-competence to fight parasitic infections. In summary, snare-injured
251 individuals may have a higher prevalence of intestinal parasites because their immune systems
252 are less competent.

253

254 Chimpanzees are known to carry multiple cestode species (Abee et al., 2012). We did not
255 identify the cestode parasites to subgroups. As a result, the cestode variable contained many
256 genera. The statistical analysis found no effect of snare injuries on the prevalence of cestode
257 parasites. However, this test is limited in value because we lumped all the cestode taxa into a
258 single group.

259

260 An intriguing question for the future is whether snare-injured chimpanzees, given their higher
261 prevalence of Strongylida infections, are more likely to use medicinal plants than control
262 individuals. Some plants eaten by chimpanzees are used by humans in traditional medicine as
263 treatment for intestinal parasite infections (Ghai et al., 2015; Krief et al., 2005). However,
264 severely injured chimpanzees, generally suffered from impaired feeding efficiency (Stokes
265 and Byrne, 2001), which may prevent them from harvesting medicinal plants and control their
266 parasite infections. Foraging data would be necessary to address this point.

267

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281

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352

353

354 **Figure legends**

355

356 **Fig.1** Prevalence of Strongylida in control and snare-injured chimpanzees (*Pan troglodytes*)
357 in Budongo Forest, Uganda, 2011-2014. The parameter estimates were taken from the
358 generalized linear model analysis (see Table 3) and were back-calculated to the original scale.
359 The effect of snaring is shown for a reference faecal sample, taken from a female adult in the
360 Sonso community during the dry season and in the absence of road workers. The bars show
361 the standard errors.

362

363 **Supporting Information**

364

365 The datasets supporting this article are available as Electronic Supplementary Material.

366 **S1_File.** Table containing the analysed data

367 **S2_R Script.** Example of the R script used to run the statistical analyses

368 **S3_Table.** The means of the 8 parasite variables