

# Cellular Oncology

## Epigenetic sampling effects: nephrectomy modifies the clear cell renal cell cancer methylome --Manuscript Draft--

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<b>Abstract:</b>	<p>Currently, it is unclear to what extent sampling procedures affect the epigenome. Here, this phenomenon was evaluated by studying the impact of artery ligation on DNA methylation in clear cell renal cell cancer. To this end, DNA methylation profiles between vascularised tumour biopsy samples and devascularized nephrectomy samples from two individuals were compared. The relevance of significantly altered methylation profiles was validated in an independent clinical trial cohort. We found that six genes were differentially methylated in the test samples, of which four were linked to ischaemia or hypoxia (REXO1L1, TLR4, hsa-mir-1299, ANKRD2). Three of these genes were also found to be significantly differentially methylated in the validation cohort, indicating that the observed effects are genuine. Based on these results, we conclude that the impact of sampling procedures in clinical epigenetic studies should</p>	

	be considered, particularly after inducing hypoxia/ischemia, which occurs in many oncological surgery procedures through which tissues are harvested for translational research.
<b>Response to Reviewers:</b>	Dear editor,  my sincerest thanks for your help in making this a better manuscript

The Editorial board of Cellular Oncology

Ghent, September 1, 2016

Dear Dr. Editor,

Please find enclosed our manuscript entitled “*Epigenetic sampling effects: nephrectomy modifies the clear cell renal cell cancer methylome*” for consideration for publication in *Cellular Oncology* as a short communication.

In a recently accepted paper (*Dynamic epigenetic changes to VHL occur with sunitinib in metastatic clear cell renal cancer*, Stewart et al, *Oncotarget*), we demonstrated an epigenetic change in the VHL gene upon clear cell renal cancer treatment. As pre- and post treatment conditions differed in the manner of sampling, we ran an additional study to analyse a possible sampling effect (biopsy vs. nephrectomy). Though this was not the case for VHL – subject of the previous paper - a clear sampling effect was observed for some loci and also appeared to be relevant in the actual treatment study. This implies that sampling procedure impacts results in cellular oncology studies. As the most likely underlying cause – hypoxia – is relevant in many such studies, and the sampling study is unique in its kind (biopsy immediately followed by nephrectomy), we are confident that this paper is of general interest to researchers in the field of cellular oncology.

Yours sincerely,



Prof. Dr. Tim De Meyer

Dept. of Mathematical Modelling, Statistics and Bioinformatics

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1 [Short Communication](#)

2

3 **Epigenetic sampling effects: nephrectomy modifies the clear cell**  
4 **renal cell cancer methylome**

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6

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37

38 **Abstract**

39

40 **Currently**, it is unclear to **what** extent sampling procedures affect the epigenome. Here, this  
41 **phenomenon** was evaluated by studying the impact of artery ligation on DNA methylation in  
42 clear cell renal cell cancer. **To\_t-his end**, DNA methylation profiles between vascularised  
43 tumour biopsy samples and devascularized nephrectomy samples from two individuals were  
44 compared. The relevance of significantly **altered methylation profiles** was validated in **an**  
45 **independent** clinical trial cohort. **We found that six genes were** differentially methylated **in**  
46 **the test samples**, of which four were linked to ischaemia or hypoxia (*REXO1L1*, *TLR4*, *hsa-mir-*  
47 *1299*, *ANKRD2*). Three **of these** genes were also **found to be** significantly differentially  
48 methylated in the validation cohort, indicating that the observed effects are **genuine**. **Based**  
49 **on these results, we conclude that** the impact of **sampling procedures** in clinical epigenetic  
50 studies should be considered, particularly **after** inducing hypoxia/ischemia, which occurs in  
51 many oncological surgery procedures **through which** tissues **are** harvested for translational  
52 research.

