A tumor DNA complex aberration index is an independent predictor of survival in breast and ovarian cancer

Hans Kristian Moen Vollan, Oscar M. Rueda, Suet-Feung Chin, Christina Curtis, Gulisa Turashvili, Sohrab Shah, Ole Christian Lingjærde, Yinyin Yuan, Charlotte K. Ng, Mark J. Dunning, Ed Dicks, Elena Provenzano, Stephen Sammut, Steven McKinney, Ian O. Ellis, Sarah Pinder, Arnie Purushotham, Leigh C. Murphy, Vessela N. Kristensen, METABRIC Group, James D. Brenton, Paul D.P. Pharoah, Anne-Lise Børresen-Dale, Samuel Aparicio, Carlos Caldas

Cancer Research UK, Cambridge Institute, University of Cambridge, Li Ka Shing Centre, Robinson Way, Cambridge CB2 0RE, UK
Department of Oncology, University of Cambridge, Hills Road, Cambridge CB2 2XZ, UK
Department of Genetics, Institute for Cancer Research, Oslo University Hospital, The Norwegian Radium Hospital, Montebello, 0310 Oslo, Norway
The K.G. Jebsen Center for Breast Cancer Research, Institute for Clinical Medicine, Faculty of Medicine, University of Oslo, Norway
Department of Oncology, Division for Surgery, Cancer and Transplantation, Oslo University Hospital, The Norwegian Radium Hospital, Montebello, 0310 Oslo, Norway
Department of Preventive Medicine, Keck School of Medicine, University of Southern California, Los Angeles, CA 90033, USA
Department of Pathology and Laboratory Medicine, University of British Colombia, Vancouver, British Colombia V6T 2B5, Canada
Molecular Oncology, British Colombia Cancer Research Center, Vancouver, British Columbia V5Z 1L3, Canada
Biomedical Informatics Division, Department of Computer Science, University of Oslo, Oslo, Norway
Center for Cancer Biomedicine, University of Oslo, Norway
Strangeways Research Laboratories, University of Cambridge, Cambridge CB1 9RN, UK
Cambridge Experimental Cancer Medicine Centre, Cambridge CB2 0RE, UK
Cambridge Breast Unit, Addenbrooke's Hospital, Cambridge University Hospital NHS Foundation Trust and NIHR Cambridge Biomedical Research Centre, Cambridge CB2 2QQ, UK
Department of Histopathology, School of Molecular Medical Sciences, University of Nottingham, Nottingham, NG5 1PB, UK

Abbreviations: BCSS, Breast cancer specific survival; CAAI, Complex arm-wise aberration index; CNA, Copy number alterations; ER, Estrogen receptor; HR, Hazard ratio; HGSOC, High-grade serous ovarian cancer; MIP, Molecular inversion probe; OS, Overall survival; PFS, Progression free survival.

* Corresponding author. Cancer Research UK, Cambridge Institute, Li Ka Shing Centre, Robinson Way, Cambridge CB2 0RE, UK. Tel.: +44 1223 769 650.
** Corresponding author. Department of Pathology and Laboratory Medicine, University of British Colombia, Vancouver, British Colombia V6T 2B5, Canada. Tel.: +1 604 675 8201.
E-mail addresses: saparicio@bccrc.ca (S. Aparicio), carlos.caldas@cruk.cam.ac.uk (C. Caldas).
1 These authors contributed equally to this work.
2 Senior and co-corresponding authors.
3 Full list of members at end of text.

http://dx.doi.org/10.1016/j.molonc.2014.07.019
1574-7891/© 2014 The Authors. Published by Elsevier B.V. on behalf of Federation of European Biochemical Societies. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/3.0/).
1. Introduction

Breast and ovarian cancer are major causes of morbidity and death (Jemal et al., 2010). Current treatment for breast cancer includes a combination of surgery, radiotherapy, chemotherapy, endocrine agents (tamoxifen or aromatase inhibitors) in ER+ cases and trastuzumab in HER2-positive cases (Goldhirsch et al., 2009). Although clinical outcome has improved dramatically with modern therapy, the challenge remains to identify patients that could be spared overtreatment, since at 15 years 58% of patients are alive despite receiving no adjuvant chemotherapy (Early Breast Cancer Trialists’ Collaborative Group (EBCTCG), 2005). Markers that could more precisely predict better clinical outcome are therefore of utmost importance.

