- 1 Incorporating non-equilibrium dynamics into demographic history inferences of a migratory
- 2 marine species
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46 ABSTRACT: 250 words

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Understanding how dispersal and gene flow link geographically separated populations over evolutionary history is challenging, particularly in migratory marine species. In southern right whales (SRWs, Eubalaena australis), patterns of genetic diversity are likely influenced by the glacial climate cycle and recent history of whaling. Here we use a dataset of mitochondrial DNA (mtDNA) sequences (n=1,327) and nuclear markers (17 microsatellite loci, n=222) from major wintering grounds to investigate circumpolar population structure, historical demography, and effective population size. Analyses of nuclear genetic variation identify two population clusters that correspond to the South Atlantic and Indo-Pacific ocean basins that have similar effective breeder estimates. In contrast, all wintering grounds show significant differentiation for mtDNA, but no sex-biased dispersal was detected using the microsatellite genotypes. An approximate Bayesian computation (ABC) approach with microsatellite markers compared scenarios with gene flow through time, or isolation and secondary contact between ocean basins, while modeling declines in abundance linked to whaling. Secondary-contact scenarios yield the highest posterior probabilities, implying that populations in different ocean basins were largely isolated and came into secondary contact within the last 25,000 years, but the role of whaling in changes in genetic diversity and gene flow over recent generations could not be resolved. We hypothesise that these findings are driven by factors that promote isolation, such as female philopatry, and factors that could promote dispersal, such oceanographic changes. These findings highlight the application of ABC approaches to infer connectivity in mobile species with complex population histories and currently low levels of differentiation. Keywords: southern right whale; approximate Bayesian computation; historical demography; connectivity; last glacial maximum; non-equilibrium population genetic; Eubalaena australis

INTRODUCTION

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Migratory marine species, such as sea turtles, sharks, sea birds and marine mammals, are by nature highly mobile, and many lack obvious barriers to long-distance movement (e.g., Benson et al., 2011; Bonfil et al., 2005; Mate & Best, 2011). Despite this, many such species also have geographically subdivided populations. A key question when managing contemporary populations of migratory marine species is whether such geographically distinct populations are linked by current dispersal or gene flow, or are now isolated and experiencing genetic drift. Such questions are of interest in understanding the evolution of population structure and are also of importance for management, given the disproportionate number of migratory marine species that are of conservation concern (Croxall et al., 2012; Dulvy et al., 2014; Hoffmann et al., 2010; IUCN, 2017). When using genetic data to investigate such questions, it is recognized that patterns of genetic variation within and among populations result from the interplay of evolutionary forces through time. Consequently, to understand the potential genetic impacts of contemporary anthropogenic processes, it is important to ask how the current genetic structure of a species reflects its history of population dynamics and adaptive challenges (Jobling, 2012). The demographic histories of migratory marine species have been strongly influenced by oceanographic and climatic processes that shape the distribution of primary productivity and prey resources (Pastene et al., 2007, Bowen et al., 2016) and suitable habitats (Fontaine et al., 2010; Munro & Burg, 2017), as well as a species' dispersal strategy and capacity (Bowen et al., 1994; Munro & Burg, 2017; Pichler et al., 2001; Veríssimo et al., 2017). Life history traits, such as philopatry (Andreotti et al., 2016; Bowen et al., 2016), social organisation (Baker et al., 1993; Palsboll et al., 1995; Whitehead, 1998) and mating systems (Palsbøll et al., 2010; Hoelzel, 1999), also influence patterns of genetic diversity in natural populations of migratory marine species.

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Recent anthropogenic impacts, such as direct hunting and habitat degradation, have also affected contemporary patterns of genetic diversity in such species (e.g., Pinsky & Palumbi, 2014). However, interpreting the impact of recent anthropogenic activity on longest-lived migratory marine species, such as baleen whales, is controversial (Alter et al., 2012; Attard et al., 2015). The long and overlapping generation times of baleen whales (20+ years: Taylor et al., 2007) may have slowed the loss of genetic diversity during the demographic bottleneck caused by whaling, which was severe for many species but relatively short-lived.

Here we focus on a migratory baleen whale species with a circumpolar distribution (Fig. 1, IWC, 2001; Richards, 2009), the southern right whale (SRW: Eubalaena australis). During winter, SRWs typically inhabit shallow, sheltered coastal areas at mid-latitudes, where females calve and both sexes socialize. During spring, SRWs migrate to offshore summer feeding grounds in mid- to high-latitudes (IWC, 2001). Given these habitat preferences, we suggest that the historical demography of SRWs has been influenced by the transition from the last glacial maximum (LGM) to the Holocene, during which sea levels rose dramatically, primary productivity increased and Antarctic sea ice cover decreased while becoming more seasonal (Allen et al., 2011; Bentley et al., 2014; Clark et al., 2009; Denis et al., 2009; Gersonde et al., 2005; Scourse, 2013). These changes impacted the shallow, near-shore marine environment used as SRW wintering areas: many such areas would have become unsuitable due to sea level rises while newer, larger potential wintering areas were created during the expansion of shallow marine habitat (Scourse, 2013). These disruptions could have plausibly precipitated increased dispersal rates among SRW wintering grounds, leading to secondary contact between previously isolated populations.

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Furthermore, whaling, and the species' subsequent recovery, may have influenced connectivity between wintering grounds (Allendorf et al., 2008). Between the 18th and 20th centuries, whalers killed an estimated 150,000 or more SRWs, driving a hemispheric population size decline from ~100,000 whales to possibly fewer than 400 whales by 1920 (IWC, 1986, 2001, 2012; Jackson, et al., 2008). Covert Soviet whaling from 1951-1971, in violation of international protection from the League of Nations introduced in 1931 (IWC, 1986), further slowed the species' recovery: the 3,368 SRWs killed (Tormosov et al., 1998) comprised an estimated 50% of the hemispheric population size at that time (Jackson et al., 2008). Today, the species has recovered in some parts of its former range. Large aggregations now occur in some key calving/nursery areas (e.g. Argentina, South Africa, Australia; see Fig. 1), while regular sightings of small numbers of SRWs occur in other parts of the historical range (Bannister, 1990; Best, 1990; Carroll et al., 2013; Cooke et al., 2001; Groch et al., 2005; Patenaude et al., 1998). The latter areas could represent remnant populations (e.g. Chile/Peru: Reilly et al., 2008) and/or areas that are undergoing recolonization from larger wintering aggregations (e.g. mainland New Zealand: Carroll et al., 2014). Recolonisation and asymmetric migration rates could have resulted from the differential rates of recovery shown by SRW wintering grounds (IWC, 2001), promoting higher levels of connectivity in the aftermath of whaling. Genetic studies of several extant SRW populations, defined by current calving grounds, using a short fragment of the mitochondrial (mtDNA) control region (275 bp), showed hierarchical population structure, indicating limited connectivity between the South Atlantic and Indo-Pacific (Patenaude et al., 2007). Female philopatry was invoked as a major cause of this pattern, as long-term studies of individually identified SRWs show long-term fidelity to natal wintering grounds (e.g., Carroll et al., 2016; Rowntree et al., 2001). Subsequent studies

integrating stable isotope and genetic data of contemporary populations, suggest some degree of maternally-directed learning of both wintering and summer feeding grounds (Carroll et al., 2015; Valenzuela et al., 2009).

Here we build on previous work using mtDNA haplotype sequences from 1,327 individuals (~10% of the current global population) and 17 nuclear DNA microsatellite genotypes for 222 individuals (~2% of the population), allowing both mtDNA and nuclear DNA diversity and population structure to be inferred for the first time on a circumpolar scale in the SRW. This extensive dataset allows us to begin to disentangle contemporary and historical factors that account for observed patterns of genetic variation, thereby illuminating the complex population dynamics of this widely distributed species. Specifically, we make inferences about past and current patterns of gene flow, while taking into account non-equilibrium population dynamics, and provide information for conservation and management of this species now recovering from centuries of whaling.

We use approximate Bayesian computation (ABC) to evaluate the relative power of alternative historical scenarios to explain the phylogeographic pattern of the mtDNA haplotypes previously described (Patenaude et al., 2007), and the potential impact of whaling on connectivity. Patenaude et al., (2007) described a mtDNA phylogeographic pattern for SRW consistent with two competing hypotheses: (A) random lineage sorting in a species with continuous gene flow or (B) secondary contact between formerly isolated populations (Avise, 2000). Under hypothesis (A), we posit that there was continued gene flow, potentially malebiased, between wintering grounds after population divergence. Under hypothesis B, we posit that the wintering grounds in the South Atlantic and Indo-Pacific became isolated following

divergence, but came into secondary contact as a result of the environmental change following the last glacial maximum (LGM, 16-20,000 years before present: Clark, 2009).

We interpret our results in the context of two different conservation management frameworks. The first framework (Wade & Angliss 1997) defines subpopulations or stocks as groups for which demographic processes operating within the group are more important for persistence than immigration from other subpopulations. The second framework (Crandall et al., 2000) views subpopulations as groups that can be defined on the basis of contemporary and historical ecological and genetic exchangeability.

MATERIALS AND METHODS

Genetic data generation and compilation

For both the microsatellite and mtDNA control region analyses, we used a combination of previously published data sources (Carroll et al., 2015; Valenzuela et al., 2009) and new data (Table 1, Supplementary Material 1, Supplementary Table 1). Allele calls were standardized between laboratories for the microsatellite data, and standard quality control measures were taken including tests for deviation from Hardy-Weinberg equilibrium and genotyping error rate estimation (Supplementary Material 1). All sampling areas were nursery grounds except for the Australian wintering habitat, which is a mixture of migratory corridors and winter nursery areas (Carroll et al., 2015). Therefore, we make some comparisons using samples from the southeast and southwest Australian nursery grounds only, and others using the entire Australian wintering habitat sample.

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Estimates of genetic diversity We estimated standardized allelic richness with FSTAT v 2.9 (Goudet, 1995), and estimated observed and expected heterozygosity for each microsatellite locus per sample partition using GenoDive v2.0 (Meirmans & van Tienderen, 2004). We estimated haplotype and nucleotide diversity for the mtDNA sequence data using Arlequin v3.5 (Excoffier & Lischer, 2010), and tested for significant differences in these statistics between sample partitions using permutation tests (Alexander et al., 2016). To obtain comparable estimates of the number of haplotypes detected between sample partitions, we randomly selected 12 individuals from each partition, with replacement, 1000 times, to estimate the mean number of haplotypes (and its standard deviation). We estimated the apparent contemporary N_e from the microsatellite genotypes using the bias-corrected version of the linkage disequilibrium method (Waples, 2006), as implemented in program NeEstimator v2.01 (Do et al., 2014). *Investigating contemporary patterns of genetic diversity* We undertook an hierarchical Analysis of Molecular Variance (AMOVA (Excoffier et al., 1992)) for both the mtDNA and microsatellite data in *Arlequin*, with the wintering grounds grouped into ocean basins, the significance of which was assessed with a permutation test (50,000 permutations, $\alpha = 0.05$). We estimated pairwise genetic differentiation between sample partitions for the microsatellite loci by calculating overall and pairwise F_{ST} and Jost's D statistic (Jost, 2008) using GenoDive and for the mtDNA data by calculating overall and pairwise F_{ST} and Φ_{ST} statistics using Arlequin. The probability of the observed level of differentiation occurring in a panmictic population was estimated using the log-likelihood G test in GenoDive (microsatellite) and the permutation test in Arlequin (mtDNA), for a total of 10,000 permutations each.