53

54 **Keywords:** Hypoxia; Cancer epigenetics; DNA methylation; Sampling effects

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58 **1 Introduction**

59

60 The use of clinical samples in epigenetics research has become routine ~~(Ferraro, 2016; Taucher~~  
61 ~~et al., 2016; Vitiello et al., 2015)[1–3][refs. Cell. Oncol. 39, 295–318, 2016; Cell. Oncol. 39, 195–~~  
62 ~~210, 2016; Cell. Oncol. 38, 17–28, 2015]~~. However, the nature of the sampling procedures may  
63 significantly compromise the resulting epigenetic profiles, leading to an “epigenetic observer  
64 effect”. In renal cell carcinoma (RCC), dynamic molecular changes occur over time and with  
65 therapy, which require serial tissue samples for elucidation [4,5][1, 2]. Despite the  
66 development of standard operating procedures for tissue acquisition and biobanking, less  
67 attention is paid to ensuring constant, robust pre-collection conditions, such as warm  
68 ischaemia time, than to post-collection handling and processing procedures, thus failing to  
69 avoid variation due to pre-analytical factors [6][3]. There are recommended tissue sampling  
70 guidelines from organisations such as the Confederation of Cancer Biobanks, which advise  
71 that warm ischaemia time should be minimised as much as possible prior to freezing of fresh  
72 tissue samples, but this is very difficult to measure and standardise [7][4]. Several studies have  
73 looked at the procurement conditions on RNA biomarker expression and the effect on  
74 previously identified cancer biomarkers [8][5], but so far there have been no such studies on  
75 DNA methylation.

76 Prolonged ischaemia leads to higher levels of tissue hypoxia, which has been shown to  
77 induce DNA demethylation in e.g. hepatoma cells [9][6]. This demethylation has been shown  
78 to be induced by methionine adenosyltransferase 2A (MAT2A), of which the expression is  
79 positively regulated by HIF-1α [9][6], which in turn is negatively regulated by VHL [10][7]. VHL  
80 plays a crucial role in RCC development [11][8]. Hypoxia, methylation and RCC form, therefore,  
81 an intricate network of which the components cannot be studied separately. Yet, different  
82 RCC sampling procedures might obfuscate the results by influencing the hypoxic conditions.  
83 More than most tumours, RCC sampling is challenging as the majority of the extirpative  
84 procedures deal with minimally invasive approaches. It is well established that changes occur  
85 in mRNA levels with increasing time after renal artery clamping in RCC [12][9]. Here, we  
86 assessed the effect of renal artery clamping on the RCC methylome.

87

88 **Patient samples and methods**

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90 Patient-matched sample sets were obtained from two patients who, at the time of open  
91 cytoreductive nephrectomy for metastatic clear cell RCC (ccRCC), had fresh ccRCC tumour  
92 biopsies taken prior to ligation of the renal artery after which matched fresh frozen tumour  
93 samples were harvested following ligation and division of the renal artery and removal of the  
94 kidney as per a previously described approach [5][2]. A total of 12 samples was collected, i.e.,  
95 3 biopsy and 3 nephrectomy samples from each patient. These samples were obtained as part  
96 of the Scottish Collaboration On Translational Research into Renal Cell Cancer (SCOTRRCC)  
97 study (East of Scotland Research Ethics Service REC 1: 10/S1402/33). For validation purposes,  
98 but also to evaluate the potential impact on clinical epigenetics research, matched tumour  
99 samples taken at the time of diagnostic renal tumour biopsy and subsequent nephrectomy  
100 were obtained from 14 patients with metastatic ccRCC. Following primary tumour biopsy,  
101 these patients were treated with three cycles of sunitinib (18 weeks) followed by  
102 cytoreductive nephrectomy after 2 weeks of sunitinib as part of the Upfront Sunitinib  
103 (SU011248) Therapy Followed by Surgery in Patients with Metastatic Renal Cancer: a Pilot  
104 Phase II Study (SuMR; ClinicalTrials.gov identifier: NCT01024205) [13][10]. All samples used  
105 in this study are listed in Supplementary file 1.

106 Extraction of genomic DNA was performed using a Qiagen DNeasy Blood and Tissue  
107 (Qiagen, UK) kit following the manufacturer's instructions. DNA methylation analyses using  
108 MBD sequencing was carried out as described previously [14][11]. A MethylCap kit  
109 (Diagenode, Belgium) was used for capturing methylated fragments from 500 ng starting  
110 material. Massively parallel sequencing of these fragments was subsequently performed on  
111 an Illumina HiSeq 2000 machine (Illumina, San Diego, CA, USA).

112 Raw data files were mapped using BOWTIE to the human reference genome  
113 Hg19/GRCh37, and summarized using an in-house developed Map of the Human Methylome  
114 [15][12] consisting of a putative genome-wide overview of potentially methylated loci  
115 ("methylation cores"). Further data analyses were performed using Python 3.4.3 and R 3.2.1.  
116 The Bioconductor quantro software package (1.2) was used to assess quantile normalization  
117 assumptions [16][13], whereas the limma software package (3.24.15) was used to identify  
118 regions featured by differential methylation patterns. Prior to limma analysis, the samples  
119 were quantile normalized and transformed using the voom function [17][14]. Only  
120 methylation cores that referred to annotated promoter regions (including exon1), and that  
121 had at least an average coverage of one mapped fragment per sample, were withheld for



122 analysis. Low coverage loci are featured by a too low power to be detected as differentially  
123 methylated, thereby unnecessarily inflating the amount of hypotheses tested, which justifies  
124 their unsupervised removal from the dataset. Regions with a False Discovery Rate (FDR) < 0.05  
125 were selected for further assessment using the sample dataset [13][19]. For the validation  
126 dataset, the FDR estimation (Benjamini-Hochberg) was based on the amount of loci to be  
127 validated.