Expression profiling of breast cancers has provided important insights into tumor biology and clinical behavior, even though clinical implementation is not yet routine (Perou et al., 2000; Sørlie et al., 2001, 2003). Based on the global pattern of gene expression, five subgroups (Luminal A, Luminal B, HER2-enriched, Basal-like and Normal-like subgroups) were identified, with different biology and clinical course. By combining DNA copy number alterations with gene expression profiles, molecular subtypes with different biology and clinical course (Curtis et al., 2012) were identified. These subtypes of breast cancer with clearly distinct genomic drivers (Curtis et al., 2012) were defined as CE, IC1, IC2, IC3 and IC4.

There is a need for new and informative markers for breast cancer, since at 15 years 58% of patients are alive despite receiving no adjuvant chemotherapy (Early Breast Cancer Trialists’ Collaborative Group (EBCTCG), 2005). Finding such markers could more precisely predict better clinical outcome. Breast and ovarian cancer are major causes of morbidity and death (Jemal et al., 2010). Current treatment for breast cancer includes a combination of surgery, radiotherapy, chemotherapy, endocrine agents (tamoxifen or aromatase inhibitors) in ER+ cases and trastuzumab in HER2-positive cases (Goldhirsch et al., 2009). Although clinical outcome has improved dramatically with modern therapy, the challenge remains to identify patients that could be spared overtreatment, since at 15 years 58% of patients are alive despite receiving no adjuvant chemotherapy (Early Breast Cancer Trialists’ Collaborative Group (EBCTCG), 2005). Markers that could more precisely predict better clinical outcome are therefore of utmost importance.

Expression profiling of breast cancers has provided important insights into tumor biology and clinical behavior, even though clinical implementation is not yet routine (Perou et al., 2000; Sørlie et al., 2001, 2003). Based on the global pattern of gene expression, five subgroups (Luminal A, Luminal B, HER2-enriched, Basal-like and Normal-like subgroups) were identified, with different biology and clinical course. By combining DNA copy number alterations with gene expression profiles, molecular subtypes with different biology and clinical course (Curtis et al., 2012) were defined as CE, IC1, IC2, IC3 and IC4.
infiltration and ER+ tumors with abundant stroma. IC5 identifies most of the HER2 amplified tumors. IC6 is a ZNF-703 driven poor prognosis luminal group. IC7 and IC8 are intermediate-risk Luminal A groups. IC9 is a Luminal B group with 8q gain. Finally IC10 are the core basal-like tumors with high genomic instability. One of our aims was to determine how CAAI affects prognosis in these different groupings.

Gene expression-based prognostic classifiers have been proposed to risk stratify patients (Paik et al., 2004; van de Vijver et al., 2002; van’t Veer et al., 2002), and are already used in some clinical environments, but their implementation in practice awaits results of ongoing trials and will always be problematic given the intrinsic instability of RNA (Borgan et al., 2011). Furthermore it remains questionable if these genomic tests are cost effective (Hall et al., 2012), or if they improve the performance of current prognostic tools that integrate clinical and pathological parameters routinely used in the clinic, such as Adjuvant! (Ravdin et al., 2001) or PREDICT (Wishart et al., 2010, 2011, 2012). PREDICT is an online tool based on population-based cancer registry data, similar to Adjuvant!, but that in addition incorporates mode of detection, HER2 status and trastuzumab benefit (Wishart et al., 2012, 2011, 2010).

Ovarian cancer is the fifth leading cause of cancer related death in US women (Jemal et al., 2010). The majority of these deaths occur in patients with advanced stage, high-grade serous ovarian carcinomas (HGSOC) (Koonings et al., 1989; Seidman et al., 2004), despite optimal cytoreduction by debulking surgery and adjuvant chemotherapy (Ozols, 1997). Major prognostic factors are age at diagnosis, performance status, histology and residual tumor size (Winter et al., 2007, 2008). Recent studies have sought to develop molecular classifications, but these do not significantly improve prognostic performance (Cancer Genome Atlas Research Network, 2011; Etemadmoghadam et al., 2009).