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We used two complementary methods to detect genetic clusters within the microsatellite data: discriminant analysis of principal components (DAPC) using the R package adegenet (Jombart & Ahmed, 2011) and STRUCTURE v2.3.4 (Pritchard et al., 2000). DAPC is a generic multivariate method that makes few assumptions about the underlying data and seeks to maximise between-group variation while minimising within-group variation. We ran DAPC with samples grouped by wintering grounds and by nursery grounds (Australian migratory corridor samples were excluded). Data were visualised by plotting samples by linear discriminant coordinates. In contrast, STRUCTURE attempts to group individuals into clusters that minimise deviations from the Hardy-Weinberg equilibrium and linkage disequilibrium. The fit of the data to K populations was assessed in STRUCTURE under the admixture and correlated allele frequency model, with and without prior information on the sampling locations of the data (location prior set as wintering ground). Ten replicates of K = 1 - 5 were conducted, each with burn-ins of one million iterations and runs of ten million Markov chain Monte Carlo (MCMC) iterations, and convergence was assessed by visually inspecting the summary statistics (e.g. F_{ST}). We used *CLUMPAK* to summarise the modes or distinct solutions for each value of *K* (Kopelman et al., 2015) and assessed the most likely value of K using the mean log likelihood from across the ten runs, summarized using STRUCTURE HARVESTER (Earl & VonHoldt, 2012). The relationship between population structure and geographic location was quantified using *ObStruct* (Gayevskiy et al., 2014). We asked whether the phylogeographic pattern originally documented in the mtDNA by Patenaude et al., (2007) was still evident with our larger dataset by estimating a phylogenetic tree using MrBayes v3.2 (Ronquist et al., 2012) and sequences from all three right whale species (E.

australis, E. japonica and E. glacialis). We used MrBayes v3.2 to simultaneously select the best

model of evolution and construct a phylogenetic tree, by sampling across the substitution model space in the Bayesian MCMC analysis itself. The analysis was conducted using the sequences of the unique SRW mtDNA control region haplotypes and previously published sequences from North Atlantic (E. glacialis) and North Pacific (E. japonica) right whales (see Supplementary File 1 for accession numbers and Supplementary File 2 for the nexus file of SRW sequences used). We undertook two runs of MrBayes, each with two chains that were run for one million iterations, using the bowhead whale ($Balaena\ mysticetus$) sequence as an outgroup. We compared the standard deviation of the split frequencies (<0.01) and the potential scale reduction factor (PSRF, should be close to 1) to detect whether convergence had been reached, and ran the program for additional iterations if required. We summarized the tree and branch length information after discarding the first 25% of trees as burn-in and used MrBayes to generate the consensus tree with clade credibility (posterior probability) values. We also constructed a median joining network (Bandelt et al., 1999) for the haplotypes using POPART (Leigh & Bryant, 2015) to examine the relationships and distributions of haplotypes (with $\epsilon = 0$).

Estimating contemporary and long-term gene flow

We used the program *BayesAss v3.0* (Wilson & Rannala, 2003) to co-estimate recent migration rates (past two generations), individual assignment and ancestries, based on microsatellite genotypes. Initial runs were conducted to calibrate the mixing parameters and to ensure that acceptance rates were in the optimal range of 0.2-0.6. During this phase we adjusted the allele-frequency mixing parameter to 0.15 but decided to keep all other parameters at their default values (0.10). We then conducted five *BayesAss* runs of ten million iterations with initial burnins of one million iterations. Parameters were sampled every 1000 iterations and traces were

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visually checked for convergence in *TRACER v1.6* (Rambaut et al., 2014). We report median migration rates with 95% HPD interval from all runs and the mean assignment probabilities of individuals across the five runs.

We tested for sex-biased dispersal with the R package *hierfstat* (Goudet & Jombart, 2015), looking for differences in F_{ST} and the variance of corrected assignment indices (vAIc) between males and females (Goudet et al., 2002). The significance of the difference was tested using null distributions generated with 1,000 permutations (Goudet et al., 2002).

We attempted to estimate long-term gene flow rates using the coalescent based program LAMARC v 2.0, as this program simultaneously estimates migration rates, growth rates and θ (Kuhner, 2006). In principle, this method can account for changes in abundance caused by whaling and also avoid positively biasing diversity estimates by accounting for migration (Kuhner, 2006). We used the Bayesian option for *LAMARC*, and ran two replicates, each comprising one chain with 100,000 sampled genealogies, sampled every 50th genealogy, discarding the first 25,000 samples of each search. To improve search performance of the Bayesian option, we followed the suggestion from the *LAMARC* manual of employing a search strategy with three heated chains (1.0, 1.1 and 1.3 respectively). We conducted one *LAMARC* run using all the microsatellite data, as the LAMARC manual recommends one longer run with heating to obtain the best results for the Bayesian option. Given the large size of the mtDNA dataset, we ran three replicates of LAMARC, each with a different set of 100 individuals (n = 25each for Argentina, South Africa and New Zealand nursery areas and the Australian wintering habitat), randomly subsampled from the larger dataset without replacement. Long-term and whaling era migration rates were also estimated with ABC analyses (see below).

We employed approximate Bayesian computation (ABC) to test competing hypotheses regarding changes in N_e and gene flow from observed microsatellite data. These initial scenarios were generated by previous work characterizing the mtDNA phylogeographic pattern, which has been described as either consistent with continuous gene flow after divergence or isolation followed by secondary contact (Patenuade et al., 2007). The mtDNA data were not included in the ABC analyses as the hypotheses being tested were generated from the mtDNA phylogeny. To test the hypothesis that whaling could have impacted connectivity between wintering grounds, we included scenarios in which rates of gene flow rate change in the whaling era. As a type of null model, we include a scenario with no gene flow after divergence.

A total of six scenarios were examined (Figure 2): continuous gene flow following population divergence at a single migration rate M_H (Scenario 1) or two migration rates: one since divergence M_H and one since the whaling era M_W (Scenario 2); isolation following divergence, with either: no subsequent gene flow (Scenario 3); or gene flow at one migration rate since the whaling era M_W (Scenario 4), or one migration rate since secondary contact, M_C (Scenario 5); or two migration rates: one since secondary contact, M_C , and one since the whaling era, M_W (Scenario 6).

All scenarios incorporated a reduction in N_e due to whaling, followed by a recovery (see Table 2 for prior distributions), and the timing of these events was fixed at nine and two generations before present, respectively. We assumed an average effective generation time of 25 years (Taylor et al., 2007) and a per-generation mutation rate for the microsatellite loci of 5×10^{-4} (Estoup et al., 2002). For each scenario, 100,000 coalescent simulations were run with fastsimcoal2 (Excoffier & Foll, 2011) and 17 summary statistics (Supplementary Table 2), were calculated in arlsumstat v3.5.2 (Excoffier & Lischer, 2010). We analysed the results in an ABC

framework with the R package *abc* (Csilléry et al., 2012), using the neural networks algorithm (Blum & François, 2010) with 1% acceptance ratio to estimate posterior parameters, a method that is suitable for high-dimensional, correlated summary statistics (Csilléry et al., 2012). Model selection was conducted by calculating the posterior model probabilities and Bayes factors (BF) in the *abc* package. BF was calculated for all possible pairs of models and was interpreted following the scale of Jeffreys (1961; see Supplementary Table 3). The ability of the ABC approach to distinguish between the best-selected models was assessed using a cross-validation function in the *abc* package. In addition, we undertook posterior predictive checks using nine summary statistics not employed in the initial ABC analysis. Posterior predictive checks were carried out by sampling 1000 combinations of model parameters from the posterior distributions for scenarios and using these as the input for coalescent simulations in *fastsimcoal2* with the same settings as used for the initial scenarios. For each of the 1000 simulations per scenario, we calculated the 95% confidence intervals and determined whether these encompassed the observed value.

RESULTS

Microsatellite genotyping and diversity statistics

In total, 222 individuals were genotyped at an average of 16.2 of 17 microsatellite loci, with an estimated error rate of 0.7% per allele (for more information see Supplementary Material 1). Microsatellite-based diversity statistics were broadly comparable across ocean basins and nursery areas (Table 1). In contrast, the mitochondrial DNA data (Table 1) showed generally

higher levels of diversity in the South Atlantic than in the Indo-Pacific nursery grounds.

Permutations confirmed that this was a statistically significant difference for both nucleotide and haplotype diversity (at $\alpha = 0.05$).

As the samples comprise overlapping generations, the estimates of N_e based on microsatellite loci actually reflect the number of effective breeders (N_b) that produced the sample. Estimates of N_b were broadly similar across ocean basins and Argentinean and South African nursery grounds (Table 1; Supplementary Table 4). For the two individual Australian nursery grounds and the New Zealand wintering ground, the estimates of N_b had undefined upper boundaries, and for the former this is likely due to the small sample sizes. For the New Zealand wintering ground, we suggest it is due to the increased variance and lower precision found when using the bias-corrected linkage disequilibrium method in populations with large N_e (Waples & Do, 2010). However, the lower bound has been shown to be reliable in such cases (Waples, 2010), so $N_b = 192$ is probably a reasonable lower bound for the New Zealand wintering ground.

Contemporary patterns of genetic diversity

The AMOVA analyses and fixation indices indicated greater variation between ocean basins (AMOVA: $F_{ST} = 0.126$ and $\Phi_{ST} = 0.131$ for mtDNA, $F_{ST} = 0.024$ for microsatellites, all at p<0.01) than among wintering grounds within ocean basins ($F_{ST} = 0.052$ and $\Phi_{ST} = 0.082$ for mtDNA, $F_{ST} = 0.004$ for microsatellites, all at p < 0.01; see Table 3 and Supplementary Table 5 for fixation indices). Direct comparison of the Indo-Pacific and South Atlantic ocean basins yielded estimates of divergence for mtDNA at $F_{ST} = 0.161$ and $\Phi_{ST} = 0.189$ (p<0.001). The divergence for microsatellite loci was $F_{ST} = 0.012$ (95% CI: 0.007, 0.018) and Jost's D = 0.041 (95% CI: 0.024, 0.063, all p<0.001).

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Microsatellite loci data clustered by ocean basins, based on the STRUCTURE analysis particularly with location prior set as nursery or wintering ground (Figure 3A, B). This was supported by log likelihood suggesting the best K = 2, with cluster corresponding to ocean basin, and the *ObStruct* analysis showing there was a strong ($R^2 = 0.95$) and significant (p < 0.0001) correlation between ocean basin and genetic cluster. When no prior population information was provided, the highest likelihood was for K = 1 (Figure 3), which is not surprising given that STRUCTURE has little power to resolve population structure $F_{ST} < 0.02$ (Latch et al., 2006). However, when K = 2, the next best fitting K, was analysed with *ObStruct*, there was a significant correlation (p < 0.0001) between ocean basin and genetic cluster ($R^2 = 0.45$). When grouped by wintering grounds, DAPC separated samples by ocean basins along linear discriminant 1 (LD1) and by wintering grounds within ocean basins along LD2 (Figure 3C). There was overlap between Indo-Pacific wintering grounds, as described previously (Carroll et al., 2015), although when grouped by nursery grounds the distinctiveness of the southwest Australian samples was emphasized (Supplementary Figure 1). As expected, the global mitochondrial phylogenetic tree for the three right whale species shows distinct clades into which North Pacific, North Atlantic and southern hemisphere individuals sort cleanly (Supplementary Figure 2). Convergence was indicated as average standard deviation of split frequencies was 0.005 and the PRSF was 1.00. Within SRWs, the Indo-Pacific sample contains just 13 haplotypes in a total sample of 769 individuals, while the South Atlantic sample contains 55 haplotypes (4.2 times as many) in a total sample of 558 (0.73 times as many). The 13 Indo-Pacific haplotypes are broadly distributed through the SRW genealogy, as are many other common haplotypes (Figure 3D). This is the Type II phylogeographic pattern, described as pronounced phylogenetic gaps between some branches in

a gene tree, with principal lineages showing no obvious geographic pattern (Avise, 2000) as previously described (Patenaude et al., 2007) for SRWs. However, the Indo-Pacific samples could possibly be derived from expansions from a few ancestral sequences, which could imply a small founding population.