128

## 129 Results and discussion

130

131 A quantro test was performed to check the suitability of the samples for quantile  
132 normalization, which was used for limma-voom. A  $p$ -value of 0.53 was obtained after 1000  
133 permutations, implying that there were no global differences in the distributions between the  
134 non-*ischaemic* biopsy and *ischaemic* nephrectomy samples. A limma-voom data analysis was  
135 subsequently performed using quantile normalization. In Table 1 seven regions are listed that  
136 were found to be differentially methylated up to an FDR of 0.05 (Fig 1 ). Only two of these  
137 seven (*AC232323.1* and *ANKRD2*) exhibited a relative hypomethylation in the *ischaemic*  
138 nephrectomy samples compared to the non-*ischaemic* samples. In one of these (*AC232323.1*)  
139 a different region (8325757) exhibited a significant relative hypermethylation.

140 The differentially methylated regions with a FDR < 0.05 were subsequently subjected  
141 to validation using methylation data obtained from matched biopsy and nephrectomy  
142 samples from the SuMR clinical trial. Three of the seven regions identified in the test set, were  
143 again significantly altered at the same level within the validation cohort (*AC232323.1* region  
144 8325757, *REXO1L1* and *OR6Q1*). A binomial test – using the FDR threshold as expected  
145 probability – rejected the null hypothesis that this would have occurred randomly ( $p = 0.004$ ).  
146 We therefore hypothesise that the shared results between the two studies are caused by  
147 *ischaemic* conditions. Of the 7 loci identified in the test set, 6 were found to have the same  
148 fold change direction in the validation cohort. However, assuming a 50% random chance of  
149 having the same fold change direction, the number of similar direction of change was not  
150 found to be significant ( $p = 0.125$ ). A lower FDR cut-off of 10% yielded 36 significant regions,  
151 yet this cut-off was deemed not sufficiently conservative as the fraction that could be  
152 validated in the validation cohort was too low to reject the null hypothesis of significant  
153 overlap ( $p = 0.085$ , data not shown).

154 Methylation meta-analyses have shown that DNA methylation is a critical event in  
155 tumorigenesis [18][15]. It is, therefore, surprising that an analysis of the effect of tissue  
156 procurement on DNA methylation has so far not been performed, as it has in other molecular  
157 analyses. From our test and validation results it is clear that ischaemic conditions, induced as  
158 part of the surgical procedure, may lead to differential methylation.

159 The results of the study presented here indicate that the RCC methylome may be  
160 modulated following renal artery ligation. Global gene demethylation in samples was not  
161 observed, but consistent demethylation of at least one individual gene (*ANKRD2*) was found.  
162 These results hold considerable significance for translational methylation research for solid  
163 tumours obtained by extirpative surgery, especially where minimally invasive surgical  
164 approaches are used. The most significantly affected gene, *AC232323.1*, encodes a long non-  
165 coding RNA (lncRNA) product. According to LNCipedia [19][16] this lncRNA is linked to the  
166 second most significantly affected gene, *REXO1L1*, and has potential direct biological  
167 relevance (<http://www.lncipedia.org/db/transcript/lnc-REXO1L2P-1:1>). *REXO1L1* was found  
168 to be hypermethylated following renal artery clamping. *REXO1L1* deletions have been linked  
169 to increased apoptosis under certain conditions [20][17], such as intense hypoxia, which may  
170 underlie the results presented above [21][18]. The third significantly affected gene, *TLR4*,  
171 plays a crucial role in kidney ischemia/reperfusion injury [22][19], and *Hsa-mir-1299* also has  
172 a possible role in apoptosis through interaction with *PIM1* [23][20]. The *OR6Q1* gene encodes  
173 an olfactory receptor and is, therefore, an unlikely candidate, although it should be noted that  
174 there is evidence for a limited expression in other tissues as well [24][21]. Finally, *ANKRD2*  
175 belongs to the conserved muscle ankyrin repeat protein (MARP) family. Expression of MARPs  
176 has been found to be induced in response to physiologic stress, injury, hypoxia and  
177 hypertrophy [25,26][22, 23] and the *ANKRD2* mRNA level has indeed been found to be  
178 upregulated under specific hypoxic conditions. Our results indicate that this upregulation may  
179 be brought about by demethylation of a specific methylation core.