Genomic rearrangements in breast cancer have distinct patterns, possibly reflecting different mechanisms of genomic instability (Hicks et al., 2006). Analysis of somatically acquired copy number alterations (CNA) has identified distinct types of structural changes (e.g. whole-arm alterations and firestorms) (Hicks et al., 2006; Russnes et al., 2010). Clustered narrow peaks of high copy number gains characterize ‘firestorms’. We developed a score, the Complex Arm-wise Aberration Index (CAAI), to quantify such complex events (Russnes et al., 2010). In our original report CAAI was shown to have independent prognostic power (Russnes et al., 2010), but this finding needs to be independently validated. Here we report such independent validation of the prognostic value of CAAI in 1950 breast carcinomas (Curtis et al., 2012), and show that it adds to the gene expression-based prognostic classifiers (the 70-gene and the 21-gene signatures) (Paik et al., 2004; van de Vijver et al., 2002). We also evaluate CAAI as a prognostic marker in 508 advanced stage HGSOC (Cancer Genome Atlas Research Network, 2011).

2. Materials and methods

2.1. Breast cancer cases

A total of 1950 breast cancer cases from the METABRIC (Molecular Taxonomy of Breast Cancer International Consortium) cohort (Curtis et al., 2012) were included in this study. Female breast cancer cases were selected on the basis of invasive histology (in-situ and benign cases were excluded, as well as cases with unknown histology). Using an eQTL-based approach (Lynch et al., 2012) a few cases with mismatched DNA/RNA were identified, and excluded. This resulted in 1950 cases with gene expression, SNP-array and clinical data available for analysis (flow chart of included samples in Appendix A). Clinical and pathological variables were collected from hospital reports. Estrogen receptor status by IHC (immunohistochemistry) was available for 1921 samples and ER status for the remaining 29 samples was scored by the expression value of ESRI (Lehmann et al., 2011). IHC progesterone receptor status was not available, hence expression values for PGR were used to score PR status (Lehmann et al., 2011). HER2 status was obtained from segmented copy number data as described in the original METABRIC report (Curtis et al., 2012). Clinical variables are presented in Supplementary Table 1.

2.2. Serous ovarian carcinomas

A total of 508 high-grade serous ovarian adenocarcinomas, recruited at the time of primary surgery, were obtained from TCGA (the Cancer Genome Atlas) (NCBI/TCGA project number 2459) (Cancer Genome Atlas Research Network, 2011). A description of the clinical variables is presented in Supplementary Table 5.

2.3. Bioinformatic and statistical analyses

CNA profiles were obtained from Affymetrix Genome-Wide Human SNP Array 6.0 (Affymetrix, Santa Clara, CA, USA). Estimation of raw copy number from arrays was performed using the CRMav2 method implemented in the "aroma.affymetrix" R-package (Bengtsson et al., 2009). For the SNP-probes the two alleles were summarized to obtain an estimate of the total copy number. Normalized probe intensity ratios were obtained with matched normal DNA for cases where such data was available, otherwise an average of a pool of 473 adjacent normal tissue samples and 270 HapMap samples were used. Raw normalized copy number estimates were segmented into regions of constant copy number with PFC (Piecewise Constant Fit) with the smoothing parameters kmin = 5 and gamma = 200 (Nilsen et al., 2012). CAAI was then scored as previously described (Russnes et al., 2010). The method computes scores that capture complex rearrangements for each chromosomal arm separately, based on the segmented copy number data (Nilsen et al., 2012), and calls the tumor CAAI positive if the score exceeds the set threshold in at least one chromosomal arm. Gene expression data was available for the METABRIC cases and used to derive the PAM50 subtype classifications (Curtis et al., 2012), as well as representations of the 21-gene (Paik et al., 2004) and 70-gene prognostic classifiers (van’t Veer et al., 2002). All statistical analyses were performed using R version 2.15.0 with the packages ’copynumber’ and ’rms’ (Gentleman et al., 2004; Harrell, 2014; Nilsen et al., 2012). A subset of 32 samples from METABRIC was also profiled with Molecular Inversion Probe (MIP) arrays, a method for copy number profiling optimized for use with paraffin-
CAAI was also tested as a prognostic marker using the breast cancer prognostic tool PREDICT (www.predict.nhs.uk) (Wishart et al., 2012, 2011, 2010). The predictive performance of CAAI in the ovarian cohort was assessed by internal validation to correct for potential overfitting, given no adequate independent dataset for validation was available, using the bootstrap resampling technique (Halabi et al., 2003; Harrell, 2001). The study complies with the Reporting Recommendations for Tumor Markers (REMARK) (McShane et al., 2005). A detailed description of materials and methods can be found in Appendix A (Supplementary material and methods).