Estimates of contemporary and long-term gene flow

We pooled Argentina and South Africa to represent the 'South Atlantic' and pooled New Zealand and Australia to represent the 'Indo-Pacific' for the *BayesAss* analysis, following the nomenclature previously used (Patenaude et al., 2007). Inspection of the traces for all five runs indicated convergence was achieved, with effective sample sizes for all parameters on each run >450. The migration rate estimates were consistent across runs (Supplementary Table 6) and the median migration rate (proportion of individuals that are migrants) from the South Atlantic to the Indo-Pacific was 0.038 (95% HPD 0.006, 0.083) and 0.028 (95% HPD <0.001, 0.068) in the reverse direction (Supplementary Figure 2). *BayesAss* analysis identified two putative first-generation and four second-generation immigrants, though mostly with low confidence (Supplementary Table 7). There was no evidence of sex-biased dispersal between ocean basins, based on either the F_{ST} or vAIc metrics using the microsatellite genotypes (p > 0.05 for all analyses).

The three runs of the coalescent sampler *LAMARC* with different random subsamples of the mtDNA dataset produced similar patterns and so the results were combined for parameter estimation using *TRACER*. While the combined effective sample sizes were sufficient (>500), the traces did not show signs of convergence for all parameters. The *LAMARC* analysis using microsatellite markers also failed to converge, so we do not present the results.

Historical demography of the SRW

The ABC analysis yielded near-zero posterior probabilities (<0.0001) for the demographic scenario with no gene flow (scenario 3) and those with continuous gene flow (scenarios 1 and 2, see Supplementary Table 8). In contrast, the posterior model probability and BF estimates (Table 2) strongly support the scenarios simulating secondary contact between previously isolated populations (scenarios 4 - 6) and we focus on the results of these scenarios in the rest of the paper (posterior distributions can be found in Supplementary File 3).

Of the three models simulating secondary contact, scenarios 5 and 6 showed the best fit to the data based on posterior model support and BF (Table 2B). The support for scenario 5, which had one constant migration rate since secondary contact, was marginally greater than for scenario 6, which had one post-secondary contact and one post-whaling migration rate (Table 2B, Supplementary Figure 5), with a BF = 1.24, which is barely worth mentioning on Jeffreys' (1961) scale for interpreting BF. The cross validation analysis showed that while scenario 4 was distinguishable from scenarios 5 and 6, these two latter scenarios were misclassified as each other >30% of the time. This implies that the ABC method was unable to distinguish between scenarios that differ by events that happened in the recent past (<10 generations) given the available dataset. In light of this, and the fact that the post-contact and post-whaling migration estimates for scenario 6 were very similar to each other and to the post-contact estimate from scenario 5 (all <0.03), we suggest that scenario 5 offers the most parsimonious scenario to explain the observed data (see Supplementary File 3 for graphs of model fit and prior and posterior distributions).

Overall, the evidence is consistent with the hypothesis that secondary contact was stimulated by environmental changes that occurred near the end of the LGM. Obvious candidates

would include sea-level rise, which was dramatic and changed the spatial distribution of suitable wintering habitat (Scourse, 2013), potentially triggering increased dispersal in search of better places to calve and socialize. We were unable to resolve the timing of the initial secondary contact, as the estimates span 11-960 generations ago (corresponding to 275 – 24,000 years, assuming a 25-year effective generation time), which includes the beginning of the present warm interglacial period.

DISCUSSION

This first investigation of global diversity and population structure for SRW using both mtDNA and nuclear DNA markers highlights a complex interplay between forces promoting isolation (e.g., philopatry and migratory fidelity) and forces promoting connectivity (e.g., climate change). The ABC results robustly support a period of historical isolation was followed by secondary contact between ocean basins. As right whales can easily swim thousands of kilometers, both the isolation and its subsequent breakdown were likely the consequences of behavioural mechanisms, presumably the same ones that continue to isolate populations today, as reflected in the heterogeneity of gene-flow estimates across the Southern Hemisphere. The present data and analyses do not provide sufficient resolution to resolve debates about the impact of whaling on SRW genetic diversity, but they do bear on a number of general issues in population management.

Connectivity through time: impacts of natural and anthropogenic changes

The finding of isolation and secondary contact between the Indo-Pacific and South Atlantic

suggests that SRWs in the two ocean basins had limited connectivity until less than 1,000

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generations ago (25,000 years, assuming a 25-year effective generation time). This implies that for thousands of generations after divergence, the populations were relatively isolated, with the most parsimonious explanation being behavioural mechanisms such as female philopatry. discussed in-depth below. The timing of secondary contact supports our hypothesis that the transition from the LGM to the Holocene precipitated the secondary contact, but other, more recent factors cannot be ruled out. The impact of profound climatic change, such as the transition to the Holocene after the LGM, will depend on the ecology of the species. For example, both Antarctic blue (Balaenoptera musculus musculus) and bowhead whales saw decreases in effective population sizes linked to the decline of their sea ice habitat after the LGM (Attard et al., 2015; Phillips et al., 2013). By contrast, the emergence of new breeding habitat and productive foraging areas coincident with deglaciation, reduction in sea ice and increasing productivity during the Holocene after the LGM, led to postglacial population expansion in a number of penguin species and southern elephant seals (Mirounga leonina) in the Southern Hemisphere (Younger et al., 2016). Other species have apparently adapted to new conditions; Pyenson & Lindberg (2011) suggest that the gray whale (*Eschrichtius robustus*) can switch prey, enabling the species to maintain a constant population size across glacial cycles.

The impact of glaciation on connectivity within species is less well studied than its impact on effective population sizes, but its effects would be expected to depend on dispersal ability, population density and ecological necessity or opportunity (Bérubé et al., 1998; Phillips et al., 2011). In the ice-adapted and vagile bowhead whale, heavy sea ice three thousand years ago did not prove a barrier to connectivity across the Holarctic, based on a comparison of historical and contemporary mtDNA data (Alter et al., 2012). In contrast, dispersal rates increased in the Steller sealion (*Eumetopias jubatus*) coincident with an increase in glaciation

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that disrupted breeding habitat (when effective population size was high; Phillips et al., 2011). We suggest that SRWs may have experienced something similar, when their breeding areas were disrupted by sea-level rise following the LGM, prompting increased dispersal between shifting constellations of winter breeding grounds.

Our point estimates of migration from the Indo-Pacific into the South Atlantic from BayesAss are tenfold higher than the estimates of post-contact migration from ABC. Whether this difference reflects a real, very recent increase in migration, or a bias in *BayesAss* estimates. probably linked to the violation of continuous migration assumption of the program, is not clear. A recent increase in migration could plausibly be linked to recovery from whaling. Allendorf et al., (2008) suggested that hunting could either increase directional gene flow into certain populations, for example, into low density areas, or it could decrease migration with concomitant reductions in population size and density. Increase in migration linked to population expansion and recolonisation has been seen in other marine mammals such as the migration from areas of high to low density implicated in the recovery of populations of gray seals (Brasseur et al., 2014). As our estimate of recent gene flow in SRW does not appear to be asymmetric, the change in migration rate could also reflect hunting's impact on behavioural processes across populations. Hunting appears to effect the timing of migratory events in sockeye salmon (Oncorhynchus nerca) (Quinn et al., 2007), as well as characteristics such as boldness (Leclerc et al., 2017) that could influence exploratory behaviour (although this is controversial; Mueller et al., 2014; Rollins et al., 2015). Disruption to breeding behaviour or aggregations could also have precipitate increased dispersal during or subsequent to the whaling era. For example, the disruption of humpback whale breeding aggregations is thought to have influenced metapopulation dynamics (Clapham & Zerbini, 2015). This idea, the social aggregation hypothesis,

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holds that hunting-induced disruption to the humpback whale's social system led to movement of animals from low density to high density breeding aggregations as the species recovered from whaling (Clapham & Zerbini, 2015). Hunting, particularly in the early pre-industrial era, targeted coastal wintering grounds used by aggregations of SRWs as calving, socializing and breeding areas (Smith et al., 2012). Disruption of behaviours at wintering grounds by whaling could have prompted dispersal events in recent generations, just as disruption of wintering grounds by the expansion of shallow seas during the Holocene could have thousands of years prior.

An alternative explanation for a recent increase in migration rates is that the demographic history of the SRWs violates the assumptions of *BayesAss*, in which case the estimates of migration rate from this programme could be biased. Bayes Ass considers a constant population size scenario and implicitly assumes that migration is continuous through time. Linkage or gametic disequilibrium observed in a population is due to recent migration, which increases concomitantly with levels of genetic differentiation amongst populations. However, the demographic history of SRWs, with secondary contact following isolation and the demographic decline due to whaling, likely increased linkage disequilibrium beyond what is expected under continuous migration and constant population sizes. This would lead to an overestimate of migration rates. Simulation studies indicate that if the model assumptions are violated, Bayes Ass still performs well if migration rates are low (<0.01) and F_{ST} is high (>0.10) (Faubet et al., 2007), however, in the present study F_{ST} is fairly low ($F_{ST} = 0.024$). Additional empirical and theoretical work should be directed toward resolving this biologically interesting question, which has potentially large implications for attempts to predict how SRWs might respond to future climate change.

More generally, it is important to consider the limitations of methods used to infer connectivity in non-equilibrium situations. The ABC framework is highly flexible and can explicitly simulate and compare scenarios involving isolation with gene flow and isolation with secondary contact scenarios. Previous suggestions that coalescent-based methods may also be able to differentiate such models based on the timing of migration estimates have proven incorrect (Gaggiotti, 2011), based on both theoretical (Sousa et al., 2011) and simulation studies (Strasburg & Rieseberg, 2011). Such investigations would violate the assumption, common to many coalescent based methods, that immigration rates have been constant throughout the lifespan of the coalescent tree (Kuhner, 2006; Sousa et al., 2011). Violation of this assumption may have contributed to the lack of convergence of *LAMARC* analyses in our study, although other problems have been noted to affect analyses using coalescent-based methods (see Putman & Carbone, 2014).

Migratory fidelity and circumpolar population structure

The present analyses confirm previous findings of hierarchical global population structure in mtDNA haplotype data (Patenaude et al., 2007) using AMOVA and fixation indices and extend these finding to microsatellite markers (Table 3, Supplementary Table 5). In particular, we document low but statistically significant differentiation between the Argentinean and South African nursery grounds at nuclear loci (microsatellite $F_{ST} = 0.001$, Jost's D = 0.004, p < 0.01) (Table 2 and Figure 3), and extends previous findings of stronger differentiation in mtDNA haplotype data (Patenaude et al., 2007).

Reduced connectivity between ocean basins is a common characteristic in cetaceans, which has been attributed to site fidelity, social structure and resource specialization in toothed

whales, and to migratory fidelity in baleen whales (Bowen et al., 2016). Migratory fidelity, like other forms of philopatry or site fidelity, is hypothesized to be adaptive as it increased the chances of finding mates and/or suitable habitat, and may be favoured by natural selection by enhancing the maintenance of co-adapted gene complexes (Greenwood, 1980; Refsnider & Janzen, 2010; Stiebens et al., 2013). In addition to many baleen whale species, such as humpback whales (Baker et al., 2013), natal philopatry is found in all seven sea turtle species (Bowen et al., 2016), many shark (Chapman et al., 2015) and sea bird species (e.g., Milot et al., 2008), showing its ubiquity in migratory marine species.