180 Despite the test and validation cohorts used, this study should be considered as proof  
181 of principle with a necessarily low power, as it was deemed unethical to expose subjects to an  
182 extra pre-operative percutaneous biopsy procedure without clinical benefit. Also, in the  
183 validation cohort, patients were treated with sunitinib, the time between biopsy and  
184 nephrectomy was longer (20 weeks) and the sequencing depths were generally more shallow,  
185 implying that additional discrepancies between both datasets may have been caused by

186 treatment, temporal or other technical reasons.

187 Taken together, [our data indicate that in both](#) a test and [a validation cohort](#) renal  
188 artery ligation modulates gene methylation in a biologically relevant fashion. As with any  
189 research using surgically resected clinical samples, future methylation studies must be  
190 designed to include robust and well-documented sample procurement techniques [in order](#) to  
191 take these [findings](#) into account.

192

#### 193 **Author's contributions**

194 Conception and design: DJH, AL, GDS, TDM, TP. Development of methodology: CVN, GDS, DJH,  
195 TP, TDM. Acquisition of data (acquired and managed patients, provided facilities, etc.): GDS,  
196 FOM, AL, DJH, TP. Analysis and interpretation of data (e.g., statistical analysis, biostatistics,  
197 computational analysis): CVN, TDM, GDS. Writing, review, and/or revision of the manuscript:  
198 CVN, AL, FOM, WVC, DD, FVN, TP, DJH, GDS, TDM. Study supervision: GDS, DJH, TDM, WVC.

199

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205 educational grant from Pfizer (TP). CVN and TDM were funded by Ghent University  
206 Multidisciplinary Research Partnership 'Bioinformatics: from nucleotides to networks'.

207

#### 208 **Compliance with ethical standards**

209

210 **Conflict of Interest:** The authors declare that they have no conflict of interest.

211

212 **Informed consent:** Informed consent was obtained from all [participants](#) included in the study.

213

214

215 **Legend tot the Figure**

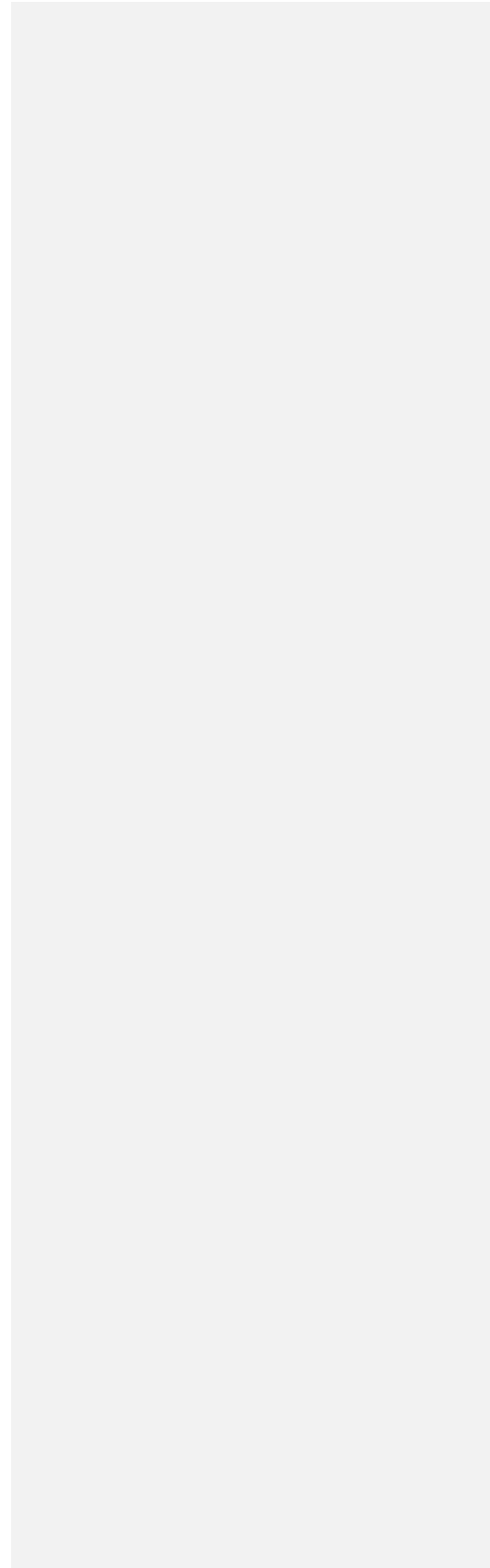
216

217 **Figure 1** Sampling procedure differences. For each of the regions with  $\alpha$  FDR < 0.05, boxplots  
218 of the methylation counts are shown. The methylation core region is shown under the gene  
219 label.