3. Results

3.1. Breast cancer

3.1.1. CAAI distribution and correlation with overall genomic instability

A total of 835 (43%) of the 1950 breast cancer cases were CAAI positive (Supplementary Figure 1). CAAI positivity was significantly associated with larger tumors, higher grade, negative ER and PR (progesterone receptor), and amplification of the HER2 gene (Supplementary Table 1). CAAI events were most frequent on 11q in ER- cases and 17q in ER+ cases. Hierarchical clustering of the CAAI positive cases based on binary dissimilarity and Ward’s method, revealed groups dominated by 1q, 8p, 11q or 17q complex alterations, and with a tendency for mutually exclusivity (Figure 1A). The proportion of CAAI positive cases in the 1950 breast cancer cases was variable across both the ten integrative subtypes recently described (Curtis et al., 2012) and across the five expression-based intrinsic subtypes (Perou et al., 2000) (Figure 1B and C).

CAAI status was significantly different distributed in the intrinsic subtypes, with CAAI positive tumors most frequently found in HER2-enriched (61.8%) and Luminal B (59.9%) subgroups. The frequency of CAAI positive tumors in Basal-like, Luminal A and Normal-like subgroups were 40.8%, 30.4% and 26.3% respectively. CAAI status was associated with higher mortality within each of the intrinsic subtypes, except for Luminal B tumors (Supplementary Figure 3). CAAI was also prognostic in IC4, IC7 and borderline significant in IC10 (Supplementary Figure 4). The HER2-enriched Pam50 subgroup overlap with, but is not equal to the clinical HER2 amplified group. In IC5 all tumors have HER2 gene amplification, but not all HER2+ tumors belong to IC5. CAAI was prognostic in HER2 amplified tumors (by SNP-array) and in the Pam50 HER2-enriched group, but not in IC5 subgroup (Supplementary Figure 5). A total of 64.4% of the HER2 amplified tumors by SNP-array were CAAI positive. Interestingly only 139 of the 279 CAAI positive HER2 amplified tumors (49.8%) had a complex alteration affecting chromosome 17q, the remainder had complex events elsewhere in the genome, most frequently affecting 11q and 8p. This shows that not all HER2 amplified cases have a complex structural alteration affecting the HER2 amplicon, but rather a more simple amplification.

The CAAI score is a continuous numeric value (range 0.0–22.4 in the 1950 breast cancer cases). To classify samples as CAAI positive and negative (dichotomous variable) we used the same threshold value of 0.5 as in our original report (Russnes et al., 2010). Using a log2 transformation of the continuous CAAI as a variable in a Cox regression model showed an HR of 1.46 (95%CI, 1.33–1.61; p < 0.001). This significant association between the continuous numeric CAAI score and outcome illustrates that results are not dependent on the chosen threshold, but since the present study is an independent validation of the prognostic value of CAAI, we used the dichotomized variable for all the multivariable models (see below).

3.1.3. Multivariable survival analysis

Results from multivariable Cox regression models for BCSS are presented in Table 1 (and for OS are shown in Supplementary Table 3). When comparing ER+ and ER- tumors the proportional hazard varies over time (Blows et al., 2010). We therefore fitted separate Cox models for ER+ and ER- cases. In the 1412 ER+ cases CAAI status, tumor size, lymph node status, age, histologic type (ILC vs. IDC) and the surrogate 21-gene predictor were significant after adjustment for covariates. Positive CAAI had an HR of 1.56 (95%CI, 1.23–1.99; p < 0.001). A total of 421 ER- cases were available for multivariable analysis, and only axillary lymph node status, histological type and CAAI status remained significant (HR 1.55 (95%CI, 1.11–2.18; p = 0.011)). Neither of the two expression-based signatures (that is surrogates to resemble Mamma Print and OncoTypeDX, see below), was found to be significant in ER- disease.