We did not find evidence of sex-biased dispersal in SRWs, using indirect genetic methods based on the microsatellite genotypes. It might be that in SRWs, different genetic patterns are evident on distinct habitats across a migratory network, reflective of different patterns of gene flow or dispersal. For example, both males and females show long-term fidelity to nursery grounds, based on photo-identification and genotype recapture studies (Carroll et al., 2013; Cooke et al., 2001). Photo-identification studies, tracking individuals, and paternity analyses, showing recent patterns of gene flow, also indicate limited connectivity between calving grounds within ocean basins, based on studies from the Indo-Pacific (Carroll et al., 2012; Pirzl et al., 2009). In contrast, gene flow between individuals that use different calving grounds could occur on shared migratory corridors, as suggested in Australian migratory habitats based on a combination of photo-identification and genetic data (Carroll et al., 2015), or on shared feeding grounds. For example, whales from South African and Argentinean wintering grounds share summer feeding areas, based on photo-identification and stable isotope data (Best et al., 1993, Rowntree et al., 2001). This could facilitate mating between whales from different

wintering grounds on migratory corridors or promote the temporary dispersal of whales between wintering grounds.

Sex biased dispersal is not necessarily common in migratory marine species. In a recent review, Chapman et al., (2015) found residency and site fidelity occurred in both sexes in more than 50 studies in sharks, although many other studies examined only one sex. In turtles, which show strong natal philopatry, genetic connectivity is attributed to the occasional wanderer (Bowen et al., 2016), rather than sex-biased dispersal. Even in a species with a similar life history pattern with strong female philopatry, the humpback whale, there was no evidence that dispersal was sex biased on a global scale, based on a large genetic analysis (Jackson et al., 2014).

Uncertain impact of whaling-era events in ABC analyses

The ABC analyses could not differentiate scenarios with differences in very recent generations, including between those scenarios with changes in gene flow linked to the whaling era (e.g., between scenarios 5 and 6). This is consistent with findings that recent events are not accurately detected by the coalescent based methods due to data-driven and theoretical constraints (Boitard et al., 2016, Wakeley et al., 2016). In the light of this finding, we ran additional analyses and found that, with the current dataset, the method did not have the power to differentiate between the scenarios with and without a whaling-related bottleneck (See R code and Supplementary File 3).

SRWs underwent a centuries-long demographic bottleneck due to whaling, and simulation studies suggest this reduced mtDNA haplotype number and diversity in the species (Jackson et al., 2008). Long-term exploitation of other cetacean species first hunted by pre-

industrial whalers is also correlated with declines in genetic diversity (Alter et al., 2012; Waldick et al., 2002). For example, bowhead whales appear to have lost unique mitochondrial lineages in contemporary populations compared with historical samples, and this was attributed to habitat loss during the Little Ice Age and/or whaling (Alter et al., 2012). Any decline in diversity in nuclear genes like microsatellites is likely to be less severe than declines in mtDNA, given the larger effective population size. Indeed, high levels of nuclear diversity have been found in some great whale species, supporting the hypothesis that they have had large, long-term effective population sizes (e.g. Ruegg et al., 2013). Furthermore, low levels of genetic diversity in pygmy blue whales (*Balaenoptera musculus brevicauda*) were found to be related to a founder event, rather than recent whaling, in an ABC study using both mtDNA and microsatellite data (Attard et al., 2015). These examples highlight the importance of placing recent anthropogenic impacts in the long-term evolutionary context of a species.

The present study provides what appear to be the first estimates of N_b for SRWs, using the bias-corrected linkage disequilibrium method (Waples, 2006), which produced similar results for the South Atlantic (N_b = 365, 95% CI 241, 712) and Indo-Pacific ocean basins (N_b = 331, 95% CI 230, 556). The estimates of N_b for the Australian, Argentinean and South African wintering grounds were broadly similar, in the low hundreds. Morin et al., (2012) estimated N_b for bowhead whale stocks using 22 microsatellite markers and also found estimates in the low hundreds: the small effective population size was attributed to whaling, but might also be due to life history traits similar across these related species. Given that the demographic bottleneck in SRWs happened in recent generations, it is likely that genomic-based methods that estimate effective population sizes through time using linkage disequilibrium methods (e.g., Hollenbeck

et al., 2016) will be needed to determine the extent to which whaling impacted levels of nuclear genetic diversity.

As with all ABC studies, we could only compare a limited number of distinct hypotheses (Beaumont, 2010): in the present case, these were suggested by previous analyses of mtDNA data (Patenaude et al., 2007). Future studies should investigate models that explicitly incorporate sex-biased dispersal and allow migration rates to vary in response to anthropogenic disturbance and other environmental variables such as bathymetry. Given that we could decisively reject the continuous migration scenarios (1-3) in favor of the isolation and secondary contact scenarios (5 and 6) using only 17 polymorphic nuclear markers, future studies using whole-genome data seem likely to have power to estimate many such parameters.

Conservation implications of findings

Effective management of migratory species requires consideration of both the overall migratory network and the different migratory habitats it encompasses. Long term photographic and genetic monitoring programs show that females return regularly to their natal calving grounds across decades (Carroll et al., 2013; Rowntree et al., 2001). As recruitment is dependent upon female reproductive success, the persistence of these calving grounds is likely reliant on these philopatric females (Avise, 2000), and is unlikely to be supplemented by recruitment from other calving grounds (Carroll et al., 2011; Clapham et al., 2008). Therefore, SRW calving grounds seem likely to be substantially demographically independent, which would imply that they qualify as separate sub-populations under the population concepts advocated by Wade and Angliss (1997). On an evolutionary scale, female fidelity to wintering grounds will have

contributed to significant differences in mtDNA haplotype frequencies between wintering grounds, as presented here and in previous studies (Baker et al., 1999; Patenaude et al., 2007).

On a broader scale, vertical transmission of migratory preferences to both feeding and calving or breeding areas suggests an argument for ecological or demographic distinctiveness of the ocean basins, under the framework advocated by Crandall et al., (2000). While not explicitly mentioned by those authors, behavioural variability is arguably as valid as morphological variability when deciding if individuals from different populations are exchangeable. Vertical transmission of socially learned behaviour, such as learned migratory routes, can shape adaptation, and by favouring the conservation of migratory traditions, promote isolation between populations (Brakes & Dall, 2016; Whiten, 2017). Recently it has been suggested that this 'second inheritance system' needs to be integrated into modeling and management of migratory marine species that face challenges from climate change, as migratory conservatism could limit responses to a changing environment (Keith & Bull, 2017). Therefore, we suggest that SRWs in the two ocean basins should be considered distinct population segments. The ABC and genetic analyses (STRUCTURE and F_{ST} results) reject the null hypothesis of historical and contemporary genetic exchangeability, and the well-documented behavioural mechanisms are consistent with the observed levels of genetic differentiation. Our findings also suggest that non-equilibrium scenarios should be considered in future studies of population structure in migratory marine species. ABC techniques will make this feasible, even where low levels of differentiation and complex population histories pose severe challenges for other inference methods.

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ACKNOWLEDGEMENTS

Biopsy collection in Argentina was conducted under permits from Dirección de Fauna y Flora		
Silvestre (DFyFS) and Secretaría de Turismo y Areas Protegidas del Organismo Provincial de		
Turismo (OPT), Chubut Province, using protocols approved by the University of Utah		
Institutional Animal Care and Use Committee under assigned protocol number 05-01003. Biopsy		
sampling was carried out under permits issued to PBB in terms of the South African Sea Fishery		
Act (no. 12 of 1988), issued 22 February 1995, 9 February 1996 and 9 July 1997. Ethics for the		
New Zealand and Australian datasets can be found in the original publication detailing the		
generation of the genetic data (Carroll et al., 2015). Claudia de Silva and Mary-Beth Rew		
contributed to the laboratory work on the South African SRW samples. ELC was supported		
while writing this paper by a EU Horizon 2020 Marie Slodowska Curie Fellowship, project		
BEHAVIOUR-CONNECT, by a Newton Fellowship from the Royal Society of London and		
Bayesian statistical training was supported by National Science Foundation (award DEB-		
1145200). Laboratory analyses conducted by ELC were funded by a small grant from the British		
Ecological Society 5076 / 6118 and Bayesian analysis was supported by training from the		
National Science Foundation under Grant No. DEB-1145200. OEG was supported by the Marine		
Alliance for Science and Technology for Scotland (MASTS) funded by the Scottish Founding		
Council (grant reference HR09011). Genetic data from the South African right whale samples		
were generated by MB and PJP with the support of UC Berkeley, University of Stockholm and		
University of Groningen. Computational Biology analyses were supported by the University of		
St Andrews Bioinformatics Unit which is funded by a Wellcome Trust ISSF award. We thank		
the Editor and three anonymous reviewers for constructive comments that improved the		
manuscript.		

658	CONFLICT OF INTEREST STATEMENT
659	No author has a conflict of interest.
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661	DATA ARCHIVING
662	Supplementary File 1: Data on the mitochondrial DNA sequences compiled and generated for
663	this study, including haplotype codes, GenBank accession numbers and original citation, where
664	applicable.
665	Supplementary File 2: Nexus file of the mtDNA haplotypes.
666	Supplementary File 3: Posterior distributions of ABC simulations
667	CircumpolarSRW_SupportingInfo_Jan18.docx: Supplementary Materials 1-3, Supplementary
668	Tables 1-8 and Supplementary Figures 1-5 can be found in the Supporting Information file.
669	
670	Microsatellite data will be submitted to dryad upon acceptance and bash and R code used in this
671	paper can be found at https://github.com/emmcarr/Circumpolar_analysis.
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674	LITERATURE CITED
675	Alexander, A., Steel, D. J., Hoekzema, K., Mesnick, S., Engelhaupt, D., Kerr, I., Payne, R., &
676	Baker, C. S. (2016). What influences the worldwide genetic structure of sperm whales
677	(Physeter macrocephalus)? Molecular Ecology, 25, 2754–2772.
678	Allen, C. S., Pike, J., & Pudsey, C. J. (2011). Last glacial-interglacial sea-ice cover in the SW
679	Atlantic and its potential role in global deglaciation. Quaternary Science Reviews, 30(19-
680	20), 2446–2458.