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## References

1. Ferraro, Altered, Primary chromatin structures and their implications in cancer development. *Cell. Oncol.* **39**, 195 (2016) doi:10.1007/s13402-016-0276-6

2. Taucher, V., Mangge, H., Haybaeck, J., Non-coding RNAs in pancreatic cancer: challenges and opportunities for clinical application. *Cell. Oncol.* **39**, 295 (2016) doi: 10.1007/s13402-016-0275-7

3. Vitiello, M., Tuccoli, A., Polisenio, L., Long non-coding RNAs in cancer: implications for personalized therapy. *Cell. Oncol.* **38**, 17 (2015) doi: 10.1007/s13402-014-0180-x

4. Sharpe, K., Stewart, G.D., Mackay, A., Van Neste, C., Rofe, C., Berney, D., Kayani, I., Bex, A., Wan, E., O'Mahony, F.C., O'Donnell, M., Chowdhury, S., Doshi, R., Ho-Yen, C., Gerlinger, M., Baker, D., Smith, N., Davies, B., Sahdev, A., Boleti, E., Meyer, T.D., Crieckinge, W.V., Beltran, L., Lu, Y.-J., Harrison, D.J., Reynolds, A.R., Powles, T., The effect of VEGF-targeted therapy on biomarker expression in sequential tissue from patients with metastatic clear cell renal cancer. *Clin. Cancer Res.* **19**, 6924-6934 (2013) doi:10.1158/1078-0432.ccr-13-1631

5. Stewart, G.D., O'Mahony, F.C., Laird, A., Eory, L., Lubbock, A.L.R., Mackay, A., Nanda, J., O'Donnell, M., Mullen, P., McNeill, S.A., Riddick, A.C., Berney, D., Bex, A., Aitchison, M., Overton, I.M., Harrison, D.J., Powles, T., Sunitinib treatment exacerbates intratumoral heterogeneity in metastatic renal cancer. *Clin. Cancer Res.* **21**, 4212-4223 (2015) doi:10.1158/1078-0432.ccr-15-0207

6. Stewart, G.D., O'Mahony, F.C., Powles, T., Riddick, A.C.P., Harrison, D.J., Faratian, D., What can molecular pathology contribute to the management of renal cell carcinoma? *Nature Rev. Urol.* **8**, 255-265 (2011) doi:10.1038/nrurol.2011.43

7. Confederation of Cancer Biobanks: Biobank Quality Standard - Collecting, Storing and Providing Human Biological Material and Data for Research (2014) <http://ccb.ncri.org.uk/wp-content/uploads/2014/03/Biobank-quality-standard-Version-1.pdf>

8. Freidin, M.B., Bhudia, N., Lim, E., Nicholson, A.G., Cookson, W.O., Moffatt, M.F., Impact of collection and storage of lung tumor tissue on whole genome expression profiling. *J. Mol. Diagn.* **14**, 140-148 (2012) doi:10.1016/j.jmoldx.2011.11.002

9. Liu, Q., Liu, L., Zhao, Y., Zhang, J., Wang, D., Chen, J., He, Y., Wu, J., Zhang, Z., Liu, Z., Hypoxia induces genomic DNA demethylation through the activation of HIF1 $\alpha$  and transcriptional upregulation of ~~mat2a~~ *MAT2A* in hepatoma cells. *Mol. Cancer Ther.* **10**, 1113 (2011) ~~Mol. Cancer Ther.~~ **10**, 1113-1123 (2011) doi:10.1158/1535-7163.mct-10-1010 cannot find this reference, please check

10. Kaelin Jr, W.G., The von Hippel-Lindau tumour suppressor protein: O<sub>2</sub> sensing and cancer. *Nat. Rev. Cancer* **8**, 865-873 (2008) doi:10.1038/nrc2502 cannot find this reference, please check