3.1.4. CAAI provides additional stratification to gene expression classifiers

The prognostic value of the two gene expression prognostic classifiers (Paik et al., 2004; van de Vijver et al., 2002) was
tested in the 1950 breast cancer cases using surrogate 21-gene and 70-gene signatures as described in supplementary information and presented in Supplementary Figure 6. CAAI status was able to further stratify each of the gene expression risk groups (Supplementary Figure 7). CAAI provides added prognostic information in all subsets, with the strongest effect in the high-risk groups. To evaluate the added predictive ability of a new biomarker, such as CAAI, is a complex task, as statistical significance is not always the same as clinical significance (Pencina et al., 2008). For this reason, we compared the performance of prognostic models in the complete dataset, and in ER⁺ and ER⁻ disease separately, using three different statistical measures proposed in the literature: the likelihood ratio test, the C-index (Harrell et al., 1996), a measure of the concordance between predicted and observed survival times, and finally U-statistics based on net classification improvements (Pencina et al., 2011), to formally test the significance of the increase in the C-index. The results of these analyses, presented in Supplementary Table 4, confirm that a model including CAAI and the two expression-based
prognostic signatures outperforms a model including the 21-gene and 70-gene signatures but excluding CAAI.

### 3.2. Ovarian cancer

In HGSOC CAAI positivity was detected in 262 of the 508 (52%) available cases. The distribution of clinical variables in the data set is shown in Supplementary Table 5. The CAAI scores were generally higher in the ovarian cohort than in breast cancer. Univariable Cox regression of the continuous CAAI-score (log2 transformed) showed a significant HR of 1.3 (95%CI, 1.2–1.5; p < 0.01). We chose a threshold of 1.0 for the subsequent analyses to reflect the more rearranged genome of ovarian cancer. Kaplan–Meier plots illustrate the prognostic impact of CAAI status on progression free survival (PFS) (p = 0.013) and OS (p < 0.001) (Figure 4). Univariable Cox regression models for all clinical variables are presented in Supplementary Table 6. CAAI positive cases had an increased risk of progression with HR 1.3 (95%CI, 1.1–1.6; p = 0.01) and for death of any cause with HR 1.4 (95%CI, 1.2–1.7; p < 0.01).

The results from multivariable Cox regression analyses are presented in Table 2. Age at diagnosis did not meet the proportional hazards (PH) assumption and the model was stratified by a categorical representation of age (<60 vs. >60). Positive CAAI increased risk of relapse, independent of other covariates with a HR of 1.3 (95%CI, 1.1–1.7; p = 0.01). CAAI was together with histological grade a predictor of OS, with HR of 1.3 (95%CI, 1.1–1.6; p < 0.01). Since no adequate external/independent dataset was available, an internal validation of the multivariable models was performed using bootstrapping to correct for potential over-fitting (Halabi et al., 2003; Harrell, 2001). Supplementary table 7 shows the result of the validation using several indexes (details in supplementary material). The amount of over-fitting in the models was modest, as reflected by the estimated optimism. Supplementary Figure 8 shows the predictions from the models at 1 and 2 years of follow up compared with the actual survival probability (calibration accuracy). The agreement between the observed and predicted survival was highly concordant; mean optimism 0.002 for PFS at both time points and 0.003 and –0.002 for OS at 1 and 2 years of observation respectively.

### 3.3. Possible clinical utility of CAAI in breast cancer

#### 3.3.1. CAAI in FFPE tumor material

Most aCGH-based analyses rely on availability of fresh frozen tumor material. Molecular inversion probe (MIP) arrays have been developed for analysis of FFPE material and this could facilitate clinical implementation of CAAI analysis. We therefore compared 32 samples hybridized to both Affymetrix SNP 6.0 and MIP platforms. Supplementary Figure 9 shows the good correlation between CAAI scores obtained with the two platforms in matched fresh-frozen and paraffin-embedded samples. Using an optimized threshold for the MIP data the agreement between methods was substantial; Cohen’s Kappa value of 0.75. These preliminary results suggest that CAAI scoring could be implemented using formalin fixed paraffin-embedded tumor material.
Figure 3 — Kaplan–Meier estimates of outcome in breast cancer. In A and B, survival estimates for CAAI positive and negative cases are shown for breast cancer specific survival (BCSS) and overall survival (OS) respectively. Outcome for lymph node negative and lymph node positive cases are shown in C and D. E and F shows survival estimates for ER+ and ER− cases.
3.3.2. Implementation of CAAI in the prognostic tool PREDICT (Wishart et al., 2012, 2011, 2010)