681 Allendorf, F. W., England, P. R., Luikart, G., Ritchie, P. A., & Ryman, N. (2008). Genetic 682 effects of harvest on wild animal populations. Trends in Ecology and Evolution, 23, 327– 683 337. 684 Alter, S. E., Rosenbaum, H. C., Postma, L. D., Whitridge, P., Gaines, C., Weber, D., Egan, M. 685 G., Lindsay, M., Amato, G., Dueck, L., Brownell, R. L., Heide-Jørgensen, M. P., Laidre, K. 686 L., Caccone, G., & Hancock, B. L. (2012). Gene flow on ice: The role of sea ice and 687 whaling in shaping holarctic genetic diversity and population differentiation in bowhead 688 whales (Balaena mysticetus). Ecology and Evolution, 2, 2895–2911. 689 Andreotti, S., Von Der Heyden, S., Henriques, R., Rutzen, M., Meÿer, M., Oosthuizen, H., & 690 Matthee, C. A. (2016). New insights into the evolutionary history of white sharks. 691 Carcharodon carcharias. Journal of Biogeography, 43(2), 328–339. 692 Attard, C. R. M., Beheregaray, L. B., Jenner, K. C. S., Gill, P. C., Jenner, M. M., Morrice, M. G., 693 Teske, P. R., & Mo, L. M. (2015). Low genetic diversity in pygmy blue whales is due to 694 climate-induced diversification rather than anthropogenic impacts. Biology Letters, 11, 695 20141037. 696 Avise, J. C. (2000). Phylogeography: the history and formation of species. Cambridge, MA, 697 USA: Harvard University Press. 698 Baker, C. S., Patenaude, N. J., Bannister, J. L., Robins, J., & Kato, H. (1999). Distribution and 699 diversity of mtDNA lineages among southern right whales (Eubalaena australis) from 700 Australia and New Zealand. *Marine Biology*, 134, 1–7. Baker, C. S., Perry, A., Bannister, J. L., Weinrich, M., Abernethy, R., Calambokidis, J., Lien, J., 701 Lamberston, R., Urban-Ramirez, J., Vasquez, O., Clapham, P., Alling, A., O'Brien, S., & 702 703 Palumbi, S. (1993). Abundant mitochondrial DNA variation and world-wide population

- structure in humpback whales. Proceedings of the National Academy of Science, 90, 8239-
- 705 8243.
- Baker, C. S., Steel, D. J., Calambokidis, J., Falcone, E., González-Peral, U., Barlow, J., Burdin,
- A. M., Clapham, P. J., Ford, J. K. B., Gabriele, C. M., Mattila, D., Rojas-Bracho, L.,
- Straley, J. M., Taylor, B. L., Urbán, J., Wade, P. R., Weller, D., Witteveen, B. H., &
- Yamaguchi, M. (2013). Strong maternal fidelity and natal philopatry shape genetic structure
- in North Pacific humpback whales. *Marine Ecology Progress Series*, 494, 291–306.
- 711 Bandelt, H. J., Forster, P., & Röhl, A. (1999). Median-joining networks for inferring intraspecific
- phylogenies. *Molecular Biology and Evolution*, 16, 37–48.
- 713 Bannister, J. L. (1990). Southern right whales off western Australia. Report of the International
- 714 Whaling Commission (Special Issue 12), 279–288.
- 715 Beaumont, M. (2010). Approximate Bayesian computation in evolution and ecology. *Annual*
- 716 Review of Ecology and Systematics, 41, 379–406.
- Paragraphy Benson, S. R., Eguchi, T., Foley, D. G., Forney, K. A., Bailey, H., Hitipeuw, C., Samber, B. P.,
- 718 Tapilatu, R. F., Rei, V., Ramohia, P., Pita, J., & Dutton, P. H. (2011). Large-scale
- movements and high-use areas of western Pacific leatherback turtles, *Dermochelys*
- 720 coriacea. Ecosphere, 2(7).
- 721 Bentley, M. J., Ocofaigh, C., Anderson, J. B., Conway, H., Davies, B., Graham, A. G. C.,
- Hillenbrand, C. D., Hodgson, D. A., Jamieson, S. S. R., Larter, R. D., Mackintosh, A.,
- Smith, J. A., Verleyen, E., Ackert, R. P., Bart, P. J., Berg, S., Brunstein, D., Canals, M.,
- Colhoun, E. A., Crosta, X., Dickens, W. A., Domack, E., Dowdeswell, J. A., Dunbar, R.,
- Ehrmann, W., Evans, J., Favier, V., Fink, D., Fogwill, C. J., Glasser, N. F., Gohl, K.,
- Golledge, N. R., Goodwin, I., Gore, D. B., Greenwood, S. L., Hall, B. L., Hall, K., Hedding,

- D. W., Hein, A. S., Hocking, E. P., Jakobsson, M., Johnson, J. S., Jomelli, V., Jones, R. S.,
- Klages, J. P., Kristoffersen, Y., Kuhn, G., Leventer, A., Licht, K., Lilly, K., Lindow, J.,
- Livingstone, S. J., Mass??, G., McGlone, M. S., McKay, R. M., Melles, M., Miura, H.,
- Mulvaney, R., Nel, W., Nitsche, F. O., O'Brien, P. E., Post, A. L., Roberts, S. J., Saunders,
- 731 K. M., Selkirk, P. M., Simms, A. R., Spiegel, C., Stolldorf, T. D., Sugden, D. E., van der
- Putten, N., van Ommen, T., Verfaillie, D., Vyverman, W., Wagner, B., White, D. A., Witus,
- A. E., & Zwartz, D. (2014). A community-based geological reconstruction of Antarctic Ice
- Sheet deglaciation since the Last Glacial Maximum. *Quaternary Science Reviews*, 100, 1–9.
- 735 Bérubé, M., Aguilar, A., Dendanto, D., Larsen, F., Di Sciara, G. N., Sears, R., Sigurjonsson, J.,
- Urban-R, J., & Palsboll, P. J. (1998). Population genetic structure of North Atlantic,
- 737 Mediterranean Sea and Sea of Cortez fin whales, *Balaenoptera physalus* (Linnaeus 1758):
- analysis of mitochondrial and nuclear loci. *Molecular Ecology*, 7, 585–599.
- 739 Best, P. (1990). Trends in the inshore right whale population off South Africa, 1969-1987.
- 740 *Marine Mammal Science*, 6, 93–108.
- Best, P., Brandão, A., & Butterworth, D. (2001). Demographic parameters of southern right
- whales off South Africa. Journal of Cetacean Research and Management Special Issue, 2,
- 743 161–169.
- Best, P., Payne, R., Rowntree, V. J., Palazzo, J., & Both, M. (1993). Long-range movements of
- South Atlantic right whales *Eubalaena australis*. *Marine Mammal Science*, 9, 227–234.
- Blum, M. G. B., & François, O. (2010). Non-linear regression models for approximate Bayesian
- 747 computation. *Statistics and Computing*, 20, 63–73.
- Bonfil, R., Meyer, M., Scholl, M. C., Johnson, R., O'Brien, S., Oosthuizen, H., Swanson, S.,
- Kotze, D., & Paterson, M. (2005). Transoceanic migration, spatial dynamics, and

- population linkages of white sharks. *Science*, *310*, 100–103.
- Bowen, B. W., Gaither, M. R., Dibattista, J. D., Iacchei, M., Andrews, K. R., Grant, W. S.,
- Toonen, R. J., Briggs, J. C., & Sea, R. (2016). Comparative phylogeography of the ocean
- planet. Proceeding of the National Academy of Sciences USA, 113, 7962–7969.
- Bowen, B. W., Kamezaki, N., Limpus, C. J., Hughes, G. R., Meylan, A. B., & Avise, J. C.
- 755 (1994). Global phylogeography of the loggerhead turtle (*Caretta caretta*) as indicated by
- mitochondrial DNA haplotypes. *Evolution*, 48, 1820–1828.
- 757 Brakes, P., & Dall, S. R. X. (2016). Marine mammal behavior: a review of conservation
- implications. Frontiers in Marine Science, 3, 87.
- Prasseur, S. M. J. M., van Polanen Petel, T. D., Gerrodette, T., Meesters, E. H. W. G., Reijnders,
- P. J. H., & Aarts, G. (2014). Rapid recovery of dutch gray seal colonies fueled by
- immigration. *Marine Mammal Science*, 31, 405–426.
- Carroll, E. L., Baker, C. S., Watson, M., Alderman, R., Bannister, J. L., Gaggiotti, O. E., Gröcke,
- D. R., Patenaude, N. J., & Harcourt, R. (2015). Cultural traditions across a migratory
- network shape the genetic structure of southern right whales around Australia and New
- 765 Zealand. Scientific Reports, 5, 16182.
- 766 Carroll, E. L., Childerhouse, S. J., Christie, M., Lavery, S., Patenaude, N. J., Alexander, A.,
- Constantine, R., Steel, D. J., Boren, L., & Baker, C. S. (2012). Paternity assignment and
- demographic closure in the New Zealand southern right whale. *Molecular Ecology*, 16,
- 769 3960–3973.
- Carroll, E. L., Childerhouse, S. J., Fewster, R., Patenaude, N., Steel, D. J., Dunshea, G.,
- Boren, L., & Baker, C. S. S. (2013). Accounting for female reproductive cycles in a
- superpopulation capture recapture framework. *Ecological Applications*, 23(7), 1677–

1690. 773 774 Carroll, E. L., Fewster, R., Childerhouse, S. J., Patenaude, N. J., Boren, L., & Baker, C. S. 775 (2016). First direct evidence for natal wintering ground fidelity and estimate of juvenile 776 survival in the New Zealand southern right whale Eubalaena australis. PLoS ONE, 11, 777 e0146590. 778 Carroll, E. L., Patenaude, N. J., Alexander, A., Steel, D. J., Harcourt, R., Childerhouse, S. J., 779 Smith, S., Bannister, J. L., Constantine, R., & Baker, C. S. (2011). Population structure and 780 individual movement of southern right whales around New Zealand and Australia. Marine 781 Ecology Progress Series, 432, 257–268. 782 Carroll, E. L., Rayment, W., Alexander, A., Baker, C. S., Patenaude, N. J., Steel, D. J., 783 Constantine, R., Cole, R., Boren, L., & Childerhouse, S. J. (2014). Reestablishment of 784 former wintering grounds by the New Zealand southern right whales. Marine Mammal 785 Science, 30, 206–220. 786 Chapman, D. D., Feldheim, K. A., Papastamatiou, Y. P., & Hueter, R. E. (2015). There and back 787 again: A review of residency and return migrations in sharks, with implications for 788 population structure and management. Annual Review of Ecology, Evolution, and 789 *Systematics*, 7, 547–570. 790 Clapham, P., Aguilar, A., & Hatch, L. T. (2008). Determining spatial and temporal scales for 791 management of cetaceans: lessons from whaling. Marine Mammal Science, 24, 183–201. 792 Clapham, P., & Zerbini, A. (2015). Is social aggregation driving high rates of increase in some 793 Southern Hemisphere humpback whale populations? *Marine Biology*, 162, 625–634. 794 Clark, P. U., Dyke, A., Shakun, J., Carlson, A., Clark, J., Wohlfarth, B., Mitrovica, J., Hostetler, 795 S., & McCabe, A. M. (2009). The last glacial maximum. Science, 325, 710–714.

Genetics Resources, 4, 359–361.

818

796 Cooke, J., Rowntree, V. J., & Payne, R. (2001). Estimates of demographic parameters for 797 southern right whales (Eubalaena australis) observed off Península Valdés, Argentina. 798 Journal of Cetacean Research and Management Special Issue, 2, 125–132. 799 Crandall, K., Bininda-Emonds, O. R. P., Mace, G. M., & Wayne, R. K. (2000). Considering 800 evolutionary processes in conservation biology. Trends in Ecology & Evolution, 15, 290-801 295. 802 Croxall, J. P., Butchart, S. H. M., Lascelle, B., Stattersfield, A. J., Sullivan, B., Symes, A., & 803 Taylor, P. (2012). Seabird conservation status, threats and priority actions: a global 804 assessment. Bird Conservation International, 22, 1–34. 805 Csilléry, K., François, O., & Blum, M. G. B. (2012). abc: An R package for approximate 806 Bayesian computation (ABC). *Methods in Ecology and Evolution*, 3, 475–479. 807 Denis, D., Crosta, X., Schmidt, S., Carson, D. S., Ganeshram, R. S., Renssen, H., Crespin, J., 808 Ther, O., Billy, I., & Giraudeau, J. (2009). Holocene productivity changes off Adélie Land 809 (East Antarctica). Paleoceanography, 24, PA3207. 810 Do, C., Waples, R. S., Peel, D., Macbeth, G. M., Tillett, B. J., & Ovenden, J. R. (2014). 811 NeEstimator v2: re-implementation of software for the estimation of contemporary effective 812 population size from genetic data. *Molecular Ecology Resources*, 14, 209–214. 813 Dulvy, N. K., Fowler, S. L., Musick, J. A., Cavanagh, R. D., Kyne, M., Harrison, L. R., Carlson, 814 J. K., Davidson, L. N. K., & Sonja, V. (2014). Extinction risk and conservation of the 815 world's sharks and rays. eLife, 3, e00590. 816 Earl, D. A., & VonHoldt, B. M. (2012). STRUCTURE HARVESTER: A website and program 817 for visualizing STRUCTURE output and implementing the Evanno method. Conservation