11. Motzer, R., Michaelson, M., Redman, B., Hudes, G., Wilding, G., Figlin, R., Ginsberg, M.,

Commented [A1]: I have double checked, but I do not find a specific mistake. Pubmed entries are respectively <https://www.ncbi.nlm.nih.gov/pubmed/?term=21460102> and <https://www.ncbi.nlm.nih.gov/pubmed/?term=18923434>

270 Kim, S., Baum, C., DePrimo, S., Li, J., Bello, C., Theuer, C., George, D., Rini, B., Activity of  
271 SU11248, a multitargeted inhibitor of vascular endothelial growth factor receptor and  
272 platelet-derived growth factor receptor, in patients with metastatic renal cell carcinoma. *J.*  
273 *Clin. Oncol.* **24**, 16-24 (2006) doi:10.1200/JCO.2005.02257  
274  
β75 129. Liu, N.W., Sanford, T., Srinivasan, R., Liu, J.L., Khurana, K., Aprelikova, O., Valero, V.,  
276 Bechert, C., Worrell, R., Pinto, P.A., Yang, Y., Merino, M., Linehan, W.M., Bratslavsky, G. Impact  
277 of ischemia and procurement conditions on gene expression in renal cell carcinoma. *Clin.*  
278 *Cancer Res.* **19**, 42-49 (2013) doi:10.1158/1078-0432.ccr-12-2606  
279  
β80 130. Stewart, G., Powles, T., Van Neste, C., Meynert, A., O'Mahony, F., Laird, A., Deforce, D.,  
281 Van Nieuwerburgh, F., Trooskens, G., Van Criekinge, W., De Meyer, T., Harrison, D.J. Dynamic  
282 epigenetic changes to VHL occur with sunitinib in metastatic clear cell renal cancer.  
283 *Oncotarget* **7**, 25241–25250 (2016) doi:10.18632/oncotarget.8308  
284  
β85 141. De Meyer, T., Mampaey, E., Vlemmix, M., Denil, S., Trooskens, G., Renard, J.-P., De  
286 Keulenaer, S., Dehan, P., Menschaert, G., Van Criekinge, W., Quality evaluation of methyl  
287 binding domain based kits for enrichment DNA-methylation sequencing. *PLoS ONE* **8**, 59068  
288 (2013) doi:10.1371/journal.pone.0059068  
289  
β90 152. Biobix: Map of the Human Methylome. [http://www.biobix.be/map-of-the-human-](http://www.biobix.be/map-of-the-human-methylome/mhm-version-2/)  
291 [methylome/mhm-version-2/](http://www.biobix.be/map-of-the-human-methylome/mhm-version-2/)  
292  
β93 163. Hicks, S., Irizarry, R., Quantro: a data-driven approach to guide the choice of an  
294 appropriate normalization method. *Genome Biol.* **16**, 117 (2015) doi:10.1186/s13059-015-  
295 0679-0  
296  
β97 174. Law, C.W., Chen, Y., Shi, W., Smyth, G.K., Voom: precision weights unlock linear model  
298 analysis tools for RNA-seq read counts. *Genome Biol.* **15**, 29 (2014) doi:10.1186/gb-2014-15-  
299 2-r2  
300  
β01 185. Esteller, M., Epigenetic gene silencing in cancer: the DNA hypermethylome. *Hum. Mol.*  
302 *Genet.* **16**, 50-59 (2007) doi:10.1093/hmg/ddm018  
303  
β04 196. Volders, P.-J., Verheggen, K., Menschaert, G., Vandepoele, K., Martens, L.,  
305 Vandesompele, J., Mestdagh, P., An update on LNCipedia: a database for annotated human  
306 lncRNA sequences. *Nucleic Acids Res.* **43**, 174-180 (2014) doi:10.1093/nar/gku1060  
307  
β08 2017. D'Apice, M.R., Novelli, A., di Masi, A., Biancolella, M., Antoccia, A., Gullotta, F., Licata,  
309 N., Minella, D., Testa, B., Nardone, A.M., Palmieri, G., Calabrese, E., Biancone, L., Tanzarella,  
310 C., Frontali, M., Sanguuolo, F., Novelli, G., Pallone, F. Deletion of REXO1L1 locus in a patient  
311 with malabsorption syndrome, growth retardation, and dysmorphic features: a novel  
312 recognizable microdeletion syndrome? *BMC Med. Genet.* **16**, 20 (2015) doi:10.1186/s12881-  
313 015-0164-3  
314  
β15 218. Greijer, A., van der Groep, P., Kemming, D., Shvarts, A., Semenza, G., Meijer, G., van de  
316 Wiel, M., Belien, J., van Diest, P., van der Wall, E., Up-regulation of gene expression by hypoxia