CAAI as a prognostic marker was added to the prognostic model PREDICT (www.predict.nhs.uk) (Wishart et al., 2012, 2011, 2010), which is based on a Cox proportional hazard model. PREDICT was developed using a case-cohort of breast cancer cases of unknown CAAI status and so the underlying, baseline hazard is representative of cases of average CAAI status. The CAAI hazard ratio estimate is for CAAI positive compared to CAAI negative cases, and so these were rescaled to give an average hazard ratio of unity using an estimated prevalence of CAAI positivity of 40 percent. We then compared the performance of PREDICT with and without the addition of CAAI in predicting breast cancer specific mortality at five years after diagnosis using calibration, discrimination and reclassification as measures of model performance. Model calibration is a comparison of the predicted mortality estimates from each model with the observed mortality. Model discrimination was evaluated by calculating the area under the receiver-operator-characteristic curve (AUC) calculated for 5-years breast cancer specific mortality. This is a measure of how well the models identify those patients with poorer survival. The AUC is the probability that the predicted mortality from a randomly selected patient who died will be

### Table 1 – Multivariable Cox regression model with breast cancer specific survival as outcome variable.

<table>
<thead>
<tr>
<th>Variable</th>
<th>ER positive cases (n = 1412)</th>
<th>ER negative cases (n = 421)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>HR</td>
<td>95% CI Lower</td>
</tr>
<tr>
<td>Any genomic complex events</td>
<td></td>
<td>1.56</td>
</tr>
<tr>
<td>CAAI pos vs. neg</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Histological grade</td>
<td></td>
<td>1.13</td>
</tr>
<tr>
<td>Categories as ordinal</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tumor size</td>
<td></td>
<td>1.50</td>
</tr>
<tr>
<td>pT2 vs. pT1</td>
<td></td>
<td>3.62</td>
</tr>
<tr>
<td>pT3 vs. pT1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Axillary lymph node status</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Positive vs. negative</td>
<td></td>
<td>2.27</td>
</tr>
<tr>
<td>HER2 status (from arrays)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Positive vs. negative</td>
<td></td>
<td>0.99</td>
</tr>
<tr>
<td>Progesterone receptor status</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Positive vs. negative</td>
<td></td>
<td>1.05</td>
</tr>
<tr>
<td>Age at diagnosis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Continuous variable</td>
<td></td>
<td>1.01</td>
</tr>
<tr>
<td>Histological type</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ILC vs. IDC</td>
<td></td>
<td>1.73</td>
</tr>
<tr>
<td>Other invasive vs. IDC</td>
<td></td>
<td>1.05</td>
</tr>
<tr>
<td>70-gene classifier</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Poor vs. good prognosis</td>
<td></td>
<td>1.25</td>
</tr>
<tr>
<td>21-gene classifier</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Moderate vs. good prognosis</td>
<td></td>
<td>1.23</td>
</tr>
<tr>
<td>Poor vs. good prognosis</td>
<td></td>
<td>2.35</td>
</tr>
</tbody>
</table>

Separate models for ER positive and negative cases. Significant p-values (<0.05) in bold. Both models were stratified for site of inclusion.

![Figure 4](image-url) — Kaplan–Meier estimates of outcome in ovarian cancer. Survival estimates for CAAI positive vs. CAAI negative cases with progression free survival (A) and overall survival (B).
higher than the predicted mortality from a randomly selected survivor. Reclassification is the extent to which a new model classified individuals into specified risk groups. PREDICT is used in a clinical setting to identify patients most likely to benefit from adjuvant chemotherapy. The Cambridge Breast Unit multi-disciplinary team uses the estimated absolute benefit of chemotherapy at ten years to define three patient groups: those with a predicted absolute benefit of less than 3 percent in whom chemotherapy is not recommended, those with a predicted absolute benefit of 3–5 percent in whom the balance of risks and benefits are discussed with the patients, and those with an absolute benefit of greater than 5 percent for whom chemotherapy is generally recommended. The absolute benefit is approximately proportional to the absolute risk. We therefore classified the 1950 cases from the breast cancer cohort into three groups based on thresholds for estimated absolute benefit at five years of 1.5 percent in whom chemotherapy is not recommended, those with a predicted absolute benefit of 3 percent in whom chemotherapy is recommended, and those with an absolute benefit of greater than 5 percent in whom chemotherapy is generally recommended.