819 Estoup, A., Jarne, P., & Cornuet, J. M. (2002). Homoplasy and mutation model at microsatellite 820 loci and their consequences for population genetics analysis. *Molecular Ecology*, 11, 1591– 821 1604. 822 Excoffier, L., & Foll, M. (2011), fastsimcoal: A continuous-time coalescent simulator of 823 genomic diversity under arbitrarily complex evolutionary scenarios. *Bioinformatics*, 27, 824 1332–1334. 825 Excoffier, L., & Lischer, H. (2010). Arlequin suite ver 3.5: A new series of programs to perform 826 population genetics analyses under Linux and Windows. Molecular Ecology Resources, 10, 827 564-567. 828 Excoffier, L., Smouse, P., & Ouattro, J. (1992). Analysis of molecular variance inferred from 829 metric distances among DNA haplotypes: Application to human DNA restriction data. 830 Genetics, 131, 479-491. 831 Fontaine, M. C., Tolley, K. A., Michaux, J. R., Birkun, A., Ferreira, M., Jauniaux, T., Llavona, 832 A., Oztürk, B., Oztürk, A. A., Ridoux, V., Rogan, E., Sequeira, M., Bouquegneau, J., & 833 Baird, S. J. E. (2010). Genetic and historic evidence for climate-driven population 834 fragmentation in a top cetacean predator: the harbour porpoises in European water. 835 Proceedings of the Royal Society B, 277, 2829–2837. 836 Gaggiotti, O. E. (2011). Making inferences about speciation using sophisticated statistical 837 genetics methods: Look before you leap. *Molecular Ecology*, 20(11), 2229–2232. 838 Gayevskiy, V., Klaere, S., Knight, S., & Goddard, M. R. (2014). ObStruct: A method to 839 objectively analyse factors driving population structure using Bayesian ancestry profiles. 840 *PLoS ONE*, *9*(1). 841 Gersonde, R., Crosta, X., Abelmann, A., & Armand, L. (2005). Sea-surface temperature and sea

842 ice distribution of the Southern Ocean at the EPILOG Last Glacial Maximum - A circum-843 Antarctic view based on siliceous microfossil records. *Ouaternary Science Reviews*, 24, 844 869–896. 845 Goudet, J. (1995), FSTAT (version 1.2): A computer note computer program to calculate F-846 statistics. Journal of Heredity, 86, 485–486. 847 Goudet, J., & Jombart, T. (2015). hierfstat: Estimation and tests of hierarchical F-statistics. R 848 package version 0.044-22. https://CRAN.R-project.org/package=hierfstat. 849 Goudet, J., Perrin, N., & Waser, P. M. (2002). Tests for sex-biased dispersal using bi-parentally 850 inherited genetic markers. *Molecular Ecology*, 11, 1103–1114. 851 Greenwood, P. (1980). Mating systems, philopatry and dispersal in birds and mammals. Animal 852 Behaviour, 28, 1140–1162. 853 Groch, K., Palazzo, J., Flores, P., Ardler, F., & Fabian, M. (2005). Recent rapid increase in the 854 right whale (Eubalaena australis) population off southern Brazil. Latin American Journal of 855 Aquatic Mammals, 4, 41–47. 856 Hoffmann, M., Hilton-Taylor, C., Angulo, A., Bohm, M., Brooks, T. M., Butchart, S. H. M., 857 Carpenter, K. E., Chanson, J., Collen, B., Cox, N. A., Darwall, W. R. T., Dulvy, N. K., 858 Harrison, L. R., Katariya, V., Pollock, C. M., Quader, S., Richman, N. I., Rodrigues, A. S. 859 L., Tognelli, M. F., Vie, J.-C., Aguiar, J. M., Allen, D. J., Allen, G. R., Amori, G., 860 Ananjeva, N. B., Andreone, F., Andrew, P., Ortiz, A. L. A., Baillie, J. E. M., Baldi, R., Bell, 861 B. D., Biju, S. D., Bird, J. P., Black-Decima, P., Blanc, J. J., Bolanos, F., Bolivar-G., W., 862 Burfield, I. J., Burton, J. A., Capper, D. R., Castro, F., Catullo, G., Cavanagh, R. D., 863 Channing, A., Chao, N. L., Chenery, A. M., Chiozza, F., Clausnitzer, V., Collar, N. J., 864 Collett, L. C., Collette, B. B., Fernandez, C. F. C., Craig, M. T., Crosby, M. J.,

- Cumberlidge, N., Cuttelod, A., Derocher, A. E., Diesmos, A. C., Donaldson, J. S.,
- Duckworth, J. W., Dutson, G., Dutta, S. K., Emslie, R. H., Farjon, A., Fowler, S., Freyhof,
- J., Garshelis, D. L., Gerlach, J., Gower, D. J., Grant, T. D., Hammerson, G. A., Harris, R.
- B., Heaney, L. R., Hedges, S. B., Hero, J.-M., Hughes, B., Hussain, S. A., Icochea M., J.,
- Inger, R. F., Ishii, N., Iskandar, D. T., Jenkins, R. K. B., Kaneko, Y., Kottelat, M., Kovacs,
- K. M., Kuzmin, S. L., La Marca, E., Lamoreux, J. F., Lau, M. W. N., Lavilla, E. O., Leus,
- K., Lewison, R. L., Lichtenstein, G., Livingstone, S. R., Lukoschek, V., Mallon, D. P.,
- McGowan, P. J. K., McIvor, A., Moehlman, P. D., Molur, S., Alonso, A. M., Musick, J. A.,
- Nowell, K., Nussbaum, R. A., Olech, W., Orlov, N. L., Papenfuss, T. J., Parra-Olea, G.,
- Perrin, W. F., Polidoro, B. A., Pourkazemi, M., Racey, P. A., Ragle, J. S., Ram, M.,
- Rathbun, G., Reynolds, R. P., Rhodin, A. G. J., Richards, S. J., Rodriguez, L. O., Ron, S.
- R., Rondinini, C., Rylands, A. B., Sadovy de Mitcheson, Y., Sanciangco, J. C., Sanders, K.
- L., Santos-Barrera, G., Schipper, J., Self-Sullivan, C., Shi, Y., Shoemaker, A., Short, F. T.,
- 878 Sillero-Zubiri, C., Silvano, D. L., Smith, K. G., Smith, A. T., Snoeks, J., Stattersfield, A. J.,
- Symes, A. J., Taber, A. B., Talukdar, B. K., Temple, H. J., Timmins, R., Tobias, J. A.,
- Tsytsulina, K., Tweddle, D., Ubeda, C., Valenti, S. V., Paul van Dijk, P., Veiga, L. M.,
- Veloso, A., Wege, D. C., Wilkinson, M., Williamson, E. A., Xie, F., Young, B. E.,
- Akcakaya, H. R., Bennun, L., Blackburn, T. M., Boitani, L., Dublin, H. T., da Fonseca, G.
- A. B., Gascon, C., Lacher, T. E., Mace, G. M., Mainka, S. A., McNeely, J. A., Mittermeier,
- R. A., Reid, G. M., Rodriguez, J. P., Rosenberg, A. A., Samways, M. J., Smart, J., Stein, B.
- A., & Stuart, S. N. (2010). The impact of conservation on the status of the world's
- vertebrates. Science, 330, 1503–1509.
- Hollenbeck, C. M., Portnoy, D. S., & Gold, J. R. (2016). A method for detecting recent changes

910

888 in contemporary effective population size from linkage disequilibrium at linked and 889 unlinked loci. Heredity, 117, 207-216. 890 IUCN. (2017). IUCN red list of threatened species. IUCN 2017. Version 2017-3. 891 http://www.iucnredlist.org. Downloaded on 05 May 2017. 892 IWC. (1986). Right whales: Past and present status. Report of the International Whaling 893 Commission Special Issue, 10, 146–152. 894 IWC. (2001). Report of the workshop on the comprehensive assessment of right whales. Journal 895 of Cetacean Research and Management Special Issue, 2, 1–60. 896 Jackson, J. A., Patenaude, N. J., Carroll, E. L., & Baker, C. S. (2008). How few whales were 897 there after whaling? Inference from contemporary mtDNA diversity. *Molecular Ecology*, 898 *17*, 236–251. 899 Jackson, J. A., Steel, D. J., Beerli, P., Congdon, B. C., Olavarría, C., Leslie, M. S., Pomilla, C., 900 Rosenbaum, H., & Baker, C. S. (2014). Global diversity and oceanic divergence of 901 humpback whales (Megaptera novaeangliae). Proceedings of the Royal Society B: 902 Biological Sciences, 281, 20133222. 903 Jeffreys, H. (1961). The theory of probability (3rd ed.). Oxford. 904 Jobling, M. A. (2012). The impact of recent events on human genetic diversity. *Philosophical* 905 *Transactions of the Royal Society B*, 367, 793–799. 906 Jombart, T., & Ahmed, I. (2011). adegenet 1. 3-1: new tools for the analysis of genome-wide 907 SNP data. Bioinformatics, 27, 3070–3071. 908 Jost, L. (2008). GST and its relatives do not measure differentiation. *Molecular Ecology*, 17, 909 4015–4026.

Keith, S. A., & Bull, J. W. (2017). Animal culture impacts species' capacity to realise climate-

911 driven range shifts. Ecography, 40(2), 296–304. 912 Kopelman, N. M., Mayzel, J., Jakobsson, M., Rosenberg, N. A., & Mayrose, I. (2015). Clumpak: 913 A program for identifying clustering modes and packaging population structure inferences 914 across K. Molecular Ecology Resources, 1179–1191. 915 Kuhner, M. K. (2006). LAMARC 2.0: Maximum likelihood and Bayesian estimation of 916 population parameters. *Bioinformatics*, 22(6), 768–770. 917 Latch, E., Dharmarajan, G., Glaubitz, J., & Rhodes, O. (2006). Relative performance of Bayesian 918 clustering software for inferring population substructure and individual assignment at low 919 levels of population differentiation. Conservation Genetics, 7, 295–302. 920 Leclerc, M., Zedrosser, A., & Pelletier, F. (2017). Harvesting as a potential selective pressure on 921 behavioural traits. *Journal of Applied Ecology*, 2013–2017. 922 Leigh, J. W., & Bryant, D. (2015). Popart: Full-feature software for haplotype network 923 construction. Methods in Ecology and Evolution, 6, 1110–1116. 924 Mate, B., & Best, P. (2011). Coastal, offshore and migratory movements of South African right 925 whales revealed by satellite telemetry. *Marine Mammal Science*, 27, 455–476. 926 Meirmans, P., & van Tienderen, P. (2004). GENOTYPE and GENODIVE: two programs for the 927 analysis of genetic diversity of asexual organisms. *Molecular Ecology Notes*, 4, 792–794. 928 Milot, E., Weimerskirch, H., & Bernatchez, L. (2008). The seabird paradox: Dispersal, genetic 929 structure and population dynamics in a highly mobile, but philopatric albatross species. 930 Molecular Ecology, 17(7), 1658–1673. 931 Morin, P. A., Archer, F. I., Pease, V. L., Hancock-Hanser, B. L., Robertson, K. M., Huebinger, 932 R. M., Martien, K. K., Bickham, J. W., George, J. C., Postma, L. D., & Taylor, B. L. 933 (2012). Empirical comparison of single nucleotide polymorphisms and microsatellites for