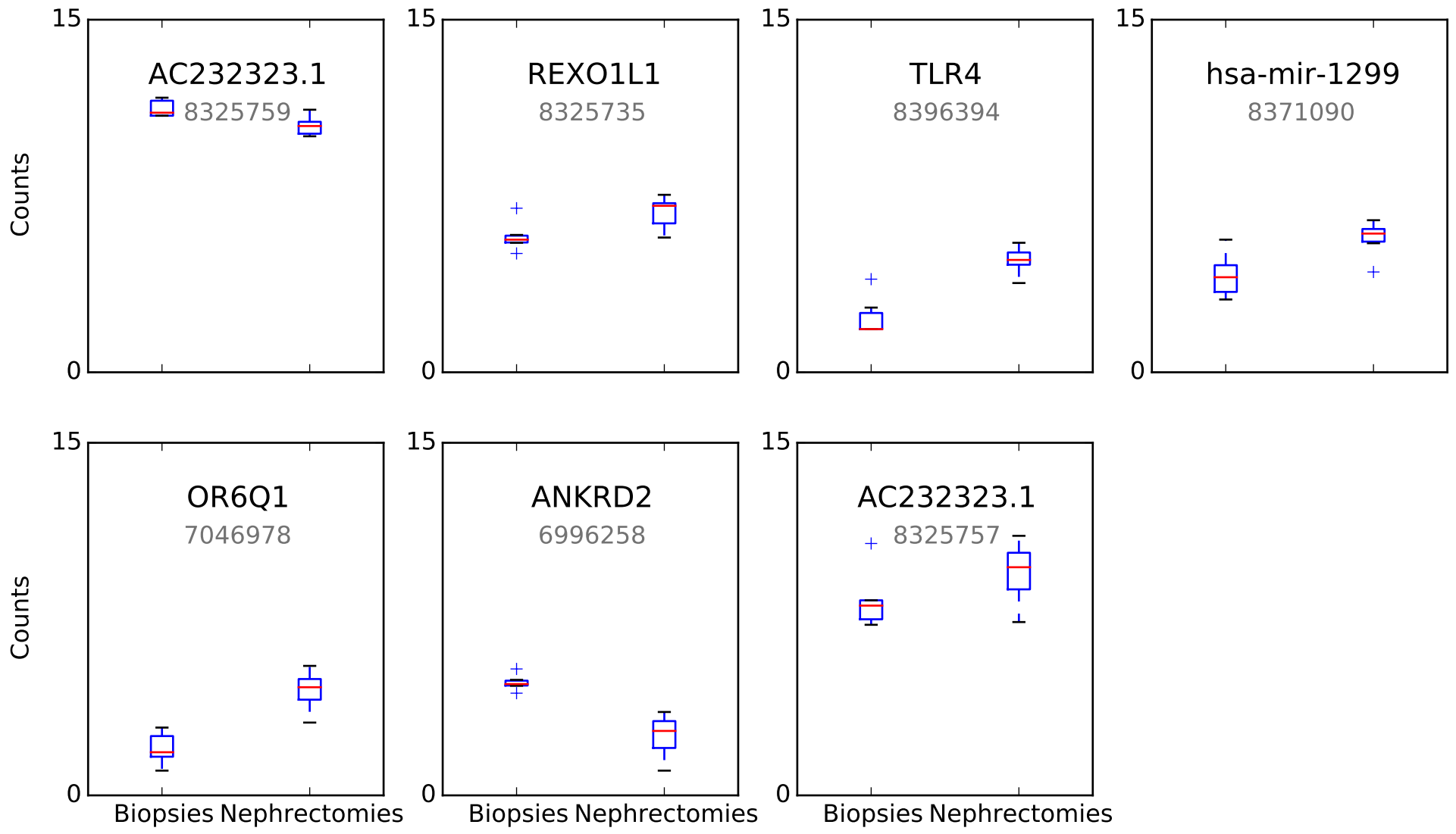
317 is mediated predominantly by hypoxia-inducible factor 1 (hif-1). *J. Pathol.* **206**, 291-304 (2005)  
318 doi:[10.1002/path.1778](https://doi.org/10.1002/path.1778)  
319  
β20 ~~2249~~. Wu, H., Chen, G., Wyburn, K.R., Yin, J., Bertolino, P., Eris, J.M., Alexander, S.I., Sharland,  
321 A.F., Chadban, S.J., Tlr4 activation mediates kidney ischemia/reperfusion injury. *J. Clin. Invest.*  
322 **117**, 2847-2859 (2007) doi:[10.1172/jci31008](https://doi.org/10.1172/jci31008)  
323  
β24 ~~230~~. Liu, Z., He, W., Gao, J., Luo, J., Huang, X., Gao, C., Computational prediction and  
325 experimental validation of a novel synthesized pan-pim inhibitor pi003 and its apoptosis-  
326 inducing mechanisms in cervical cancer. *Oncotarget* **6**, 8019-8035 (2015)  
327 doi:[10.18632/oncotarget.3139](https://doi.org/10.18632/oncotarget.3139)  
328  
β29 ~~244~~. Safran, M., Dalah, I., Alexander, J., Rosen, N., Stein, T.I., Shmoish, M., Nativ, N., Bahir, I.,  
330 Doniger, T., Krug, H., Sirota-Madi, A., Olender, T., Golan, Y., Stelzer, G., Harel, A., Lancet, D.,  
331 GeneCards version 3: the human gene integrator. *Database (Oxford)* **2010**, baq020 (2010)  
332 doi:[10.1093/database/baq020](https://doi.org/10.1093/database/baq020)  
333  
β34 ~~252~~. Miller, M.K., Bang, M.-L., Witt, C.C., Labeit, D., Trombitas, C., Watanabe, K., Granzier, H.,  
335 McElhinny, A.S., Gregorio, C.C., Labeit, S., The muscle ankyrin repeat proteins: Carp,  
336 ankrd2/arpp and darp as a family of titin filament-based stress response molecules. *J. Mol.*  
337 *Biol.* **333**, 951-964 (2003) doi:[10.1016/j.jmb.2003.09.012](https://doi.org/10.1016/j.jmb.2003.09.012)  
338  
β39 ~~263~~. Band, M., Joel, A., Avivi, A., The muscle ankyrin repeat proteins are hypoxia-sensitive: In  
340 vivo mrna expression in the hypoxia-tolerant blind subterranean mole rat, spalax ehrenbergi.  
341 *J. Mol. Evol.* **70**, 1-12 (2009) doi:[10.1007/s00239-009-9306-6](https://doi.org/10.1007/s00239-009-9306-6)