The model was stratified for age. Significant p-values (<0.05) in bold. 
n = 368 (38 excluded due to missing data).

### 4. Discussion

In this study we validate CAAI, a measure of complex structural alterations in cancer genomes, as an independent prognostic biomarker in breast cancer, and reveal its significant prognostic value also in ovarian cancer. The size of the METABRIC breast cancer cohort (1950 cases) and the comprehensive clinical data available enabled stratification by relevant subgroups, and showed that CAAI is an independent predictor of outcome in both ER+ and ER− disease. The prognostic effect was found in both chemotherapy naive and chemotherapy treated patients. The novel finding of the prognostic effect of complex focal DNA copy number change in ovarian cancer, captured by CAAI, is interesting as the method was developed for breast cancer. Since it has proven challenging to identify prognostic markers in ovarian cancer, the modest effect of CAAI needs independent validation.

Analyses of copy number data rely on a robust smoothing of the raw data into estimates of the underlying copy number state. Different algorithms for such analysis exist and the individual methods have different settings to control the sensitivity and the specificity of calls. In this analysis PCF was used after normalization of CEL files, to make it as similar to our previous results as possible. This was essential since our aim was to validate the prognostic value of CAAI in breast cancer. Threshold parameters were kept constant for the validation in breast cancer, but were adjusted in ovarian cancer due to the more aneuploid background of these genomes. As for most novel tests, thresholds for calling of positives over negatives could be optimized and improved. The prognostic effect of CAAI used as a continuous score, both in breast and ovarian cancer, was clear, indicating that results are not dependent on the thresholds used.

Because we had matched gene expression profiling data for all the 1950 breast cancers cases we could use this data to implement surrogate indices for the 21-gene and 70-gene prognostic gene expression classifiers that are currently being validated in two large clinical trials; MINDACT (Cardoso et al., 2007, 2008; Rutgers et al., 2011) (ClinicalTrials.gov nr.
NCT00433589) and TAILORx (Sparano, 2006; Sparano and Paik, 2008) (ClinicalTrials.gov nr. NCT00310180). We could therefore show, using a multivariable Cox regression model, that the prognostic value of CAAI was independent of these gene expression classifiers. Moreover we could also robustly evaluate, using three statistical measures, the added value of CAAI in a model that also included the 21 and 70 gene expression signatures. These results indicate that even if MINDACT or TAILORx demonstrate the clinical utility of gene expression classifiers, CAAI would add further prognostic information and therefore have clinical impact.

CAAI was designed to capture focal complex copy number aberrations. Our comparison with overall genome instability, measured by GII and number of breakpoints, shows that CAAI is only modestly correlated with these. This finding suggests that there are distinct biological processes underlying these distinct patterns. The CAAI index captures localized regions of copy number change, but in a genomic profile that is characterized by extensive structural variation, the index may call other patterns as well. On the other hand the method is sensitive to score genomes that overall have few changes, but with distinct localized clustered events. A total of 35.6% (154/433) of the HER2 amplified tumors (by SNP-array) are CAAI negative, showing that a single amplicon containing a known driver gene does not necessarily result in a sample being called CAAI positive. Luminal B and the HER2-enriched tumors are more often CAAI positive than others. Intriguingly CAAI was not significantly associated with survival in Luminal B tumors. We noted that curves in the KM-plot joined towards the end of the 15 years observation time, probably reflecting non-breast cancer mortality with very long follow up time.

The exact mechanism by which firestorms affect clinical outcome is not known. Firestorms are thought to arise from breakage-fusion-bridge cycles, either associated with fragile sites, or as result of recombination at palindromic sites at shortened telomeres (Hicks et al., 2006), resulting in focal amplifications, frequently at sites of known oncogenes such as ERBB2 or CCND1. These genomic events could lead not only to increased tumor aggressiveness (proliferation, invasive-ness and metastasis) but also underpin clonal evolution as a driver of drug resistance (Aparicio and Caldas, 2013). All of these features would combine to portend worse outcome for CAAI positive breast and ovarian cancers.

The potential clinical utility of CAAI was demonstrated in breast cancer, showing it could be implemented without requiring fresh frozen tumor material, and improving calibration and discrimination of the clinically used PREDICT prognostication tool by the addition of CAAI. A test based on DNA extracted from formalin fixed paraffin-embedded tissue blocks, rather than RNA from fresh frozen tumor material, would significantly facilitate implementation in routine diagnostic labs.