934 population and demographic analyses of bowhead whales. *Endangered Species Research*. 935 19(2), 129–147. 936 Mueller, J. C., Edelaar, P., Carrete, M., Serrano, D., Potti, J., Blas, J., Dingemanse, N. J., 937 Kempenaers, B., & Tella, J. L. (2014). Behaviour-related DRD4 polymorphisms in invasive 938 bird populations. *Molecular Ecology*, 23(11), 2876–2885. 939 Munro, K. J., & Burg, T. M. (2017). A review of historical and contemporary processes affecting 940 population genetic structure of Southern Ocean seabirds. Emu Austral Ornithology, 117, 4— 941 18. 942 Palsboll, P. J., Clapham, P. J., Mattila, D. K., Larsen, F., Sears, R., Siegismund, H. R., 943 Sigurionsson, J., Vasquez, O., & Arctander, P. (1995). Distribution of mtDNA haplotypes in North Atlantic humpback whales: The influence of behaviour on population structure. 944 945 Marine Ecology Progress Series, 116, 1–10. 946 Palsbøll, P. J., Peery, M. Z., & Bérubé, M. (2010). Detecting populations in the "ambiguous" 947 zone: kinship-based estimation of population structure and genetic divergence. *Molecular* 948 Ecology Resources, 10f, 797–805. 949 Pastene, L. A., Goto, M., Kanda, N., Zerbini, A. N., Kerem, D., Watanabe, K., Bessho, Y., 950 Hasegawa, M., Nielsen, R., Larsen, F., & Palsbøll, P. J. (2007). Radiation and speciation of 951 pelagic organisms during periods of global warming: The case of the common minke whale, 952 Balaenoptera acutorostrata. Molecular Ecology, 16, 1481–1495. 953 Patenaude, N. J., Baker, C. S., & Gales, N. (1998). Observations of southern right whales on 954 New Zealand's subantarctic wintering grounds. *Marine Mammal Science*, 14, 350–355. 955 Patenaude, N. J., Portway, V., Schaeff, C., Bannister, J. L., Best, P., Payne, R., Rowntree, V. J., 956 Rivarola, M., & Baker, C. S. (2007). Mitochondrial DNA diversity and population structure

957 among southern right whales (Eubalaena australis). Journal of Heredity, 98, 147–157. 958 Phillips, C. D., Gelatt, T. S., Patton, J. C., & Bickham, J. W. (2011). Phylogeography of Steller 959 sea lions: relationships among climate change, effective population size, and genetic 960 diversity. Journal of Mammalogy, 92(5), 1091–1104. 961 Phillips, C. D., Hoffman, J. I., George, J. C., Suydam, R. S., Huebinger, R. M., Patton, J. C., & 962 Bickham, J. (2013). Molecular insights into the historic demography of bowhead whales: 963 Understanding the evolutionary basis of contemporary management practices. *Ecology and* 964 Evolution, 3(1), 18–37. 965 Pichler, F. B., Robineau, D., Goodall, R. N. P., Meÿer, M. A., Olivarría, C., & Baker, C. S. 966 (2001). Origin and radiation of Southern Hemisphere coastal dolphins (genus 967 Cephalorhynchus). Molecular Ecology, 10, 2215–2223. 968 Pinsky, M. L., & Palumbi, S. R. (2014). Meta-analysis reveals lower genetic diversity in 969 overfished populations. *Molecular Ecology*, 23, 29–39. 970 Pirzl, R., Patenaude, N. J., Burnell, S., & Bannister, J. L. (2009). Movements of southern right 971 whales (Eubalaena australis) between Australian and subantarctic New Zealand 972 populations. Marine Mammal Science, 25(2), 455–461. 973 Pritchard, J. K., Stephens, M., & Donnelly, P. (2000). Inference of population structure using 974 multilocus genotype data. Genetics, 155, 945–959. 975 Putman, A. I., & Carbone, I. (2014). Challenges in analysis and interpretation of microsatellite 976 data for population genetic studies. *Ecology and Evolution*, 4, 4399–4428. 977 Pyenson, N. D., & Lindberg, D. R. (2011). What happened to gray whales during the 978 pleistocene? the ecological impact of sea-level change on benthic feeding areas in the north 979 pacific ocean. *PLoS ONE*, 6(7).

980 Ouinn, T. P., Hodgson, S., Flynn, L., Hilborn, R., & Rogers, D. E. (2007). Direction selection by 981 fisheries and the timing of sockeve salmon (Oncorhynchus nerka). Ecological Applications, 982 *17*, 731–739. 983 Rambaut, A., Suchard, M., Xie, D., & Drummond, A. (2014). Tracer v1.6, Available from 984 http://beast.bio.ed.ac.uk/Tracer. 985 Refsnider, J. M., & Janzen, F. J. (2010). Putting eggs in one basket: Ecological and evolutionary 986 hypotheses for variation in oviposition-site choice. Annual Review of Ecology, Evolution, 987 and Systematics, 41, 39-57. 988 Reilly, S. B., Bannister, J. L., Best, P., Brown, M. W., Brownell, R., Butterworth, D., Clapham, 989 P., Cooke, J., Donovan, G. P., Urban, J., & Zerbini, A. N. (2008). Eubalaena australis 990 (Chile-Peru subpopulation). In IUCN 2009. IUCN Red List of Threatened Species. Version 991 2009.2. www.iucn.redlist.org. 992 Richards, R. (2009). Past and present distributions of southern right whales (Eubalaena 993 australis). New Zealand Journal of Zoology, 36, 447–459. 994 Rollins, L. A., Whitehead, M. R., Woolnough, A. P., Sinclair, R., & Sherwin, W. B. (2015). Is 995 there evidence of selection in the dopamine receptor D4 gene in Australian invasive starling 996 populations? Current Zoology, 61(3), 505–519. 997 Ronquist, F., Teslenko, M., Van Der Mark, P., Ayres, D. L., Darling, A., Höhna, S., Larget, B., 998 Liu, L., Suchard, M. A., & Huelsenbeck, J. P. (2012). MrBayes 3.2: Efficient Bayesian 999 phylogenetic inference and model choice across a large model space. Systematic Biology, 1000 *61*, 539–542. 1001 Rowntree, V. J., Payne, R., & Schell, D. (2001). Changing patterns of habitat use by southern 1002 right whales (Eubalaena australis) on their nursery ground at Península Valdés, Argentina,

1003 and in their long-range movements. Journal of Cetacean Research and Management 1004 *Special Issue*, 2, 133–143. 1005 Ruegg, K., Rosenbaum, H. C., Anderson, E. C., Engel, M., Rothschild, A., Baker, C. S., & 1006 Palumbi, S. R. (2013). Long-term population size of the North Atlantic humpback whale 1007 within the context of worldwide population structure. Conservation Genetics, 14, 103–114. 1008 Hoelzel, A. R. (1999). Impact of population bottlenecks on genetic variation and the importance 1009 of life history; a case study of the northern elephant seal. Biological Journal of the Linnean 1010 Society, 68, 23–39. 1011 Scourse, J. (2013). Quaternary- sea-level and palaeotidal changes: A review of impacts on, and 1012 responses of the marine biosphere. In R. N. Hughes, D. Huges, & I. P. Smith (Eds.). Oceanography and Marine Biology: An Annual Review (Volume 51, pp. 1-70). Taylor and 1013 1014 Francis. 1015 Smith, T. D., Reeves, R. R., Josephson, E., & Lund, J. N. (2012). Spatial and seasonal 1016 distribution of American whaling and whales in the age of sail. *PLoS ONE*, 7, e34905. 1017 doi:10.1371/journal.pone.0034905. 1018 Sousa, V. C., Grelaud, A., & Hey, J. (2011). On the nonidentifiability of migration time 1019 estimates in isolation with migration models. *Molecular Ecology*, 20(19), 3956–3962. 1020 Stiebens, V. A., Merino, S. E., Roder, C., Lee, P. L. M., & Eizaguirre, C. (2013). Living on the 1021 edge: how philopatry maintains adaptive potential. *Proceedings of The Royal Society B*, 1022 *280*, 20130305. 1023 Strasburg, J. L., & Rieseberg, L. H. (2011). Interpreting the estimated timing of migration events 1024 between hybridizing species. *Molecular Ecology*, 20, 2353–2366. 1025 Taylor, B. L., Chivers, S. J., Larese, J., & Perrin, W. F. (2007). Generation length and percent

1026 mature estimates for IUCN assessments of cetaceans. Administrative Report LJ-07-01, 24p. 1027 Tormosov, D., Mikhaliev, Y., Best, P., Zemsky, V., Sekiguchi, M., & Brownell, R. (1998). 1028 Soviet catches of Southern right whales Eubalaena australis 1951-1971. Biological 1029 Conservation, 86, 185–197. 1030 Valenzuela, L. O., Sironi, M., Rowntree, V. J., & Seger, J. (2009). Isotopic and genetic evidence 1031 for culturally inherited site fidelity to feeding grounds in southern right whales (Eubalaena 1032 australis). Molecular Ecology, 18(5), 782–791. 1033 Veríssimo, A., Sampaio, Í., McDowell, J. R., Alexandrino, P., Mucientes, G., Queiroz, N., da 1034 Silva, C., Jones, C. S., & Noble, L. R. (2017). World without borders—genetic population 1035 structure of a highly migratory marine predator, the blue shark (*Prionace glauca*). Ecology 1036 and Evolution, 7(13), 4768–4781. 1037 Wade, P., & Angliss, R. (1997). Guidelines for assessing marine mammal stocks: Report of the 1038 GAMMS workshop April 3-5, 1996, Seattle, WA. U.S. Department of Commerce, NOAA 1039 Technical Memoradum NMFS-OPR-12, 93 pp. 1040 Wakeley, J., King, L., & Wilton, P. R. (2016). Effects of the population pedigree on genetic 1041 signatures of historical demographic events. Proceedings of the National Academy of 1042 Sciences of the United States of America, 113, 7994–8001. 1043 Waldick, R. C., Kraus, S. D., Brown, M. W., & White, B. N. (2002). Evaluating the effects of 1044 historic bottleneck events: an assessment of microsatellite variability in the endangered 1045 North Atlantic right whale. *Molecular Ecology*, 11, 2241–2249. 1046 Waples, R. S. (2006). A bias correction for estimates of effective population size based on linkage disequilibrium at unlinked gene loci. Conservation Genetics, 7, 167–184. 1047 1048 Waples, R. S., & Do, C. (2010). Linkage disequilibrium estimates of contemporary Ne using

1049	highly variable genetic markers: A largely untapped resource for applied conservation and
1050	evolution. Evolutionary Applications, 3, 244–262.
1051	Whitehead, H. (1998). Cultural selection and genetic diversity in matrilineal whales. Science,
1052	<i>282</i> , 1708–1711.
1053	Whiten, A. (2017). A second inheritance system: the extension of biology through culture.
1054	Interface Focus, 7(5), 20160142.
1055	Wilson, G. A., & Rannala, B. (2003). Bayesian inference of recent migration rates using
1056	multilocus genotypes. Genetics, 163, 1177–1191.
1057	Younger, J. L., Emmerson, L. M., & Miller, K. J. (2016). The influence of historical climate
1058	changes on Southern Ocean marine predator populations: A comparative analysis. Global
1059	Change Biology, 22(2), 474–493.
1060	

Table 1: Summary of the number of samples and diversity statistics for mitochondrial DNA control region (mtDNA) and microsatellite markers (17 loci) for southern right whale nursery areas in Argentina, (ARG), South Africa (SAF), New Zealand (NZ) and southwest Australia (SWA), southeast Australia (SEA), the overall Australian wintering habitat (AUS-WH) and ocean basins (Indo-Pacific; IP and South Atlantic; SA). For mtDNA, the sample size (n_m), number of haplotypes (n_{hap}), standardised haplotype richness (n_{hap}), haplotype (n_{hap}) and nucleotide (n_{hap}) diversities (with standard deviation, n_{hap}) are shown. For microsatellites, the sample size (n_{hap}), standardized allele richness (n_{hap}) and effective population size (n_{hap}) lowest allele frequency 0.02 and jackknifed confidence intervals CI) are shown.