Table 1 Renal clamping differential results. One gene can have several methylation cores in the promotor region. 'AveExp' = average expression count. 'FDR' = Benjamini-Hochberg adjusted P-values, smaller than 0.05.

<b>METHYLATION CORE ID</b>	<b>ANNOTATION</b>	<b>LOG-FC</b>	<b>AVE-EXP</b>	<b>P-VALUE</b>	<b>FDR</b>
<b>8325759</b>	AC232323.1	0.731	10.84	1.256e-07	0.0055
<b>8325735</b>	REXO1L1	-1.184	6.28	1.842e-07	0.0055
<b>8396394</b>	TLR4	-2.483	3.55	8.634e-07	0.0173
<b>8371090</b>	hsa-mir-1299	-1.638	4.90	2.028e-06	0.0273
<b>7046978</b>	OR6Q1	-2.521	3.22	2.269e-06	0.0273
<b>6996258</b>	ANKRD2	2.249	3.66	3.262e-06	0.0327
<b>8325757</b>	AC232323.1	-1.242	8.88	4.777e-06	0.0411



figure 1



Supplementary table 1: Clinico-pathological characteristics of the two test patients.

Clinico-pathological Characteristic	Test patient 1	Test Patient 2
Age (years)	59	73
Gender	Male	Male
Fuhrman grade	2	2
T stage	3a	3a
M stage	1	1
Warm ischemia time* (mins)	55	60
Previous Treatment	3 cycles of Sunitinib 50mg once daily. Nephrectomy 4 days following cessation.	None

Warm ischemia time is the time from clamping of the renal artery to tumour sampling and snap freezing on dry ice.

Supplementary table 2: Clinico-pathological characteristics of the 14 validation cohort patients all with metastatic ccRCC and having tumour biopsy followed by sunitinib therapy and subsequent cytoreductive nephrectomy.

Age, median (range)	67 (52-78)
Male gender (%)	11 (78.6)
MSKCC prognostic risk (%)	
Intermediate	11 (78.6)
Poor	3 (21.4)
Metastatic sites (%)	
1-2	10 (71.4)
3+	4 (28.6)
Clear cell tumour grade, identified at nephrectomy (%)*	
1-2	7 (50)
3-4	7 (50)
Median PFS, months (95% CI)	17.5 (7-21)