Acknowledgments

We thank all the patients that contributed to the study. The authors also thank Peter Van Loo, Hege G Russnes and Gro Nilsen for support, help with analyses and fruitful discussions. The results published here are in part based upon data generated by The Cancer Genome Atlas pilot project established by the NCI and NHGRI (project 2459). Information about TCGA and the investigators and institutions who constitute the TCGA research network can be found at http://cancergenome.nih.gov/. Funding: METABRIC was funded by Cancer Research UK and the British Columbia Cancer Foundation. HKMV received funding from European Association for Cancer Research (EACR) travel fellowship award 2010, Kjøpmann Einar Unsgaard og hustru Kitty Unsgaards legat 2010 and The Norwegian Radium Hospital Foundation. This study is a part of the EUROCAN platform (FP7, Grant number 260791). This work was partly supported by the Research Council of Norway through its Centres of Excellence funding scheme, project number 179571. AP has received financial support from the Department of Health via the NIHR Comprehensive Biomedical Research Centre award to Guy’s and St Thomas’ NHS Foundation Trust in partnership with King’s College London and King’s College Hospital NHS Foundation Trust. SA has received funding from BC Cancer Foundation, Canadian Breast Cancer Foundation BC/Yukon, and Canada Research Chairs Program.

Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.molonc.2014.07.019.

The METABRIC Group

Co-chairs Carlos Caldas1,2, Samuel Aparicio3,4

Steering committee James D. Brenton1,2, Ian Ellis5, David Huntsman3,4, Sarah Pinder6, Arnie Purushotham6, Leigh Murphy7, Carlos Caldas1,2, Samuel Aparicio3,4

Tissue and clinical data source sites: University of Cambridge/Cancer Research UK Cambridge Research Institute Carlos Caldas (Principal Investigator)1,2, Helen Bardwell2, Suet-Feng Chin1,2, Christina Curtis1,2, Zhihao Ding2, Stefan Grafl2, Linda Jones9, Bin Liu12, Andy G. Lynch1,2, Irene Papa- theodorou1,2, Stephen J. Sammut8, Gordon Wishart2, British Columbia Cancer Agency Samuel Aparicio (Principal Investigator)1,4, Steven Chia4, Karen Gelmon4, David Huntsman3,4, Steven McKinney3,4, Caroline Speers4, Gulisa Turashvili3,4, Peter Watson3,4,7, University of Nottingham Ian Ellis (Principal

Author contributions

HKMV, OMR, PDPP and CC lead the analysis. HKMV and CC wrote the manuscript with contributions from all authors. SF-C, ChC, GT, SS, OCL, YY, CKN, MD, ED, VNK, SM contributed with data collection and data analysis. GT, EP, SIs, IOL, SP, AP and LCM provided clinical and pathological expertise. JDB, PDPP, ALBD, SA and CC supervised data collection and analysis. SA and CC co-conceived and oversaw the METABRIC study and are joint senior authors and project co-leaders.
Investigator)5, Roger Blamey5, Andrew Green5, Douglas Macmillan6, Emaid Rakha5; King’s College London Arnie Purushotham (Principal Investigator)6, Cheryl Gillett6, Anita Grigoriadis6, Sarah Pinder6, Emanuele di Rinaldi6, Andy Tutt6; Manitoba Institute of Cell Biology Leigh Murphy (Principal Investigator)7, Michelle Parisien7, Sandra Troup7

Cancer genome/transcriptome characterization centers: University of Cambridge/Cancer Research UK Cambridge Research Institute Carlos Caldas (Principal Investigator)1,2, Suet-Feung Chin (Team Leader)1,2, Derek Chan1, Claire Fielding2, Ana-Teresa Maia1,2, Sarah McGuire2, Michelle Osborne2, Sara M. Sayalero 2, Inmaculada Spiteri2, James Hadfield2; British Columbia Cancer Agency Samuel Aparicio (Principal Investigator)3,4, Gulisa Turashvili (Team Leader)3,4, Lynda Bell5, Katie Chow5, Nadia Gale5, David Huntsman3,4, Maria Kovalik5, Ying Ng6, Leah Prentice5