1	0	6	8
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	mtDNA				microsatellites					
Region	$n_{\rm m}$	n_{hap}	$ns_{hap}\pm SD$	$h\pm \mathrm{SD}$	π±SD (%)	2n	$k\pm SD$	$H_{OB}\pm SD$	$H_{EXP}\pm SD$	N_e
		•	•							(95% CI)
ARG	208	28	8.91 ± 1.24	0.94 ± 0.01	2.27±1.16	92	5.37 ± 1.49	0.72 ± 0.03	0.77 ± 0.03	181 (120, 344)
SAF	350	37	9.03 ± 1.28	0.94 ± 0.01	2.47 ± 1.26	94	5.47 ± 1.38	0.71 ± 0.04	0.76 ± 0.03	239 (142, 656)
SA	558	55	10.04 ± 1.10	0.97 ± 0.01	2.52 ± 1.28	186	5.46 ± 1.45	0.72 ± 0.03	0.76 ± 0.03	365 (241, 712)
SWA	16	5	4.44 ± 0.63	0.71 ± 0.09	1.79 ± 01.00	34	5.16 ± 1.25	0.80 ± 0.04	0.78 ± 0.02	195 (61, ∞)
SEA	12	5	5.71 ± 0.92	0.82 ± 0.07	2.17 ± 1.22	24	5.39 ± 1.37	0.80 ± 0.04	0.78 ± 0.02	$317(49, \infty)$
AUS-WH	77	10	5.32 ± 1.06	0.78 ± 0.03	1.90 ± 0.99	156	5.47 ± 1.35	0.78 ± 0.03	0.78 ± 0.02	286 (169, 792)
NZ	692	11	3.88 ± 0.83	0.69 ± 0.01	1.49 ± 0.79	102	5.41 ± 1.47	0.72 ± 0.03	0.76 ± 0.03	412 (192, ∞)
IP	769	13	4.10 ± 0.90	0.71 ± 0.01	1.60 ± 0.84	258	5.46 ± 1.37	0.76 ± 0.03	0.78 ± 0.03	331(230, 556)
Total	1327	60	7.91 ± 1.47	0.89 ± 0.01	2.21 ± 1.13	442	5.54 ± 1.44	0.74 ± 0.03	0.77 ± 0.03	-

Table 2: Statistics for the approximate Bayesian computation (ABC) analysis of the historical demography of the southern right whale. A. Prior log-uniform distributions used for ABC analysis and posterior values obtained for the South Atlantic (SA) and Indo-Pacific (IP) ocean basins under Scenarios 4 - 6. Posterior mode and 95% HPDs are reported for migration rates (M_C : constant migration rate since secondary contact; M_W : migration rate since whaling era), effective population sizes (N_e), and time in generations since secondary contact (CONTACT) and population divergence (DIVERGENCE). Effective population sizes are shown schematically in Figure 2 and are divided into historical or pre-whaling (HIST N_e), bottleneck (BOT N_e) and recovery (REC N_e). B. Posterior support, as shown by the posterior model probability, Bayes factor (relative to Scenario 5) and number of posterior predictive summary statistics that encompassed observed value.

A.	Prior	Scenario 4	Scenario 5	Scenario 6
Migration rate since secondary				
contact (M_C)				
M_C SA to IP	0.001-0.50	-	0.0281 (0.0087, 0.2068)	0.0114 (0.0023, 0.4259)
M_C IP to SA	0.001-0.50	-	0.0008 (0.0002, 0.0825)	0.0128 (0.0015, 0.4349)
Migration rate since whaling era				
(M_W)				
M_W SA to IP	0.001-0.50	0.0164 (0.0034, 0.0717)	-	0.0025 (0.0002, 0.0830)
M_W IP to SA	0.001-0.50	0.0235 (0.0052, 0.2091)	-	0.0025 (0.0013, 0.0718)
Effective population sizes (N_e)				
$HIST N_e SA$	1,000-100,000	3,874 (1,057, 80, 909)	3,916 (1,131, 85,781)	3,188 (920, 65,783)
$HIST N_e IP$	1,000-100,000	4,635 (1,275, 92,942)	3,880 (1,131, 82,748)	3,908 (973, 88,744)
BOT N_e SA	20-2,000	128 (34, 2,447)	90 (23, 2,216)	74 (21, 1,430)
BOT N_e IP	20-2,000	88 (22, 1,936)	88 (23, 1,743)	111 (29, 1,998)
$REC N_e SA$	1,000-20,000	1,358 (466, 9,433)	888 (344, 8,126)	2,494 (706, 15,728)
$\operatorname{REC} N_e \operatorname{IP}$	100-20,000	4,447 (1,179, 24,463)	2008 (814, 10,436)	2,013 (660, 12,851)
Timing (generations)				
CONTACT	10-100,000	Fixed	38 (12, 962)	74 (20, 2,187)
DIVERGENCE	2,000-200,000	189,621 (122,048, 200,959)	194,697 (103,040, 209,241)	198,944 (109,871,
				211,017)
В.		Scenario 4	Scenario 5	Scenario 6
Posterior model probability		0.0048	0.5614	0.4338
Bayes factor		117.14	1.00	1.29
Posterior predictive checks		6/9	7/9	7/9

Table 3: Genetic differentiation indices between southern right whale southern right whale nursery areas in Argentina, (ARG), South Africa (SAF), New Zealand (NZ) and southwest Australia (SWA), southeast Australia (SEA), and the overall Australian wintering habitat (AUS-WH). Pairwise F_{ST} based on mtDNA haplotype frequencies (bottom left quadrant) and microsatellite loci (top right quadrant). The 95% confidence intervals for microsatellite-based F_{ST} , shown in parentheses, was estimated using bootstrapping over loci (999 permutations).

	ARG-WG	SAF	SWA	SEA	AUS-WH	NZ		
ARG		0.001**	0.021***	0.022***	0.013*** (0.008,	0.012***		
		(0.000, 0.003)	(0.013, 0.031)	(0.007, 0.021)	0.019)	(0.006, 0.019)		
SAF	0.048***		0.023***	0.017**	0.014*** (0.008,	0.013***		
			(0.012, 0.034)	(0.005, 0.031)	0.020)	(0.007, 0.019)		
SWA	0.143***	0.136***		0.013*	-	0.016**		
				(0.000, 0.028)		(0.001, 0.035)		
SEA	0.104***	0.098**	0.098*		-	0.000		
		0.44.7111				(0.000, 0.003)		
AUS-WH	0.123***	0.115***	-	-		0.004**		
N	0.1004444	O 1 O O ale ale ale	O 1 4 O skalada	0.000	0.072444	(0.000, 0.010)		
NZ	0.190***	0.189***	0.142***	0.000	0.073***	-		
*p<0.05; **p<0.01, ***p<0.001								

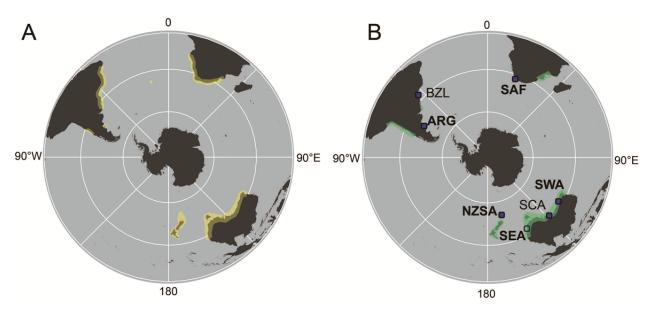


Figure 1: Map of A. historical and B. contemporary southern right whale winter habitat. The contemporary distribution is divided into large wintering aggregations (blue circles) and the areas with sporadic sightings (green shading: see Supplementary Material 2 for references). Samples included in this study are from nursery grounds with bold acronyms: ARG (Argentina), SAF (South Africa), SWA (southwest Australia), and NZSA (New Zealand sub-Antarctic), but not from the BZL (Brazilian) and SCA (southcentral Australia) nursery grounds, also marked on the map. Also included are samples from the SEA nursery ground (southeast Australia, marked by black circle) and mainland New Zealand wintering habitat, which are part of the sporadic sightings.

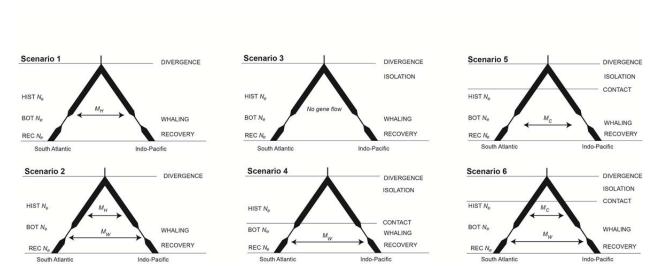


Figure 2: Scenarios compared using the approximate Bayesian computation framework. The six scenarios include continuous gene flow following population divergence with migration varying by scenario; Scenario 1: single migration rate M_H ; Scenario 2: two migration rates: one since divergence M_H and one since the whaling era M_W ; or isolation following divergence, with either: Scenario 3: no subsequent gene flow; Scenario 4: gene flow at one migration rate since the whaling era M_W ; Scenario 5: one migration rate since secondary contact, M_C ; or Scenario 6: two migration rates: one since secondary contact, M_C , and one since the whaling era, M_W (Scenario 6). In all scenarios, the populations diverge at time DIVERGENCE and maintain one HIST N_e until whaling, when each population declines to bottleneck population size BOT N_e during the whaling era and subsequently recover to REC N_e population size.

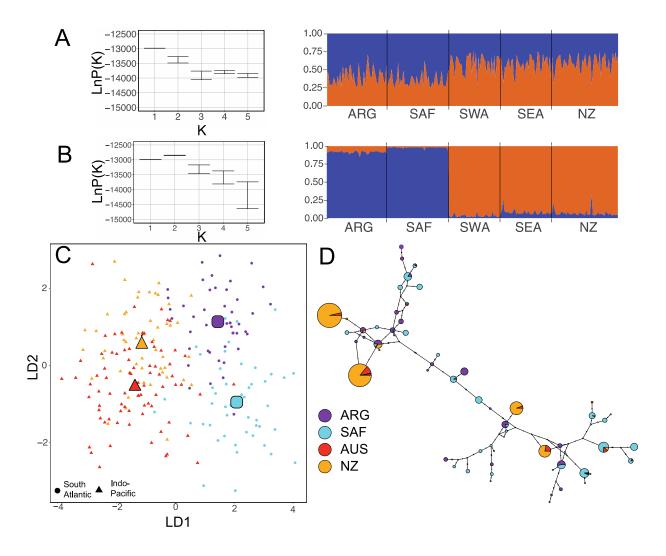


Figure 3: Inference of population structure of southern right whale wintering habitats. Samples collected in Argentina (ARG) and South Africa (SAF) are pooled to form the South Atlantic dataset; samples from Australia (AUS) and New Zealand (NZ) are pooled to form the Indo-Pacific dataset. **A and B:** *STRUCUTRE* results (left); mean log likelihood (LnP(K)), for K = 1 – 5, and (right) the proportion of each individual's genome that assigns to each cluster when K = 2 for (A) standard admixture setting and (B) location prior implemented **C**. Individuals plotted by linear discriminants (LD) from DAPC conducted with samples grouped by wintering ground. The large symbols show the centroid of each wintering ground. **D**. Median joining haplotype

- network of mtDNA haplotypes. Haplotypes are coloured by wintering ground they were sampled
- in, using key shown. Inferred, unsampled haplotypes are shown by small black circles.

